

IDENTIFICATION PROGRAMME FOR POISONOUS
AND HALLUCINOGENIC MUSHROOMS
OF INTEREST TO FORENSIC SCIENCE.

A THESIS

presented by

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BEWARE OF MUSERONS, MOCH PURSLANE,
GOURDES, AND AL OTHER THINGES,
WHICHE WYLL SONE FUTRIFIE.

Thomas Elyot

The Castel of Helth (1541)



PSILOCYBE SEMILANCEATA

Psilocybe semilanceata collected

by the author.

Original watercolour

by R. Furlong.

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SUMMARY.

An efficient differential diagnosis of mushroom poisoning is essential for a successful treatment of this type of intoxications.

An integrated multidisciplinary approach to the identification of poisonous and hallucinogenic mushrooms by a completely interactive, - on-line - identification programme is proposed.

Twenty four characters showing 127 possible states have been chosen to define each taxon included in the data bank. These characters, microscopical, ecological, medical and chemical are described and discussed.

Essential information is retrieved with each taxon identified, as well as a complete up to date bibliographic record.

223 poisonous or suspect species of mushrooms are included in the data bank which contains a total of over 30'000 data. A reference collection of 223 Scottish species of suspect mushrooms has been built and used to check, complement or expand the microscopical, ecological and chemical data.

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CHAPTER I : INTRODUCTION

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1.1. Mycotoxicology of the Higher Fungi

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I. INTRODUCTION.

With the advent of modern scientific and technological developments, areas once confined to religion, folklore, art and even gastronomy have become the concern of science. The study of higher fungi and their toxicology are two fields which have been neglected until recent years. Only in the last two decades has substantial systematic research warranted the application of highly sophisticated equipment in the evaluation, identification and diagnosis of mushroom poisoning. However, this scientific approach in no way denigrates the value of historical records obtained from folk and medical sources, it merely renews and increases the potential for pure research by suggesting new horizons. Information acquired from such sources should be thoroughly tested and stored as a basis for a completely integrated approach to the study of the mycotoxicology of the higher fungi.

The present thesis is a modest contribution to what will hopefully be a major field of research and development. It has been helped by recent extensive contributions which, although they do not introduce new concepts, are a wealth of recorded historical, medical and scientific information (Arietti and Tomasi, 1969/1975; Flammer, 1980; G rault, 1976; Heim, 1963/1978; Lincoff and Mitchel,

1977; Rumack and Salzman, 1978). The success of these reports may be attributed to the fact that the information is concentrated in monographic form, rather than diluted in chapters included in more general works. However, they also fail because the subjects are presented in a way biased towards the individual authors' speciality, together with all its specialised jargon. This makes diagnostic keys difficult to follow and identification becomes a difficult task to the practising scientist, whether he is a physician, toxicologist, forensic scientist, mycologist or a naturalist.

Information sciences and electronics have given the scientific community, in the form of computers and micro-processors, a very potent tool for data storage, fast retrieval and up-dating of information. These new concepts have to be applied if one wants to give a wider sector of the scientific community access to the available up to date information. For a meaningful use of these facilities, facts have to be consistent and complete with respect to the characters used for information retrieval.

An efficient and integrated approach to the identification of poisonous higher fungi is proposed, taking full advantage of modern computer services.

1.1 Mycotoxicology of the Higher Fungi.

Although mushrooms have been eaten since well before recorded history, and the toxicity of some of them has been known from as far back as Greek and Roman times, the methodic and scientific study of mycotoxicology only started at the end of the eighteenth century.

Following a study of the toxicity of certain species of mushrooms on animals, Paulet (1793) described a number of toxic species, gave warnings on how to prevent accidents and also suggested possible means of treating poison-cases. He was soon followed in the nineteenth century by a whole generation of scientists, such as Elias Fries (1794-1878) in Uppsala, who established the basis of modern taxonomy for the higher fungi, Mathieu J. B. Orfila (1787 - 1853) who in 1813, at the age of 26, pioneered the era of a new science, toxicology, in his "Traité des poisons". Finally, chemists who, with Schmiedeberg and Koppe (1869), isolated muscarine thus opening the door to a century of research on Amanita muscaria which culminated in 1954 with the synthesis of muscarine by Eugster and Waser.

In the 1920's and thirties, the number of general works, treaties and theses on poisonous mushrooms flourished. This went some way towards establishing the

reliability of folklore data and still provides a major source of information on toxic fungi (Sartory and Maire, 1921; Martin-Sans, 1929 and Henry, 1931; to name but a few). With the exception perhaps of Heim's general work in 1963, interest in mycotoxicology seemed to wane, and only specific aspects of research, mainly by three major groups headed by the Wielands in Heidelberg, Eugster in Zürich and Hofmann in Basle, continued (Lynen and Wieland U., 1937; Wieland H. and Hallermayer, 1941; Wieland T. and Wieland O., 1972; Wieland T. and Faulstich, 1978; Waser and Eugster, 1954; Müller and Eugster, 1965; Good et al., 1965; Catalfomo and Eugster, 1970; Hofmann et al., 1958; Hofmann et al., 1959).

Following advancements in medicine, fungal biochemistry, modern taxonomy and toxicology accompanied by a parallel development of a general interest in wild mushrooms as food or, in some cases, as a drug used for recreational purposes, a number of textbooks have appeared. Of particular interest are the publications of Arietti and Tomasi (1969/1975), Lincoff and Mitchel (1977), Heim (1963/1978), Rumack and Salzman (1978) and Flammer (1980). Theses by Gérard (1976) on the higher fungi and their intoxications and by Bornet (1980) on intoxications due to mushrooms other than Amanita phalloides are also of importance. These books represent today's

state of the art. Lincoff and Mitchel (1977) and Flammer (1980) with the help of mycologically or medically orientated identification methods set out an efficient and logical, albeit traditional, approach to the management of emergency toxicology in the case of mushroom poisoning. One major drawback is that this scheme of diagnosis and treatment requires a broad specialist knowledge of mycology/anatomy, botany, chemistry and/or medicine. Such broad training would only be feasible if the majority of toxicological emergencies arose from this particular type of poisoning, which is not the case. In fact, it would be bad management to induce laboratory directors, in charge of toxicological centres, to divert their experts' normal priorities towards such a marginal problem.

Statistics on mushroom poisoning are scarce, scattered and most of the time not reliable. To illustrate the argument one need only look at enquiries made to the British National Poisons Information Centres (Moffat, 1980). In 1977, 32'892 enquiries were processed, of which only about 10 % concerned plants and insect bites. Certainly less than 10 % of those concerned mushrooms with for instance, only 16 such enquiries for Scotland (Margot, 1977). Most of these 16 reports concerned small children who ate unidentified mushrooms and were sent to hospital by worried parents, rather than because poisoning

symptoms developed.

Over a thirty year period (1920 - 1950), Ramsbottom (1953) reported 39 fatalities due to mushroom poisoning in England and Wales. This number is in marked contrast to the total number of deaths caused by poisoning in England and Wales in a single year (1975). These latter 2984 deaths mainly involved manufactured drugs (Moffat, 1980).

A more worrying example is highlighted in a recent thesis by Borner (1980). In Switzerland, where there is a control organisation of mushroom collections and a fairly extensive network of specialists trained in the identification of mushrooms (VAPKO : Vereinigung der amtlichen Pilzkontrollorgane), there were only 806 calls to the "Centre Suisse d'information toxicologique" in Zürich relating to mushroom intoxications between 1966 and 1978 out of a total 100'554. Of these 806 calls, 134 concerned poisoning with Amanita phalloides and related species, 87 suspected mushroom poisonings were found to be unconnected with fungi and 342 cases concerned unidentified species. Of the remaining 243 calls, mushroom identification was conclusive in all but 15. In one case of intoxication involving a whole family, the parents and child were sent to different hospitals and the child subsequently died. The parents were found

to have been intoxicated with Amanita phalloides (Death Cap), whereas the death of the child was attributed to Armillaria mellea (Honey Fungus), a perfectly edible mushroom under usual circumstances. Bornet painted a bleak picture about the value of the identification of mushrooms involved in past poisoning cases and their use in statistics. He judged that only approximately one third of the identifications were acceptable, although some of his criteria for acceptance he considered, in his own words, "parfois subjectifs et discutables" *.

These reports are sufficient to highlight the artificiality of such well thought-out books and leave no alternative but to have at hand a number of specialists (i.e. a mycologist, an analytical chemist, a toxicologist, a kidney or liver specialist, etc) who can be contacted in an emergency to help the practitioner.

Whilst such an arrangement might lead to a correct identification of the problem, it might also introduce an unacceptable and perhaps fatal time delay before diagnosis is complete and treatment instituted.

The traditional approach to the identification of mushrooms usually incorporates the use of diagnostic keys, that is the identification is obtained after a set of questions

* "sometimes subjective and debatable"

has been answered. These are in a sequential order, and at each step two (dichotomous keys) or more (polytomous keys) answers are possible. Each answer, in turn, leads to a new question which has two or more answers, etc until a species is diagnosed. The alternative parts of questions in keys are called leads because they lead from one question to another. Each fundamental item of information in a key is called a character. Characters, when observed, are seen to show various states (or values or attributes). For example "colour" is a character and "red", "blue", "yellow", etc are each one state of this character. In an hypothetical key an answer of "red" to the question "What is the colour of the cap?" will lead after x steps to the identification of Amanita muscaria, whereas "green" would lead to Amanita phalloides after y steps.

Should one of the characters be missing near the start of a key, an identification becomes impossible and the problem remains in its entirety.

Another problem arises from possible errors made during the use of the keys by identifying characters not clearly defined in a given sample. If the error occurs near the beginning of the key, the consequences may be critical: the wrong lead will be followed and the user will continue in the wrong part of the key. If the above

example is used, the colour of a dry or old cap might be identified as brown, instead of green, thereby leading the user to a totally wrong and potentially dangerous conclusion. The fact that an answer is necessary at each step of the key is a serious drawback because samples provided in emergency toxicology are rarely complete and in good condition. Therefore every chance exists that the keys either cannot be used, or worse, can lead to the wrong answer with all the implications that follow.

On the other hand, with the increased interest in natural food resources and also in hallucinogens by a marginal sector of the population, it is clear that cases of accidental intoxication will increase, as has been demonstrated lately (Watling, 1980; Short et al., 1980). Detailed information is therefore required on the identification of the mushrooms concerned, whether they are submitted for analysis as large and fresh samples or, more likely, as old fragments such as from stomach washings or meal remnants. With respect to the latter situation, the identification of the mushroom and any information on the pharmacokinetics of any toxins present would be of great help to the physician and clinical chemist in deciding the prognosis of the patient.

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II. REQUIREMENTS FOR AN IDENTIFICATION PROGRAMME.

From the above introduction, it is clear that a mass of information exists which, if made accessible to the clinician, will lead to a proper management of mushroom poisoning cases. However, because the problem is marginal in the context of the total number of poisoning cases, it cannot be expected, at present, for this information to be made available as quickly and efficiently as it should be. As a first step towards a successful treatment, it is necessary to establish an efficient mushroom identification method initially using existing data, but capable of expansion to include knowledge of toxic fungi accumulated by studying certain limited aspects of the chemistry, pharmacology and toxicity of certain species or genera.

2.1 Requirements for an Efficient Identification Method.

A list of suspect and known poisonous mushroom species should be recorded based upon information derived from historical and medical sources as well as the state of knowledge of each species from the point of view of their taxonomy and chemistry. A number of fundamental items of information should then be selected

from the data matrix : these are the diagnostic characters. They should fulfil certain criteria so as to offer a high degree of usefulness for the purpose of identification. These criteria are governed mainly by their ease of observation and the information content of the character used. This latter concept is not absolute and has to be weighed in relation to the objects to be identified or taxa under study. (sing. "taxon" = taxonomic group of any rank. Every individual fungus is treated as belonging to a number of taxa of consecutively subordinate ranks, among which the rank of species is basal (Hawksworth, 1974)).

These characters and their states, for obvious reasons, should show as little variation within taxa as possible and the states should be mutually exclusive. It is also essential that any selection and sequence of characters can be used in the identification process, because the material under investigation may be incomplete or fragmentary. Furthermore, the user might have a limited knowledge of only certain types of characters used in the key, or might even want to limit himself to the use of these characters.

If the material, or the user, cannot generate enough characteristic information to reduce the identity

of a specimen to a single taxon, a short list of taxa should be obtained which might just provide the information required in an emergency. Keys where any selection of characters may be used, and in any order, are called multi-access keys (Pankhurst, 1978). This is the first requirement for an efficient identification system independent from the data matrix itself.

Another requirement is that the identification should not necessarily depend on the correct observation of all the characters; that is, some errors should be tolerated. Some extremely useful characters might be difficult to observe or untypical of a given taxon under some extreme circumstances and might be genuinely mistaken. Rather than have no identification due to a mistaken observation, the computer can be told what number and which characters are permissible as disagreements. This is a precaution against the effect of erroneous characters in the specimen description called variability limit by Morse (in Pankhurst, 1978). That means that the data observed are compared with all the sets of data of the taxa included in the data bank and identified with whichever of these agrees exactly, or is the most similar when no agreement is found.

Conventional multi-access keys produced on punched cards fit these requirements and have been used in a

number of fields. However, they suffer some major drawbacks which rule them out of the present project: they are not easy to publish, they are expensive, bulky and fragile. Furthermore, they are slow to use and not easy to update.

These drawbacks illustrate further requirements for an efficient system of identification in the case of mushroom poisoning. The system should be fast, that is, the time scale should be in terms of seconds rather than minutes or even hours as is common with traditional systems. It should also be readily updated and corrected without involving major alterations to the whole system. The cost/benefit ratio should be low, so that the system can have a wide application. Finally, and above all, such an identification should be reliable and accurate as far as the matching or identification process is concerned. For this reason, the data should be carefully and thoroughly tested and checked so that, provided the observed data is accurate, there is a high degree of certainty that the results are strictly accurate.

In short, the system should not be intelligent, but capable of making complex decisions and matchings in a matter of seconds once all the necessary data has been provided. Computers are ideal tools for such an identification scheme once the factual data is complete and

consistent, in that they can produce accurate and fast identifications. Computers are available to most hospitals, universities, toxicological centres and forensic science laboratories and this justifies their use in such fields as mycotoxicology.

In order to have an identification system as fool-proof as is humanly possible, a number of requirements should be further included into the system, bearing in mind computer capabilities and programme logic.

The main and first requirement is that it should be easy for the user to handle. He should not have to code the information collected but should be able to use his own natural language to communicate with or command the machine. The user should also be free to use and choose any character at random included in the programme and finally to select how many and which characters are permissible as disagreements if no perfect match is found.

These facilities are offered by a completely interactive or on-line identification programme. The user has a computer terminal, in the case of a large time-sharing computer system, or a desk-top microcomputer with a keyboard for typing instructions, and a printer or a visual display unit (VDU) for obtaining the answers in the form

of a dialog.

2.1.1. Summary.

The requirements for an efficient management of mushroom poisoning through the identification of the causative agent cover two basic aspects: the data matrix and the key system.

The data matrix is a series of descriptions of the taxa. The descriptions have to be complete and accurate if an automatic system is to be used. From that, a number of characters, which are easy to observe and/or have a high information content, have to be selected to be used in the identification process.

The key system must provide a multi-access facility, be fast and easy to update. It should also be reliable, accurate, have a low cost/benefit ratio and a language understandable to the user. Finally it should be capable of accommodating a limited amount of incorrect information in such a way that the whole system does not fail.

An on-line identification programme is offered here as a tool fulfilling all these requirements.

2.2. The Data Matrix.

Before any computing, or before data can be coded in a format or in a way which is useful for identification, the data must be collected. It may be obtained by a literature or field research, or by analytical means. The total data can be conveniently divided into two partially overlapping categories: that of the data retrieval and the diagnostic data.

2.2.1. Data Retrieval.

Part of the data matrix, the data retrieval, should consist of information one would wish to retrieve in an emergency. Experience and good sense has shown that the following information is required in most poisoning cases:

- identity of the poisonous species
- its frequency of occurrence
- the toxin involved
- the favoured treatment
- bibliographic information
- the relative toxicity
- the geographical distribution.

2.2.1.1. Identity of the Poisonous Species.

As in most taxonomical systems, there is no universal agreement on the identity of families, genera and species. Different taxonomists use different names for identical species. For example, the genus Pholiota is included in the family Cortinariaceae by British authors (Dennis et al. 1960) whereas it is included in the family Strophariaceae by their American (Singer, 1975) or German-speaking counterparts (Moser, 1978); the genus Hypholoma (Moser, 1978; Dennis et al. 1960) is called Naematoloma by American authors (Singer, 1975). Similarly, the species Psilocybe wassonii described by Heim (1957) is called Psilocybe muliercula by Singer and Smith (1958), and so on. These intricate inconsistencies warrant the use of a taxonomical reference, which is used throughout the present work unless otherwise stated. That reference is the "Check List of British Agarics and Boleti" (Dennis et al. 1960) and its subsequent amendments and complements (Orton, 1964; 1969; 1972; 1976 a; 1976 b; Watling, 1967; 1970; 1972; Henderson et al. 1969; Orton & Watling, 1979; Rayner, 1974).

2.2.1.2. Frequency of Occurrence

It may be important to know just how common a given species may be in order to assess if that species is likely to be involved in a large scale poisoning such as described by Grzymala (1957) for Cortinarius orellanus which occurred in Poland in 1952. From such information it should also be possible to assess to a certain extent the danger to public health and safety. The frequency of occurrence of mushrooms is a very difficult factor to determine due not only to the lack of extensive records but also because of variability associated with differing weather conditions, geographical location and other ecological factors. With all its limitations, a frequency of occurrence in Great Britain is included in the data matrix as a guide or an estimate. It arises from information gathered by the British Mycological Society (Rayner, 1980) during the forays of autumn 1958 to spring 1976. Frequencies are expressed as the number of localities at which the species was found, determined as a percentage of the total number visited, i.e. 253 locations. It provides a fairly good and, at least, objective estimate of the relative frequency of species included in the British Check List (Dennis et al. 1960). Despite its limitations, this is likely to be preferable to the rather subjective evaluations

to be found in traditional fungal floras. In such a survey it is natural that easily identified, or widely known species will appear to be more frequent than they are, whereas less easily identified species may appear to be rarer. Furthermore, since the forays take place each year in May and September, the frequency of species that fructify mainly during these two months will be overestimated. This frequency value may also be affected by how the sporophore is distributed, that is, it may grow in high densities in a limited number of areas and show a lower frequency (i.e. less important toxicologically) than another mushroom which is widespread throughout the country, but producing only a small amount of sporophores. It should be clear here that the reverse is true, i.e. that a high density species is more likely to be involved in a case of mass poisoning such as experienced in Poland in 1952 (Grzymala, 1957), although the frequency value may tend to indicate otherwise. Mycologists, throughout Britain, are recording data, which, as time goes on, may prove more accurate and will replace the present values where adjustments prove necessary.

2.2.1.3. Toxins.

The identity of the toxins is to be found in both the data retrieval and the diagnostic data. It is given here for completeness and to help the non-chemist. This repetition is necessary because the key is intended for specialists in widely different fields. The taxonomist, or clinician identifying a poisonous mushroom will also want to know the identity of any toxin associated with that particular mushroom. If the toxin proves to be unknown, the specimen should be forwarded to an analyst/chemist/toxicologist in order that the chemical entity responsible for intoxication can be identified and its toxicity established. This information can eventually be used to extend the data system.

2.2.1.4. Favoured Treatment.

For practical purposes, the fourth data to be retrieved is a short outline of the treatment generally accepted as best at the time of presentation of this thesis.

In some cases, where the toxin is well known, such as muscarine, the treatment is simple and an antidote to muscarine can be administered. In other cases, the

toxin is either unknown or treatment for a particular poison has not been firmly established, usually because of a lack of knowledge of what happens in the body when such a toxin is ingested. Extensive research is still necessary if the efficient management of all mushroom intoxications is to be achieved.

2.2.1.5. Bibliographic Information.

While the present programme has some tolerance for errors, the results have still to be checked by comparison with full descriptions, illustrations, or, better even, with preserved or voucher specimens. Therefore three bibliographic references are included which allow the user to find complementary information.

The three texts selected are reviews, or the most complete work on a given aspect of a certain species. When modern relevant information is found, preferably in English and of easy access (i.e. in widely available publications), it is selected and preferred to original work published in different languages and in sometimes obscure journals.

The first reference gives a complete taxonomical description of the species, that is to say it includes

macroscopical features, smell, taste, microscopical features, ecological data and, in most recent publications, herbarium references. Often a comment describes and discusses differences between closely related taxa.

The second reference deals with the isolation and identification of the toxin(s) where known, or gives a survey of what is known about the chemistry of the given species under investigation. It does not necessarily relate to the species itself, but to the toxin known or suspected to be present in the species.

Lastly, the third reference gives detailed information on the symptoms of the intoxication and its treatment or proposed treatment. As before, the text relates to the toxin(s) and not necessarily to the species under investigation.

2.2.1.6. Relative Toxicity.

An assessment of the relative toxicity of described species is given. This assessment is purely informative and cannot be measured in any real terms. It is based on medical records which indicate that some mushrooms provoke often fatal intoxications, as is the case of Amanita phalloides, estimated to be at the source of

75 to 90 % of all deaths due to mushroom poisoning. Some are dangerous, but rarely fatal such as Entoloma sinuatum, some rarely dangerous such as Agaricus xanthodermus (no death reported, this species is eaten without ill-effects by many), or even only upsetting under certain circumstances, such as Coprinus atramentarius (conditional intoxication in the presence of alcohol). How to distinguish the degree and severity of intoxication is not clear and is based on historical repute. Until proper statistics exist on the concentration of the different toxins in the fungi, the frequency of occurrence in a given area and the reliable identification of all mushrooms involved in poisoning cases, this relative estimation of the toxicity will remain an educated guesswork (i.e. very subjective).

2.2.1.7. Geographical Distribution.

The distribution of European species is well documented and it is even possible to distinguish North or South European flora or mountain flora. This is true too for North American species, but otherwise records are scattered, sometimes inaccurate and in any case incomplete. The indication of a geographical distribution in this thesis is as known at the time of filing of the data, and should be regularly up-dated.

2.2.1.8. Discussion and Example.

The final format of the retrieval data has been modified and completed after discussion and consultation with interested sectors of the scientific community. Extensive testing has shown that all the necessary data is retrieved and that it provides an efficient service for use in an emergency situation providing an accurate diagnosis of the source of the intoxication is established.

Figure 1 illustrates the form in which the data is retrieved. It is all condensed in a maximum of six lines of text. The species is identified by its generic name Psilocybe, specific name semilanceata and the taxonomic authors who described and named the species (Fr.)Kum. (Fr. = Fries who described first Agaricus semilanceatus; name altered to Psilocybe semilanceata by Kum. = Kummer; who elevated Psilocybe to the rank of a genus). It is followed by six lines of text giving all the necessary information to deal with an emergency case involving this mushroom.

Figure 1: Data retrieval's presentation: an example.

```
POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
PSILOCYBE SEMILANCEATA (FR)KUM UK FRES 5.9 PSILOCYBIN/INDOL HALL.
PROPOSED TREATMENT PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT
BEHAV, QUIET POSSIB DIAZEPAM. CHLORPPOMAZINE TAXON VON MICHAELIS (1977)
Z PILZK 43, 305-310 CHEM HOFMANN ET AL (1959)HELY CHIM ACTA 42, 1557-1572
MED LINCOFF & MITCHEL (1977)TOX & HALLUC. MUSHROOM POISONING, VAN NOST-
RAND R, NY HALLUCINOGEN, UPSETTING EU; N. AM.
```

2.2.2. Diagnostic Data.

The other set of data, a prerequisite for an identification programme, is a data bank consisting of a compilation of diagnostic characters and their respective states found in different species. This thesis describes a wide range of characters identified from different scientific sources, chemical, morphological, pharmacological, geographical, historical and ecological.

The choice of diagnostic characters is of prime importance since the whole identification process relies solely on the knowledge of these characters and their corresponding states. A number of factors have to be taken into account, before a character can be chosen as part of a programme. It has to answer a number of general criteria.

The first criterion is that the character should be stable within certain limits and its states should not be alterable from one to another. For example, if one takes the character "colour of the cap", in the case of Russula, the dyes giving the cap colour are water soluble and the state shown may be "red" in dry weather conditions or "white" after a shower! i.e. this character is not stable. All characters which change quickly in a mushroom, such

as colour, texture, stickiness, shape, smell and flavour and, for that matter most macroscopical data should be excluded from an identification system because in most emergency poisoning cases the sample is fragmentary, half digested, cooked and/or decomposed since they are often collected from litter bins or as left-overs after a meal.

A second criterion is the ease of observation of a character. An easily observed and stable character is likely to be important in a diagnostic key because it can be used by relatively unexperienced personnel.

A third and important criterion is the information content of a given character, i.e. a character that splits species quantitatively in roughly equivalent groups is a good character with a high information content whereas characters dividing species in grossly unequal groups might be highly diagnostic for a particular group of species, but of little use for distinguishing species in general.

Other useful criteria are the independence of characters and the discreteness of the states of the different characters. For example, the cause and the effects are dependent characters. In the case of mushroom poisoning, the analysis of the toxins and the observation of the symptoms define two separate but dependent characters.

Used concurrently, they do not further the identification and the use of only one character is required. Similarly, continuous characters are not useful in the identification process because they cannot be defined within precise limits and overlap between states is almost inevitable. Only when the states show widely different values can they be of some help to the identification.

All these criteria are rarely united for a given character and a balance between good and bad features should be made before it is included in the key. The ease of observation probably outweighs all the other criteria when the users of a key are from widely different academic disciplines, and by no means specialists in the field of mycotoxicology.

Unfortunately, the information content of easily observable characters is not always very good and, therefore, highly informative characters also have to be included to allow for species identification when necessary, rather than identification of a group of closely related species. The discriminative power of a character is not an absolute concept and has only meaning in relation to the taxa which are taken into consideration in the data bank.

Each discipline included under the all embracing term of mycotoxicology has its own specialised characters which should be represented in the key, thus allowing a reasonable diagnosis based on the one speciality with perhaps only one or two easily identified characters pertaining to other disciplines.

Because of all these reasons, the measure of the information content or of the separation coefficient for each character becomes meaningless and will even vary considerably depending on the qualifications or ability of the user of the key. The choice of each character included is discussed in its own right and in relation to other characters as seen by the author. An objective weighting being impossible, the choice of characters has therefore been subjective and based on personal experience. The usefulness of certain characters (microscopical) is demonstrated by their extensive use in traditional keys.

2.2.2.1. Character Choice.

For practical reasons, the characters are grouped by user's specialities rather than in the order they are used in the programme. Each character's name is followed by a number in brackets indicating its position

in the key. For example "Toxins" (14) indicates that the character "toxins" is the character number 14 in the list used by the computer.

Four separate groups of characters are included:

- 1) microscopical characters (title 2.2.2.1.1.)
- 2) ecological characters (title 2.2.2.1.2.)
- 3) medical characters (title 2.2.2.1.3.)
- 4) chemical characters (title 2.2.2.1.4.)

2.2.2.1.1. Microscopical Characters.

2.2.2.1.1.1. Introduction.

These characters are traditionally used by modern taxonomists and follow the definitions laid down in the "Introduction to the Agarics and Boleti" of the British Fungus Flora (Henderson et al. 1969) and further detailed by Largent et al. (1977). Being the traditional means of identification of mushrooms, these characters are indeed sufficient to permit the identification of the taxa included in the key.

For practical reasons, states of characters which are numerous, or continuous have been categorised in groups of related states which are still found to carry

a useful information content, in order to allow a useful split within the included taxa.

With the publication of the first observation of fungal spores in 1588 by Giambattista della Porta (Ainsworth, 1976), one of the most useful elements for mushroom identification was discovered.

Eight diagnostic characters based on the morphology of the spores are used here, whereas only five, three and one are based on the hymenium structures, the hyphal trama and the cuticle respectively. (see Figure 2).

These characters are almost always present, even in the smallest fragmentary material, and some keep their diagnostic value even after they have been through extreme conditions (e.g. spores are still found in perfect condition in gastric lavage of poisoned people exhibiting a rapid onset of symptoms).

Prior to describing each character and its location in detail, it is assumed that the reader/user of the key has some experience of basic microscopy. An ordinary student's microscope with magnification capabilities of 30 to 300 x is perfectly adequate for most, if not all, the microscopical examinations.

Whenever a reagent is mentioned for a special

purpose (e.g. staining), its composition is referred to and described in the appendix (title 6.1)

2.2.2.1.1.2. Anatomic characters.

Each character used for identification has a special purpose and a specific position in the anatomy of a carpophore. A number of sections can be made, and are illustrated in Figure 2. All the sections are made on the pileus and three sections show all the microscopical features of interest:

A: section 1: surface of the pileus

B: section 2: transverse section of the gills

C: section 3: part of a gill

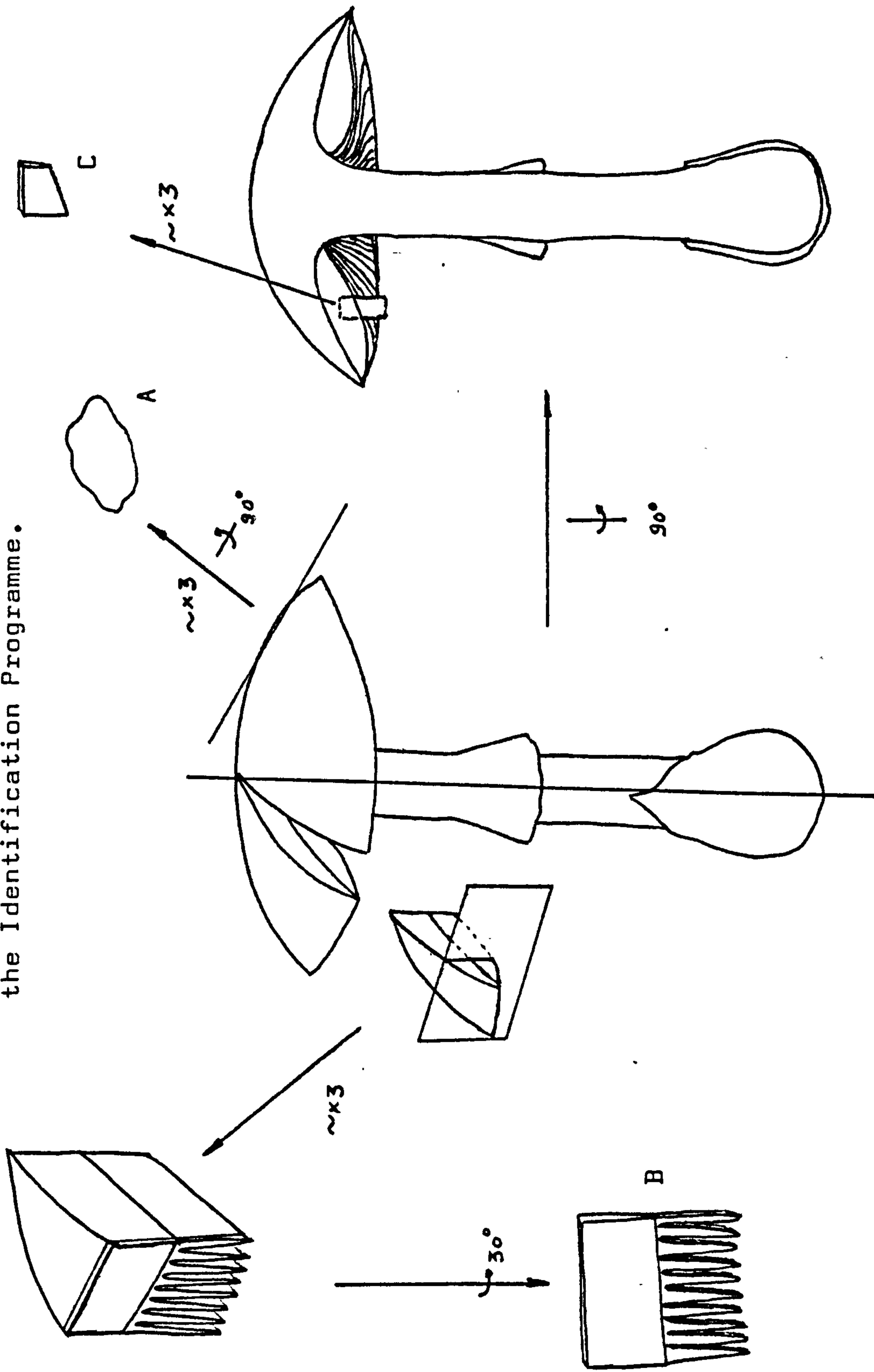
The position of each character used here is always located in relation to the above sections in the subsequent chapters.

2.2.2.1.1.3. Cuticle Structure. (18)

States: - cellular
- filamentous
- absent

A section of the surface of the pileus (section 1), which can be described as a scalp shows the general

Figure 2: Dissection of a Carpophore of a Mushroom to Locate the Characters Used in the Identification Programme.



A: section 1: surface of the pileus B: section 2: transverse section of the gills
C: section 3: part of a gill

configuration of the outer surface of the pileus called the cuticle or pileipellis. This configuration can be one of two types in the Basidiomycetes: either cellular or filamentous. These states are illustrated in Figure 3.

This is one of four characters used by modern taxonomists to split the Basidiomycetes into families and, as such, is an important diagnostic character (See Table 1). This feature is totally absent from the carpophores of the Ascomycetes which have their hymenial layer on their external surface.

This feature is unlikely to be found in gastric lavage, but can, for instance, still be observed in remnants of meals.

2.2.2.1.1.4. Transverse Section of the Gills.

This section (section 2; Fig.2) gives the most rewarding results because all but three microscopical features can be studied on this section.

2.2.2.1.1.5. Pileus Trama (16)

States : - homoiomerous
- heteromerous

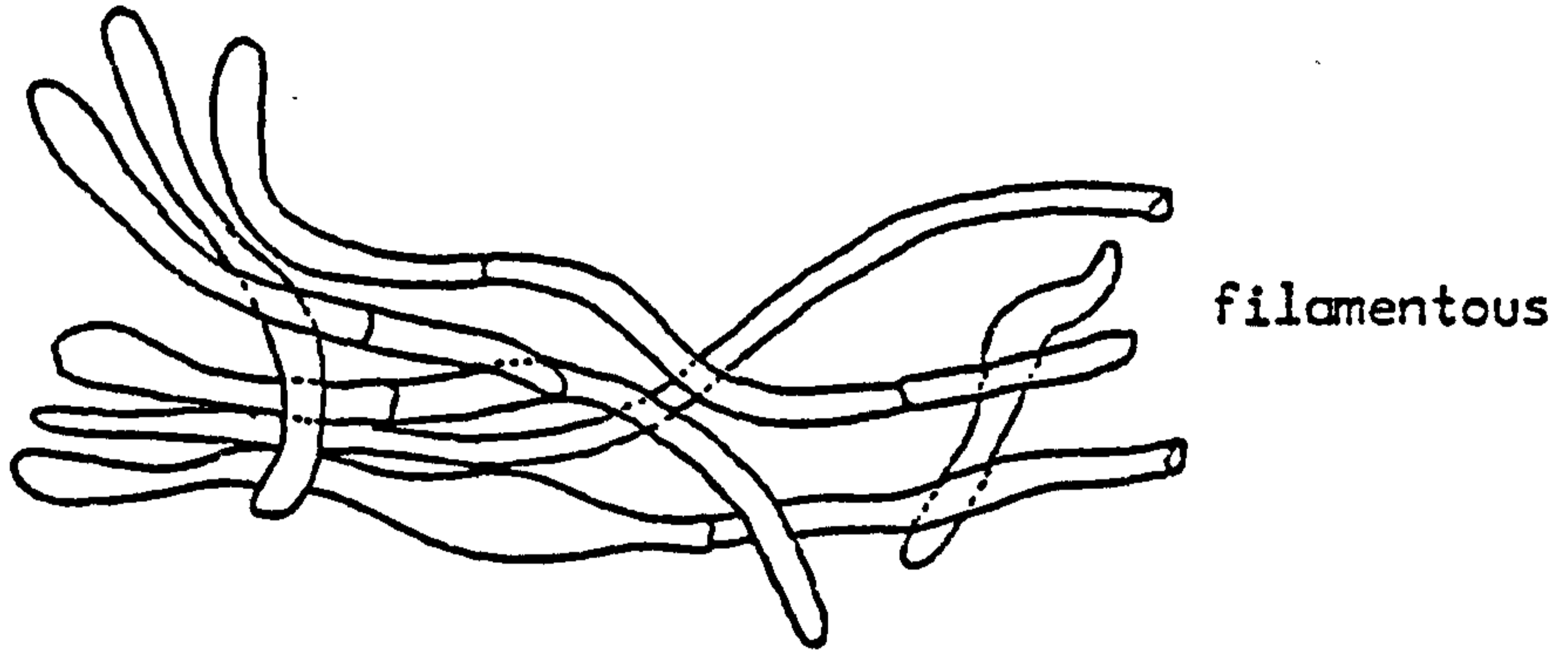
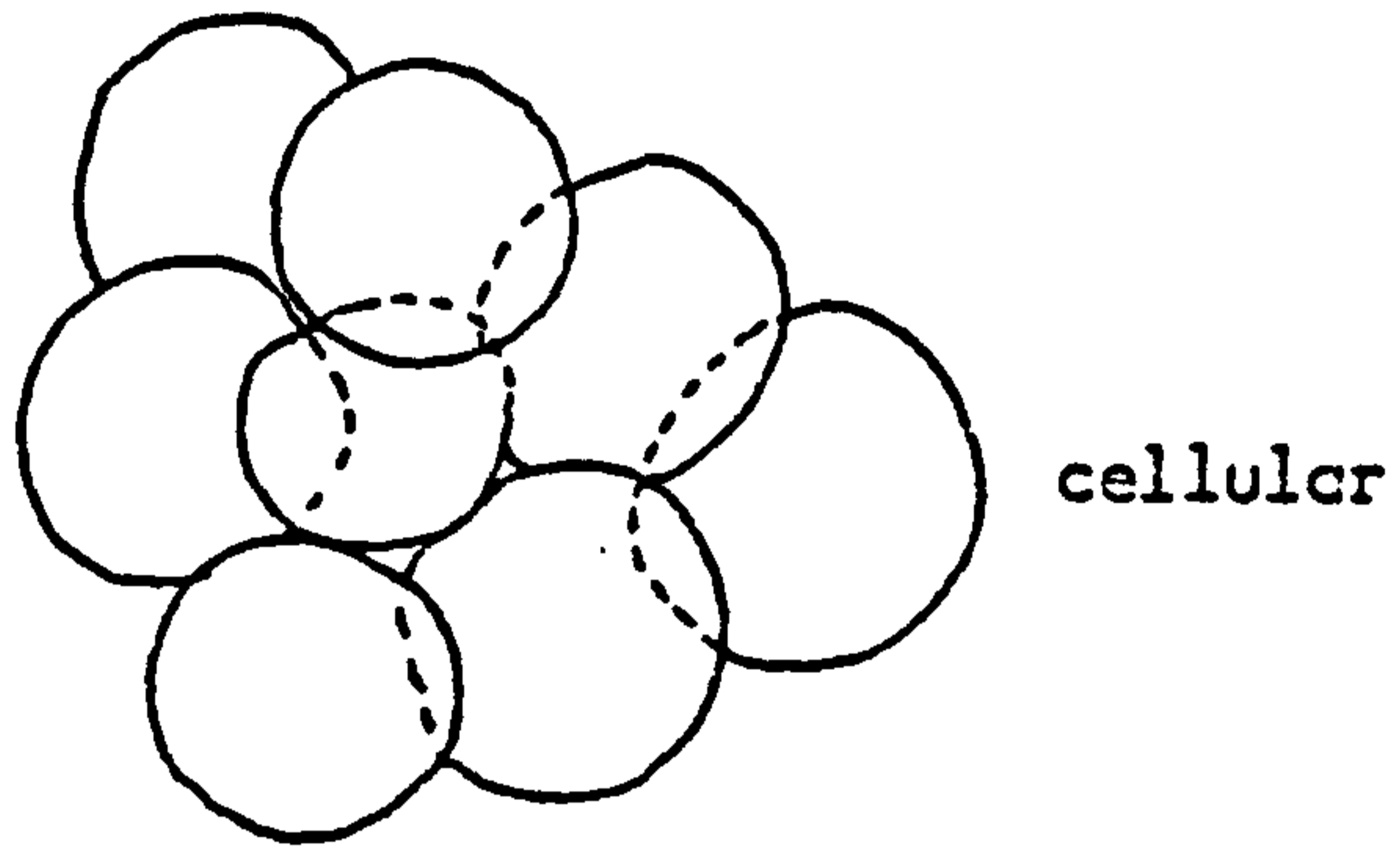


Figure 3 : Cuticle Structure

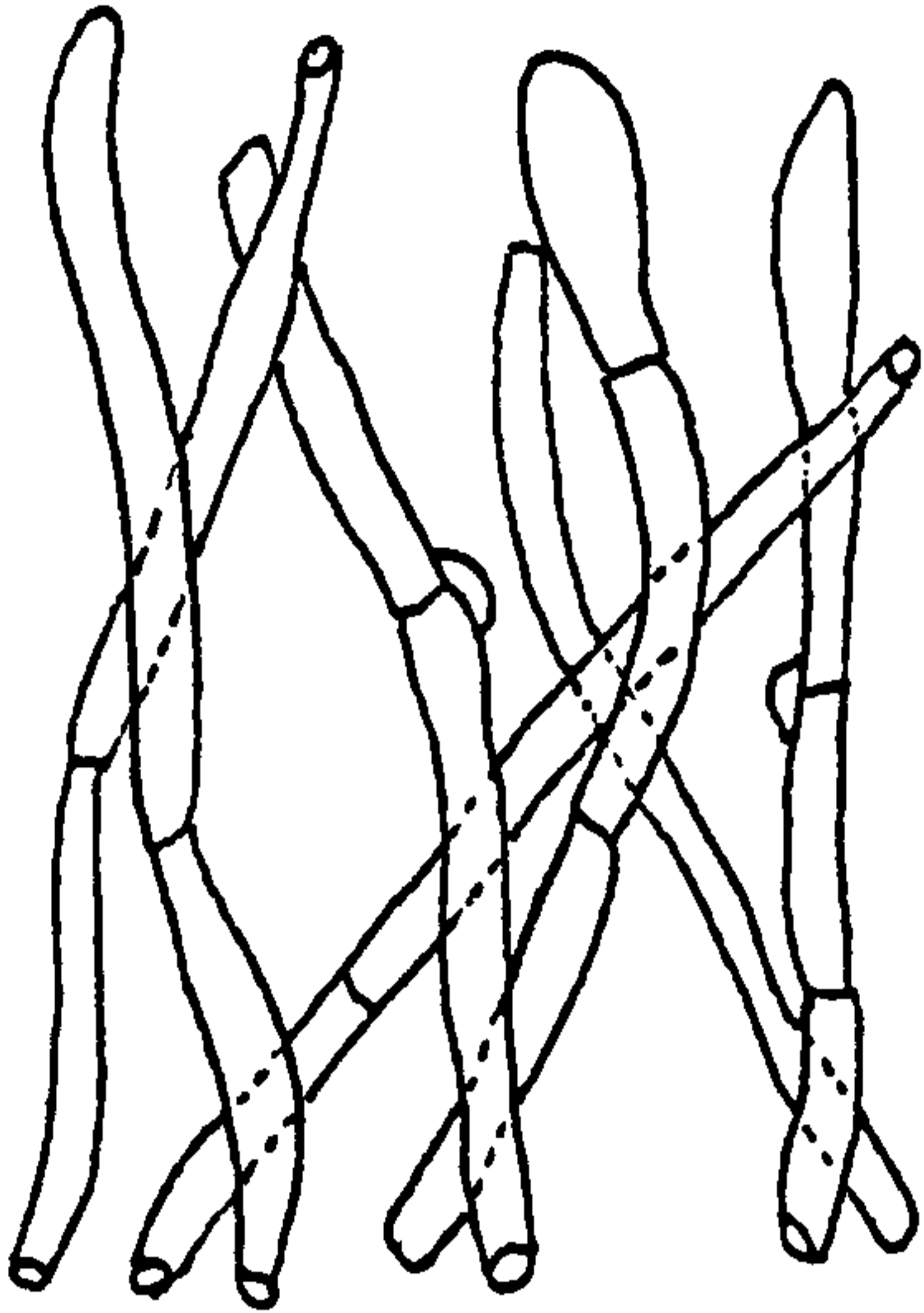
The pileus trama describes the arrangement of the hyphae in the flesh of the pileus. This trama can be one of two types: the flesh can be made of filamentous intertwined cells and form an homogeneous tissue called homomerous or it can be made of intertwined filamentous cells intersperced with nests of spherical cells. The latter arrangement is called heteromerous trama. Figure 4 illustrates these two types of trama.

This is the second of four characters used by the taxonomists to split the Agaricales into their sixteen recognised families. Although this is an important taxonomic character, it is not very useful in our context because only one large family has the feature "heteromerous" trama and only a few mildly poisonous mushrooms are found in that family (Russulaceae).

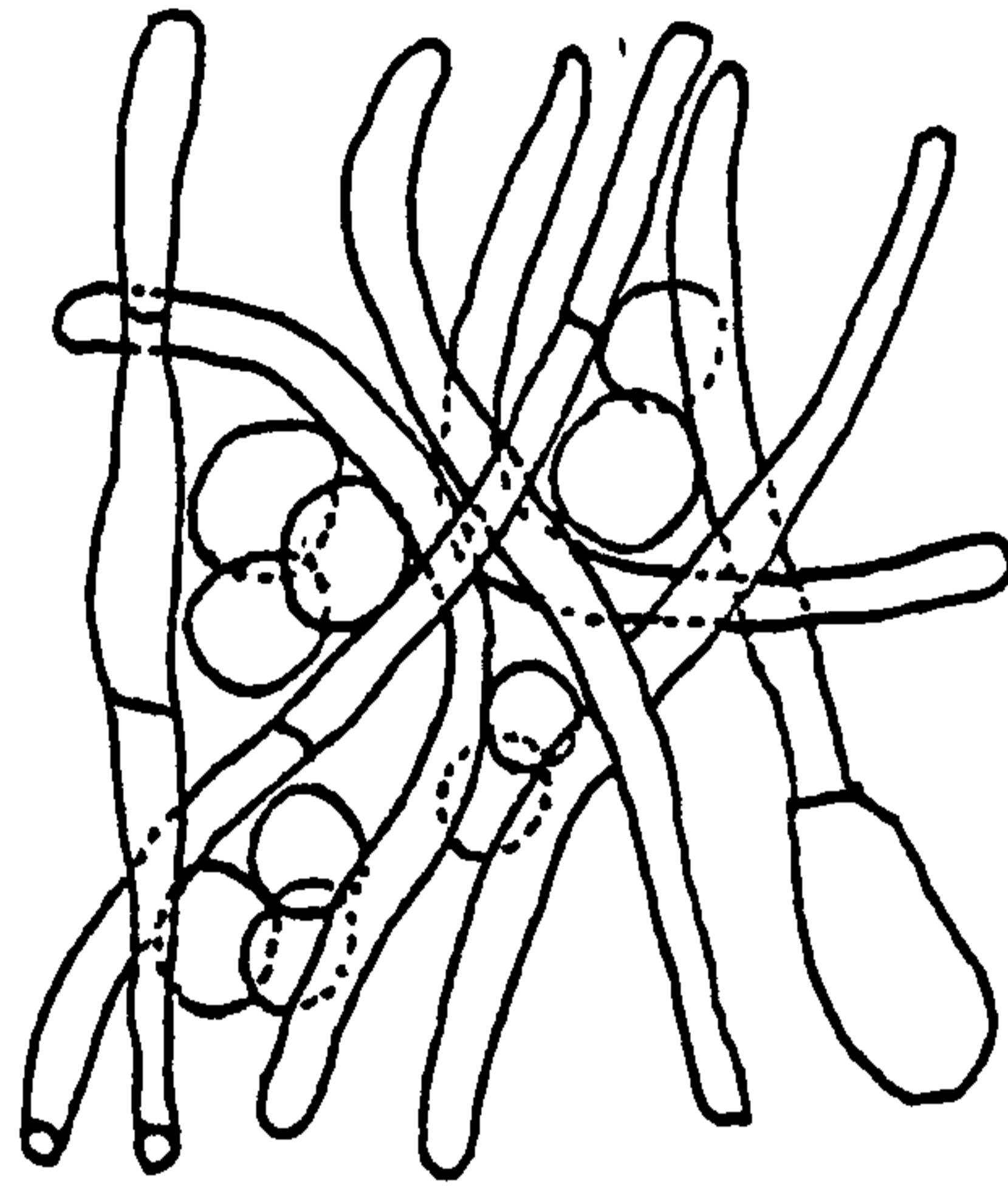
This feature can also be observed in sections of the flesh of ascocarps.

2.2.2.1.1.6. Gill Trama.(17)

- States :
- regular
 - inverse
 - bilateral
 - irregular
 - absent



homoimerous



heteromerous

Figure 4: Structures of the Pileus Trama.

As previously mentioned, gills are absent in the Ascomycetes and therefore this character cannot be observed and is qualified as "absent".

The gill trama present four different states which give a useful split within the Agaricales. It is the third character used by modern taxonomists to define the 16 families of the Agaricales (see Table 1).

The four states are described by the arrangement of the hyphal cells down the gills or lamellae. It may be "regular", where the cells are arranged in straight parallel lines, "bilateral" where the cells diverge down towards the surface of the gills like the branches of a fir tree, "inverse" where the hyphae appear to converge down towards the centre of the trama and finally "irregular" where no specific arrangement can be observed. These states are illustrated in Figure 5.

The feature "irregular" qualifies all the genera and families which show an indefinite structure, and as such, includes mushrooms with a heteromerous pileus trama extending into the gills.

2.2.2.1.1.7. The Hymenium.

On the surface and edge of the gills, a differentiated

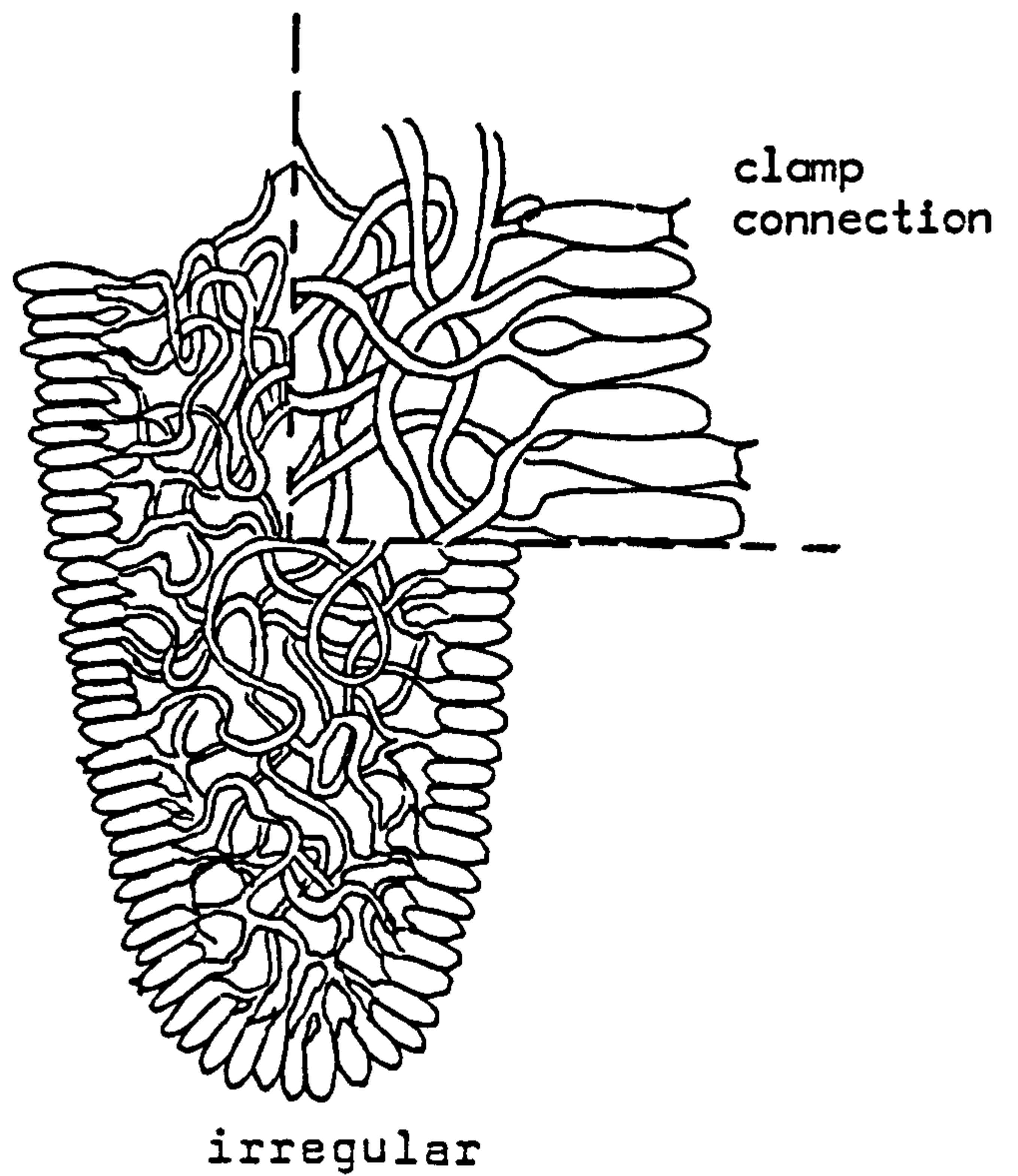
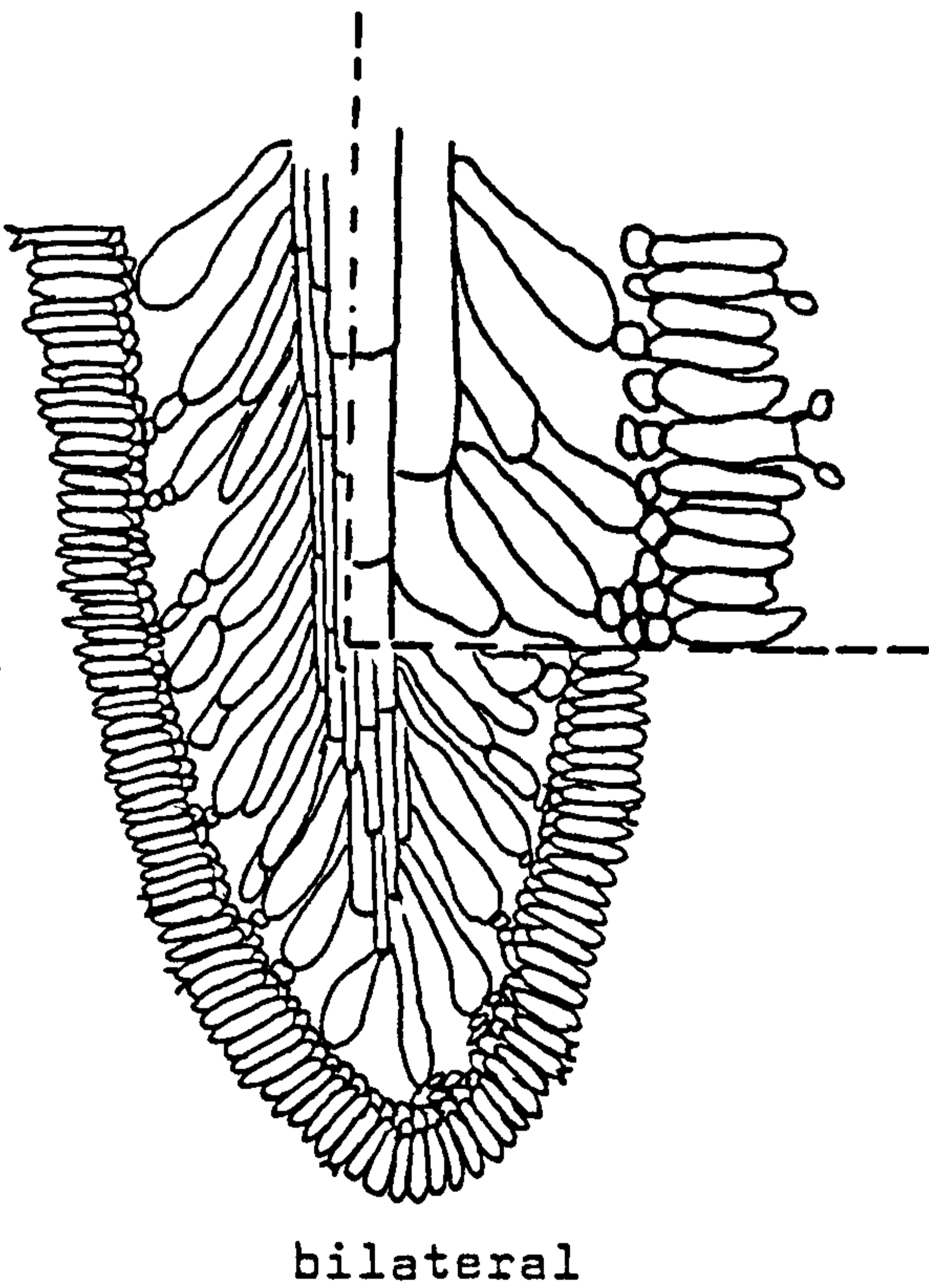
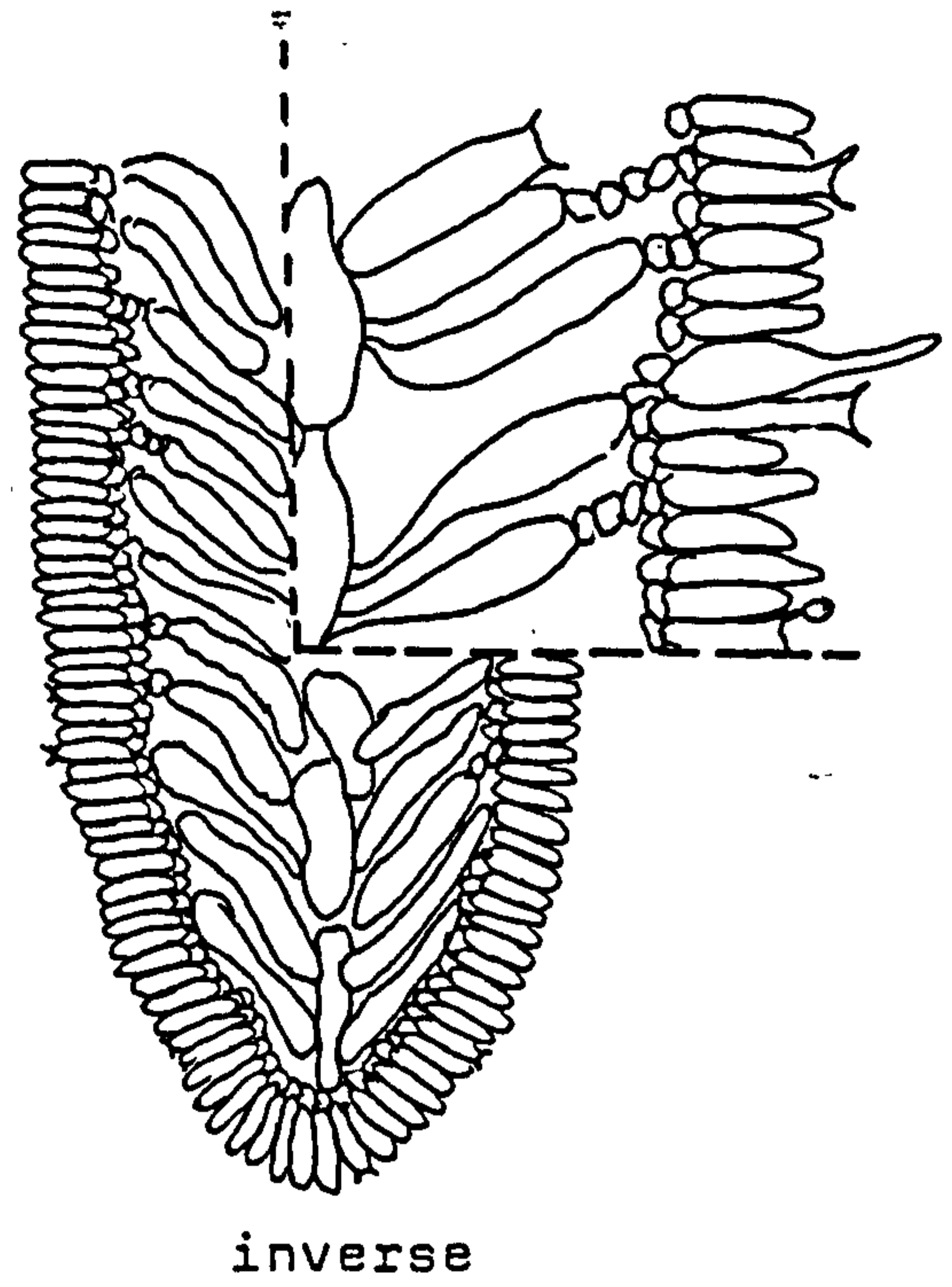
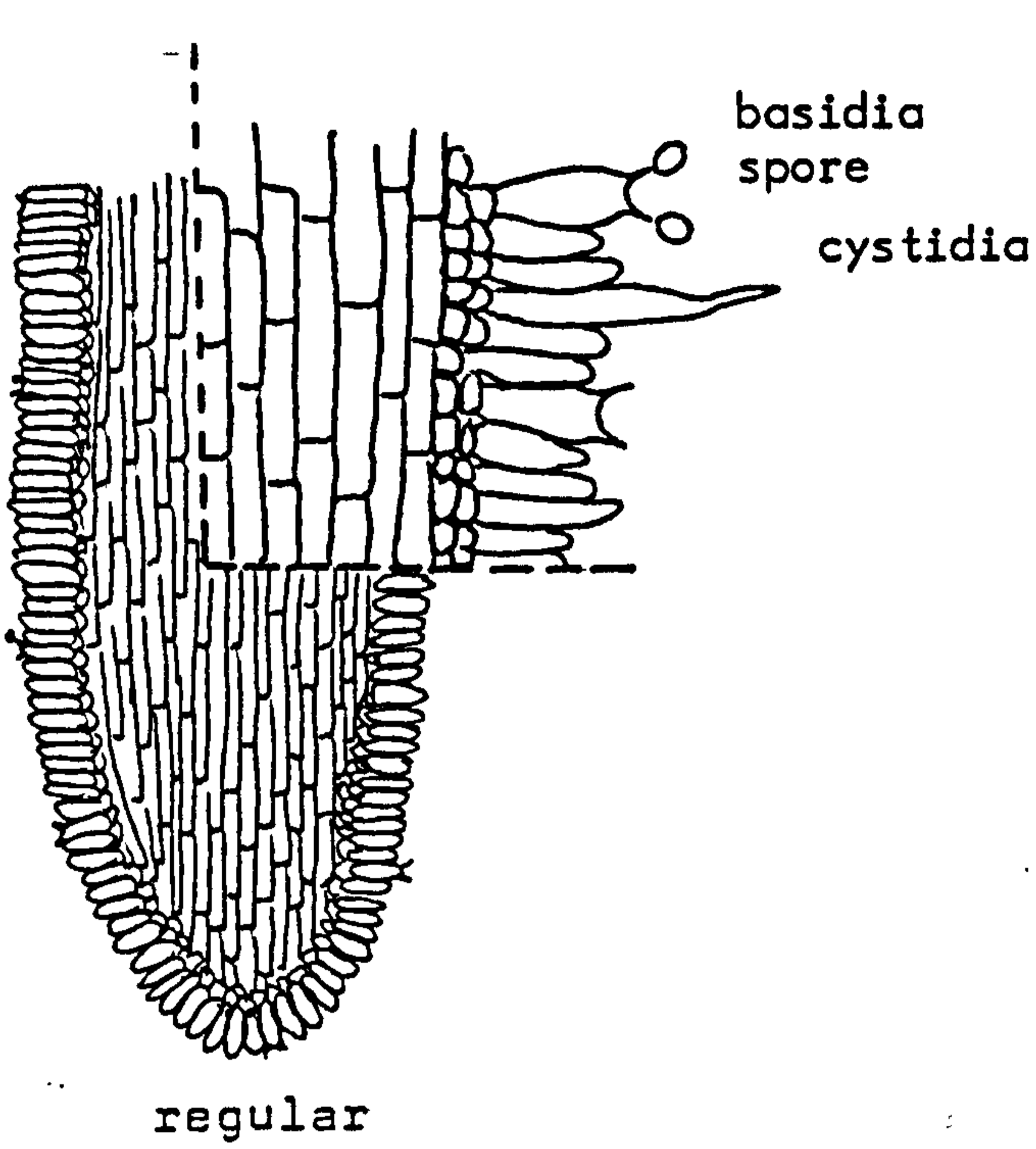


Figure 5: Gill Trama and Gill Surface (Hymenium)

layer of cells, reproductive and sterile, may be found: the hymenium. In the Ascomycetes, the genera of interest here bear their hymenium on the outer surface of the cap.

Two types of cells have interesting diagnostic features for identification: the reproductive cells, the basidia, spore bearing structures of the Basidiomycetes, or the asci, spore enclosing structure of the Ascomycetes and the sterile cells or cystidia. These latter cells are absent in the Ascomycetes and may be present on the margin (marginal cystidia = cheilocystidia) or on the faces (facial cystidia = pleurocystidia) of the gills of the Agaricales. They have specific shapes, colour and sizes which are going to be discussed subsequently.

The spore, which on germination gives rise to a new mycelium, is produced in the hymenial layer. Over the past two centuries it has been the most widely used feature for the identification of mushrooms. Under extreme treatment, spores are very resistant and retain their diagnostic features. It is such an important and stable structure that it accounts for eight identification characters used here. Although not totally independent, these characters are all included because they are easy to observe and qualify.

2.2.2.1.1.8. Colour of the Spore Print. (13)

- States :
- white
 - pink
 - brown
 - black

The most important character used in modern and traditional keys is the colour of the spores, best observed as a spore print.

Although the range of colours is large, it can be conveniently divided into the four above mentioned states.

The state "white" includes the white, the pale cream to cream and the pale to brightly yellow (greenish in one case) coloured spores. "Pink" varies from bright pink to salmon, coral and peach while "brown" includes various shades of brown from dull clay-brown or ochraceous to bright rust-brown. Finally "black" covers the purplish, and the purple black to black shades. Reference colours are found in the "Flora of British Fungi: Colour Identification Chart" (Henderson et al. 1969) available separately from the Royal Botanic Garden in Edinburgh.

The spore print colour is the fourth microscopical character used by taxonomists to divide the Agaricales

into families (Table 1). It is now possible using the first four anatomical characters described above to place a mushroom into a family. The obvious result of this is that if a mushroom is involved in a poisoning case and these four features are available, then the species must belong to only a few related species. This is demonstrated and summarised in Table 1.

2.2.2.1.1.9. Spore Colour under the Microscope. (6)

<u>States</u> :	- clear	<u>Substates</u> :	- amyloid
			- inamyloid
			- dextrinoid
			- uncertain
	- pale brown		
	- pale pink		
	- dark	<u>Substates:</u>	- resistant
			- dissolves
			- uncertain

The colour of a spore print is one of the most useful and easily observed characters, but it is rarely available in a poisoning situation because a fresh specimen is required before a spore print can be made!

Characters Families	pileus trama	gill trama	spore print	cuticle	other characteristic features
Russulaceae	Het	Irr	w	Fil	amyloid spores
Boletaceae	Hom	Bil	br	Fil	no gills:tubes
Paxillaceae	Hom	Bil	br	Fil	
Gomphidiaceae	Hom	Bil	bl	Fil	
Amanitaceae	Hom	Bil	w	Fil	
Plutaceae	Hom	Inv	p	Fil/Cel	
Entolomataceae	Hom	Reg	p	Fil	angular spores
Agaricaceae	Hom	Reg	bl	Fil	
Lepiotaceae	Hom	Reg	w	Fil/Cel	dextrinoid spores
Strophariaceae	Hom	Reg	bl	Fil	
Cortinariaceae	Hom	Reg	br	Fil	
Coprinaceae	Hom	Reg	bl	Cel	
Bolbitiaceae	Hom	Reg	br	Cel	
Hygrophoraceae	Hom	Reg/Bil	w	Cel/Fil	long basidia
Tricholomataceae	Hom	Reg/Bil	w	Cel/Fil	
Pleurotaceae	Hom	Irr	w	Fil	

Table 1 : see the legend of the facing page

Table 1: The sixteen families of the Agaricales
as defined by four anatomical characters
(based on a field course by Dr. Roy Watling,
Royal Botanic Garden, Edinburgh)

The abbreviations are the first letters of
the respective states of the different

characters: Het = heteromerous

Hom = homoimerous

Irr = irregular

Bil = bilateral

Reg = regular

Inv = inverse

w = white

br = brown

bl = black

p = pink

Fil = filamentous

Cel = cellular

They are discussed in chapters 2.2.2.1.1.3;
2.2.2.1.1.5; 2.2.2.1.1.6 and 2.2.2.1.1.8.

While an experienced microscopist and mycologist is able to categorise the spore colour accurately from its microscopical examination, it cannot be expected that the same result will be obtained from the non-specialist but he should be able to distinguish the states "clear" (also called transparent or hyaline) and "dark" fairly accurately. These two states correspond exactly to the states "white" and "black" described in the previous chapter (2.2.2.1.1.8.) The unexperienced analyst might not be able to distinguish between the two intermediate states, "pale brown" and "pale pink" ("brown" and "pink" of the previous character respectively). Users should be aware of this drawback and avoid this character if they are uncertain with its determination, despite its tremendous informative content.

The state "clear" is divided into four substates. A chemical reagent, Melzer's Iodine (see appendix 6.1.1.) is necessary to observe these substates. When the reagent is unavailable to the analyst, the feature remains "uncertain". Otherwise, three reactions may occur. The spore surface can become blue to purple in colour and is usually referred to as "amyloid". Alternatively, the spore can remain unchanged or clear and is termed "inamyloid". If the spore turns reddish to purplish brown it demonstrate the feature called "dextrinoid" by British authors (Henderson et al. 1969) or "pseudoamyloid" by American

authors (Singer, 1975). The chemical significance of these reactions is not known.

Melzer's reagent is relatively unstable and it makes its application impractical, but this drawback is compensated by the usefulness of the reaction in separating some of the most dangerous toxic mushrooms into definite small groups.

Similarly, the state "dark" is further divided into three substates after treatment with concentrated sulphuric acid. The pigments of the dark spores either resist (i.e. substate "resistant") or dissolve in the acid (i.e. substate "dissolves"). When the acid is unavailable and the reaction is not attempted the substate is defined as "uncertain". This reaction has a limited interest and is only useful in separating genera within the Coprinaceae family (see Table 2).

2.2.2.1.1.10. Spore Length and Spore Breadth. (11 and 12)

- Length states :
- less than 6 μm
 - 6 - 8 μm
 - 8 - 10 μm
 - 10 - 12 μm
 - 12 - 17 μm
 - more than 17 μm

BASIDIOMYCETES.

Order- Agaricales

I. Agaricaceae

Agaricus Fr.

II. Amanitaceae

Amanita (Fr.) Gray

III. Bolbitiaceae

Conocybe Fayod

IV. Boletaceae

Boletus Fr.

Tylopilus Karst.

V. Coprinaceae

Copelandia Bres.

Coprinus (Fr.) Gray

Panaeolus (Fr.) Quél.

VI. Cortinariaceae

Cortinarius Fr.

Galerina Earle

Gymnopilus Karst.

Hebeloma (Fr.) Kum.

Inocybe (Fr.) Fr.

VII. Gomphidiaceae

VIII. Hygrophoraceae

Hygrocybe (Fr.) Wünsche

IX. Lepiotaceae

Lepiota (Fr.) Gray

Leucocoprinus Pat.

X. Paxillaceae

Paxillus Fr.

XI. Pleurotaceae

XII. Entolomataceae

Entoloma (Fr.) Kum.

Nolanea (Fr.) Kum.

XIII. Russulaceae

Lactarius Gray

Russula (Fr.) Gray

Table 2: Systematic List of Genera Containing Poisonous
Species of Mushrooms Included in the Data Bank.

XIV. Strophariaceae

Hypholoma (Fr.)Kum.

Psilocybe (Fr.)Kum.

Stropharia (Fr.)Quél.

XV. Tricholomataceae

Clitocybe (Fr.)Kum.

Tricholoma (Fr.)Kum.

Tricholomopsis Sing.

XVI. Plutaceae

ASCOMYCETES.

Order- Pezizales

A. Morchellaceae

Morchella Dill.

B. Helvellaceae

Gyromitra Fr.

Helvella L.

Table 2 (end)

- Breadth states :
- less than $4\mu\text{m}$
 - 4 - $6\mu\text{m}$
 - 6 - $8\mu\text{m}$
 - 8 - $10\mu\text{m}$
 - more than $10\mu\text{m}$

Spore dimensions in terms of spore length and breadth are two continuous quantitative characters and have a relatively poor diagnostic value, especially if one takes into account the biological variations. States are expressed in discrete ranges, large enough to allow for the variations and small enough to retain some diagnostic value. States being continuous in reality, they overlap in some cases and are not mutually exclusive, this being one major drawback with this type of character, where multiple answers are the rule rather than the exception. It therefore becomes practically useless in the identification of medium sized spored species. It is nevertheless retained because of its ease of observation and, when only a few characters are available for identification, it may help to reduce the number of species suspected for a given intoxication.

Mature spores should always be measured, preferably from spore deposits. Abnormal spores such as those originating from two-spored basidia, where most of the spores

originated from four-spored basidia, should be avoided.

2.2.2.1.1.11. Spore Shape. (7)

- States :
- globose
 - ellipsoid
 - angular
 - various

The spore shape is an extremely useful character when it comes to the identification of poisonous mushrooms although it is partially dependent on the sizes of the spores (or vice-versa; e.g. a globose spore has a length equal to the breadth!). The range of shapes is extremely large, but can be usefully reduced into the four above mentioned categories. It demonstrates all the advantages of a good character: it is stable, selective and easy to observe. For these reasons it is widely used in traditional as well as modern keys.

2.2.2.1.1.11.1. Globose Spores.

Globose spores should theoretically be perfectly spherical, i.e. their length/breadth ratio should be 1. For practical reasons it includes all the rounded shaped spores with a length/breadth ratio no bigger than 1.2. This ratio is used by some modern taxonomists who

maintain that it is more meaningful than a separate measure of the length and the breadth since it appears to be less prone to variations within a species.

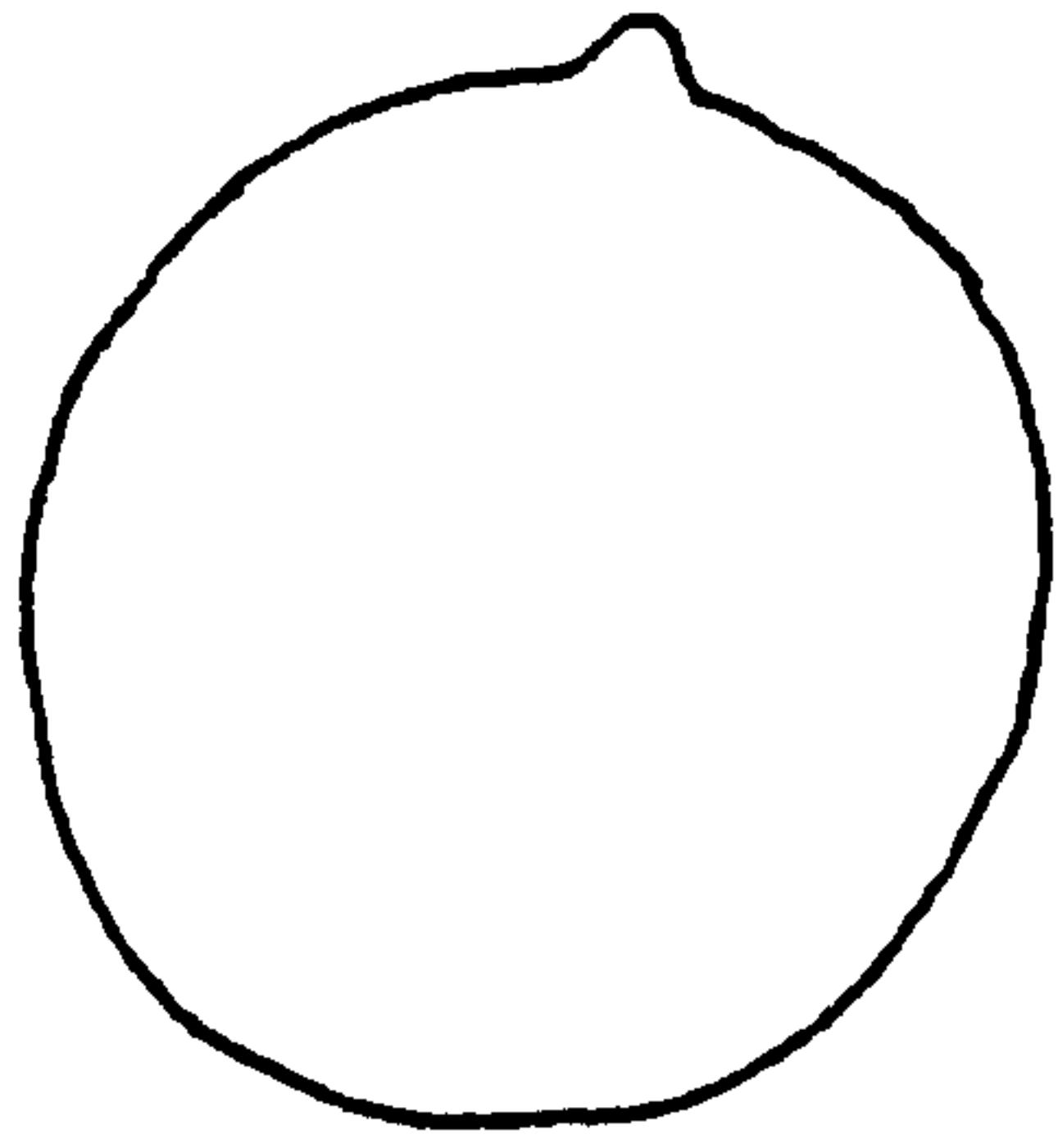
Globose spores include spores defined by mycologists as subglobose, dacryoid (=lachryform), cordate and rostrate (Figure 6).

2.2.2.1.1.11.2. Ellipsoid Spores.

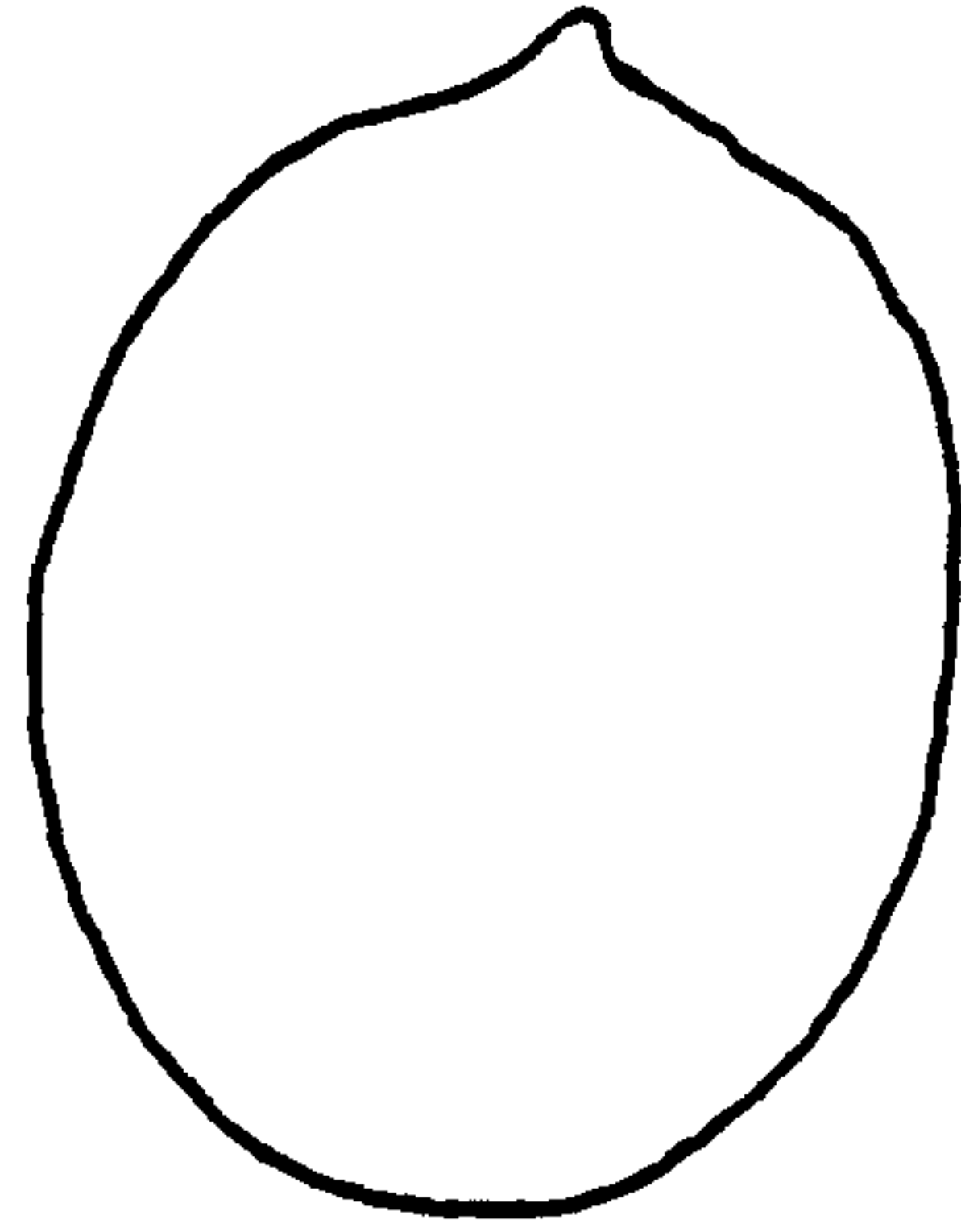
Ellipsoid spores include a whole variety of shapes and refer to the largest group of fungi. The shapes are all curved and have a length/breadth ratio always greater than 1.2. They can be ovate, pyriform, amygdaliform, fusiform, phaseoliform (bean-shaped), etc (Figure 7).

2.2.2.1.1.11.3. Angular Spores.

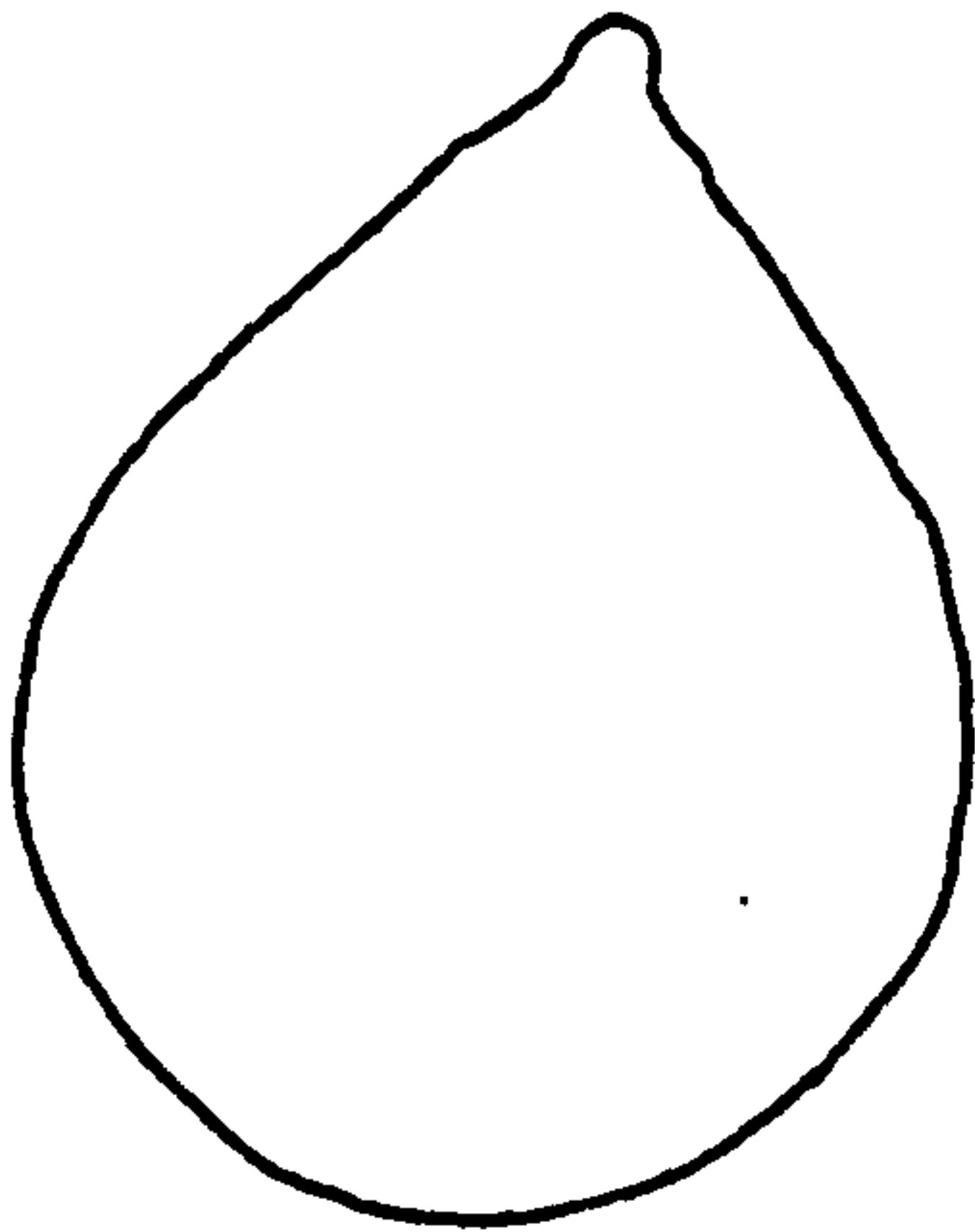
All the spores which exhibit distinct angular patterns, no matter what their orientation is, have their place here. The whole family of the Entolomataceae is separated from all other families by this character. The appearance of the spores may range from almost square to polygonal. (Figure 8)



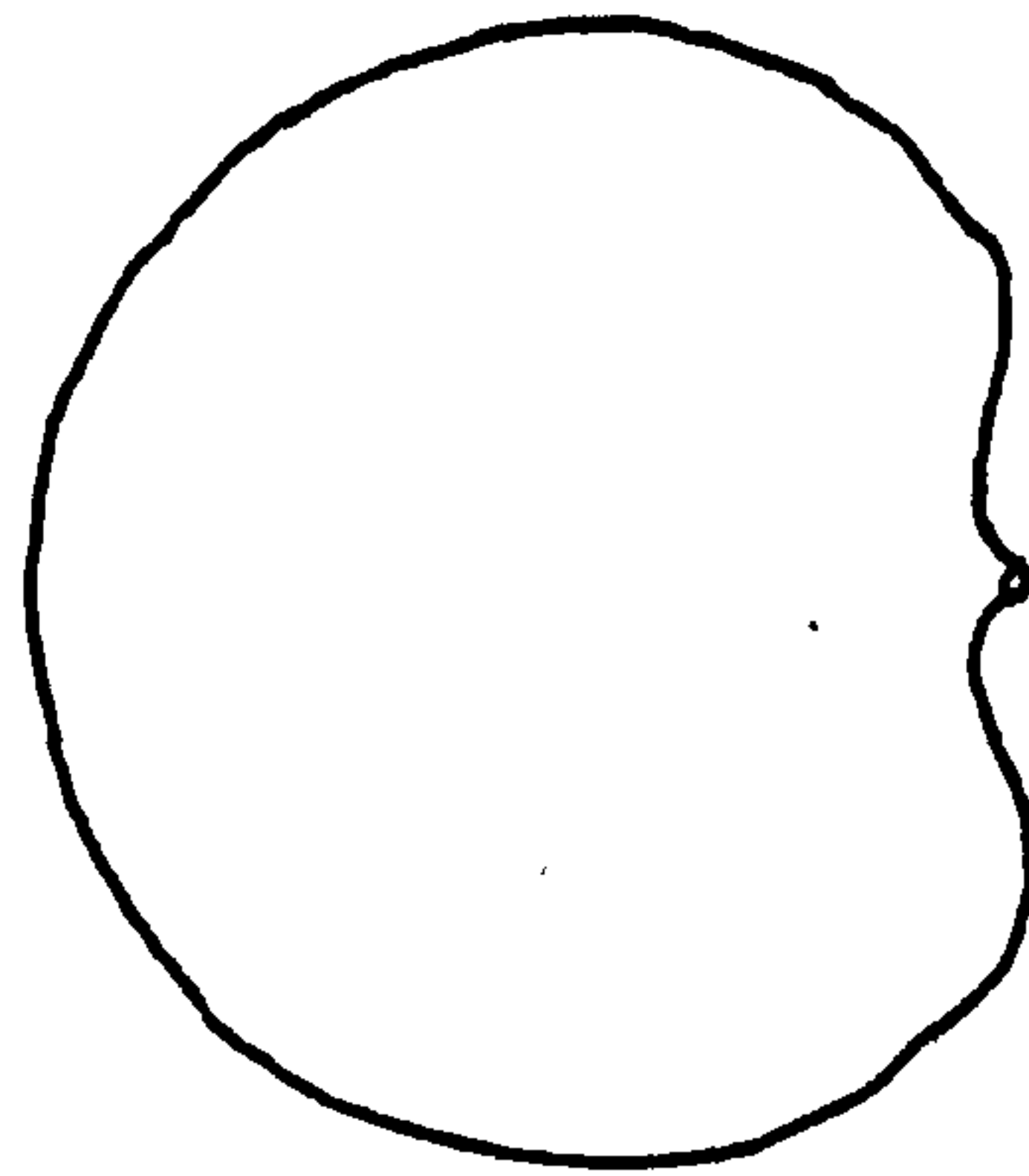
globose



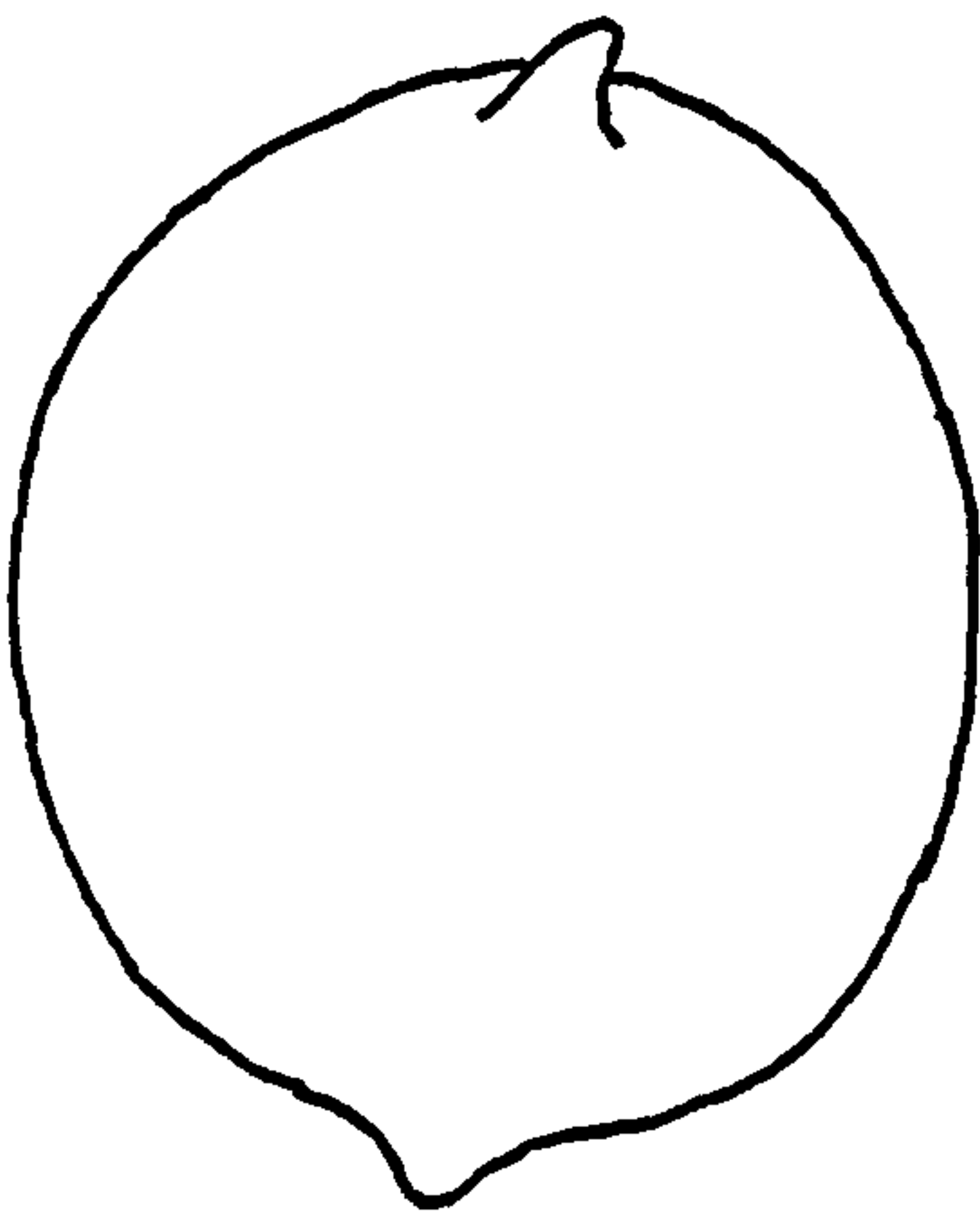
subglobose



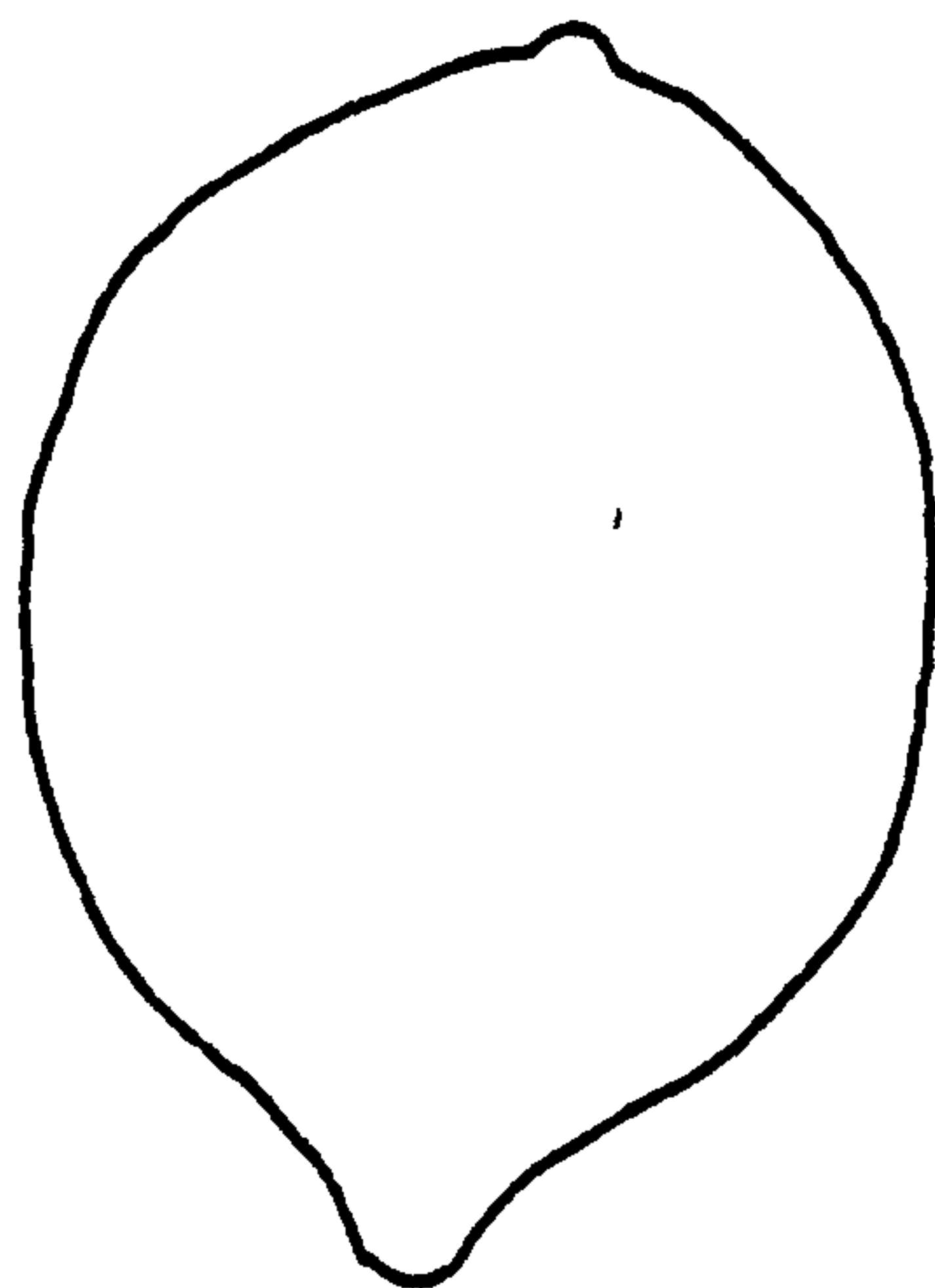
dacryoid



cordate

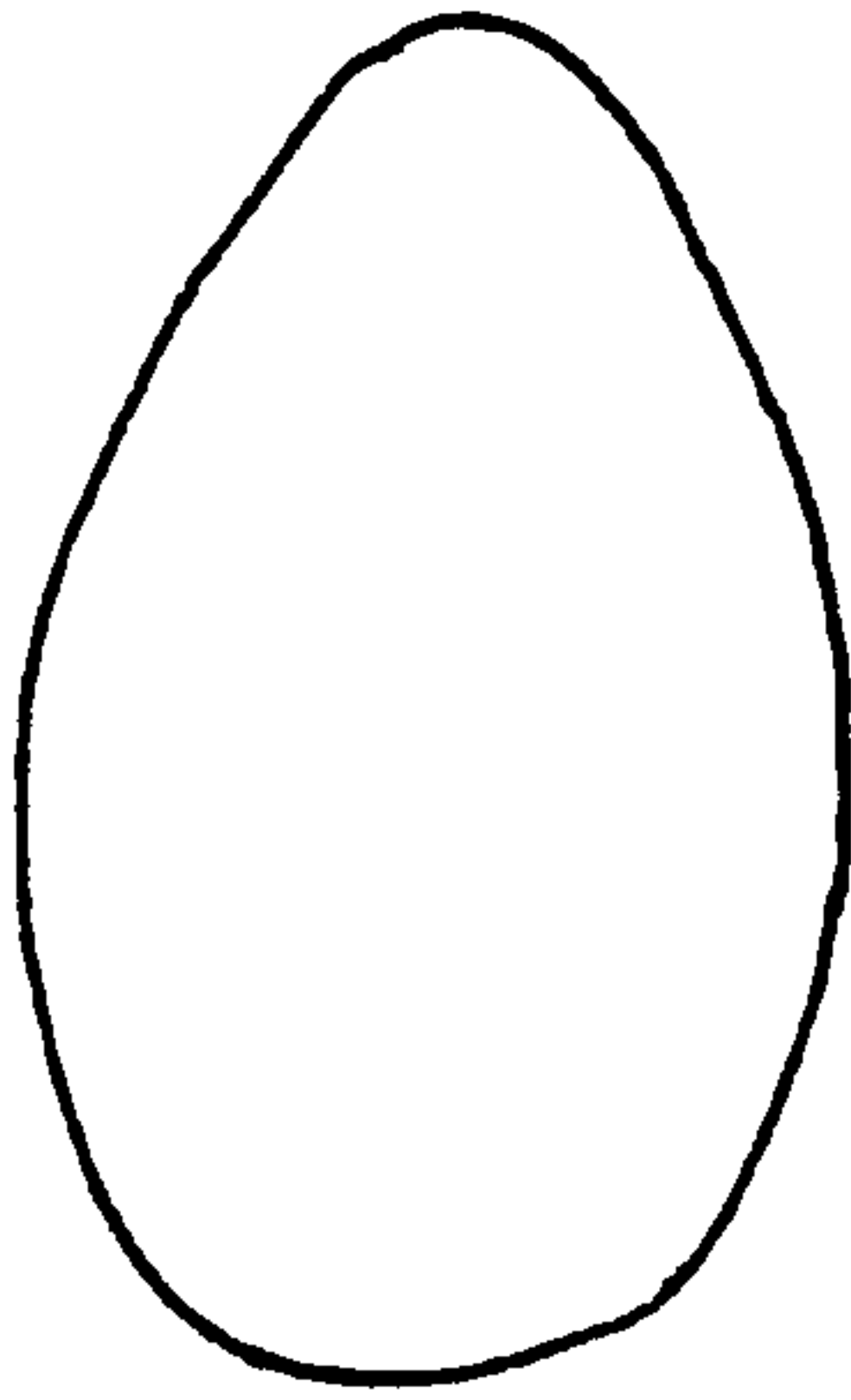


rostrate

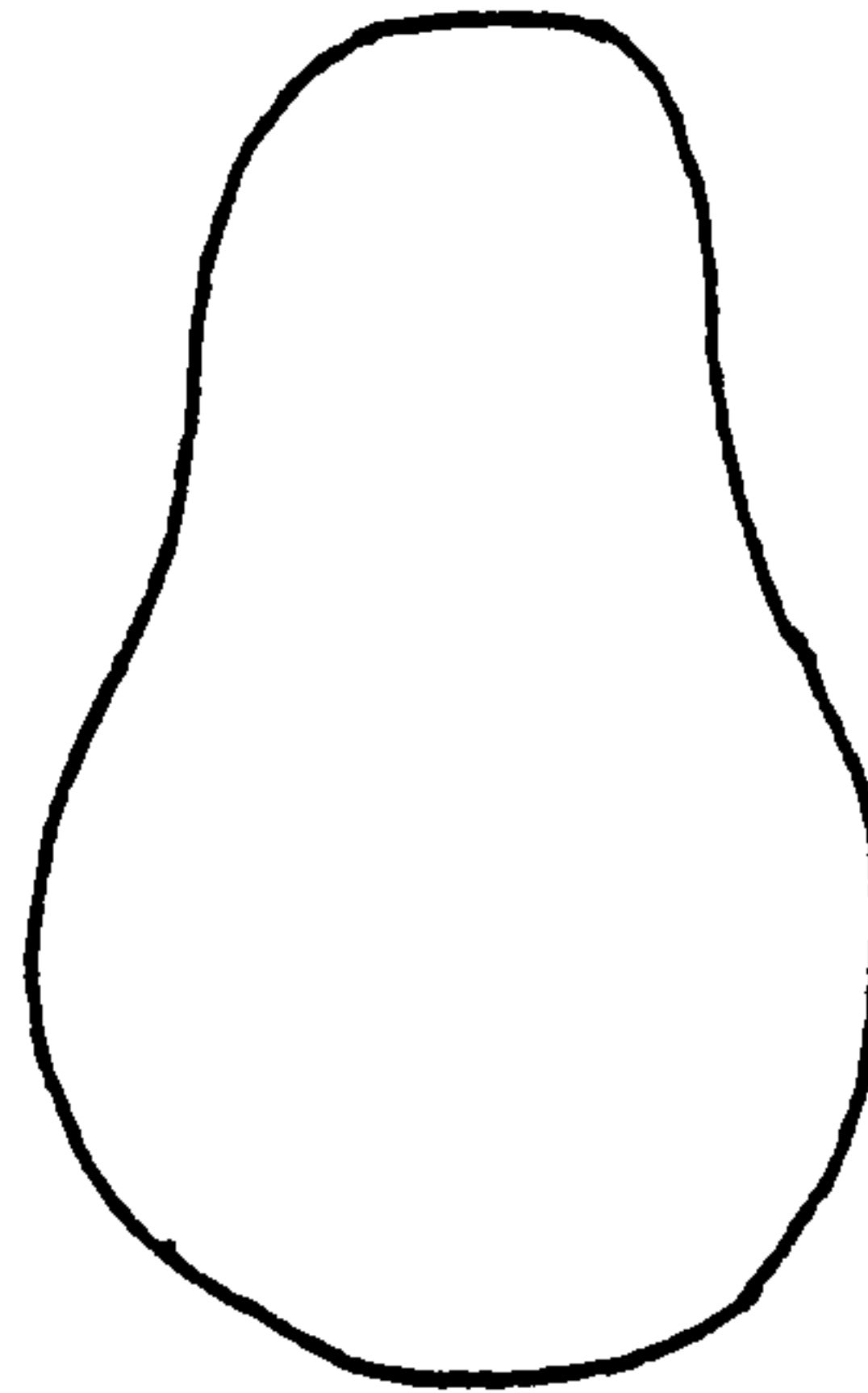


citriform

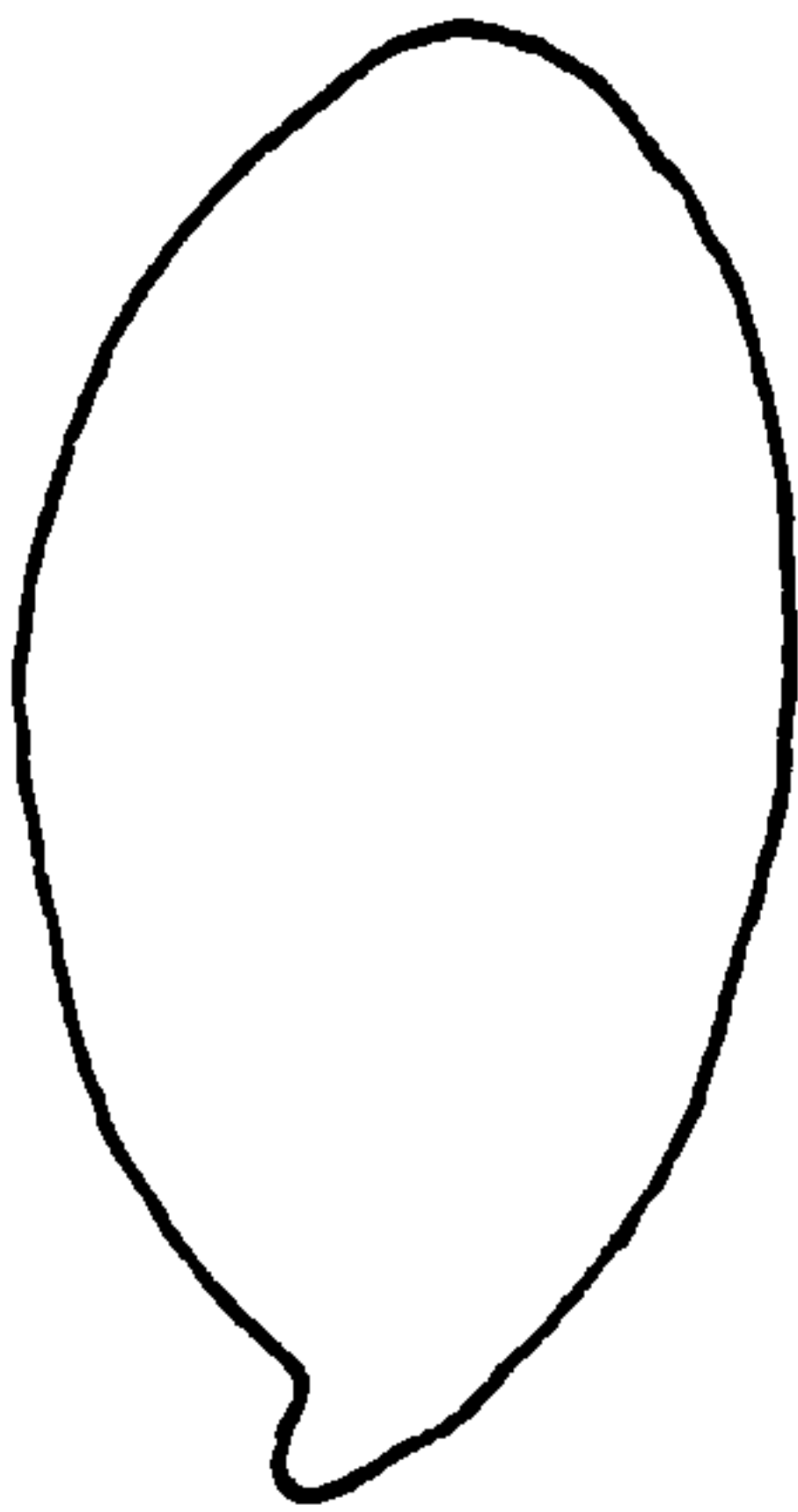
Figure 6: Examples of Globose Spores



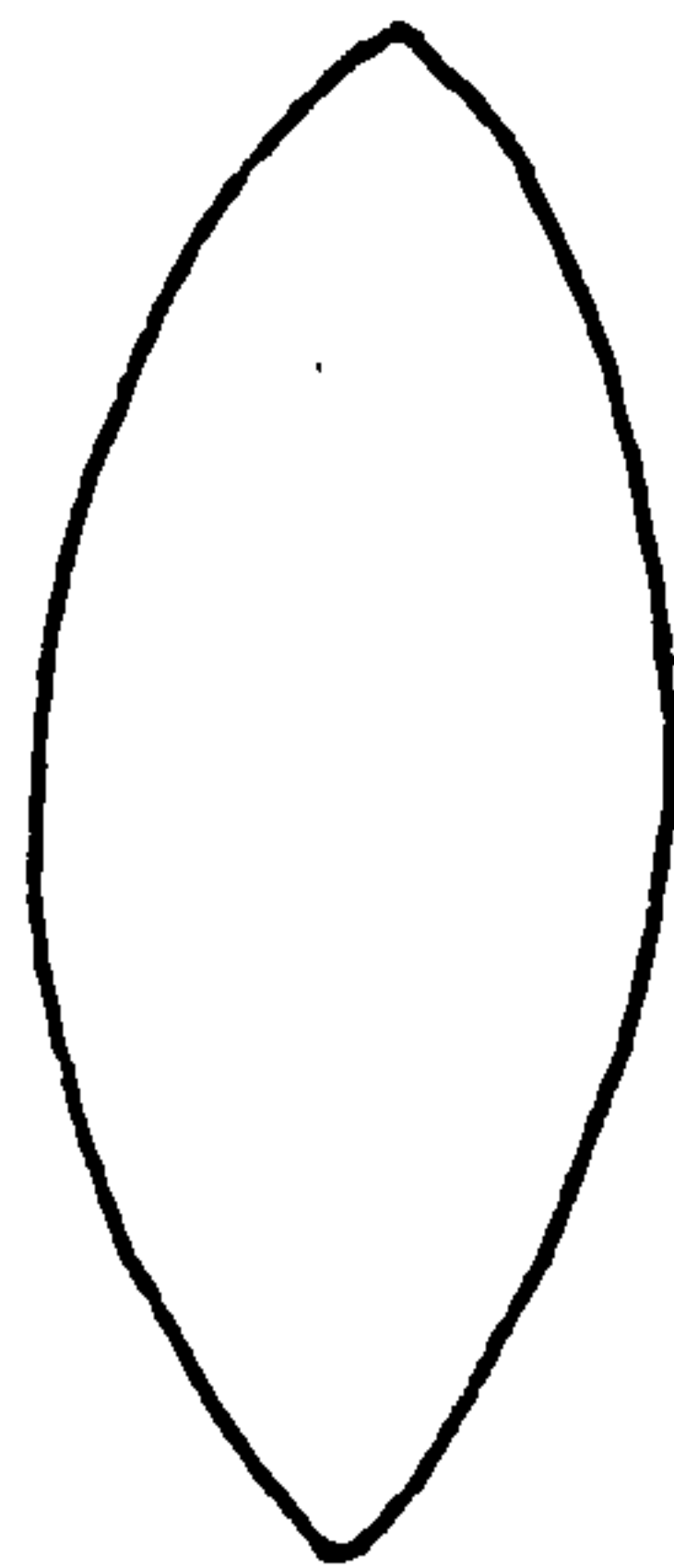
ovate



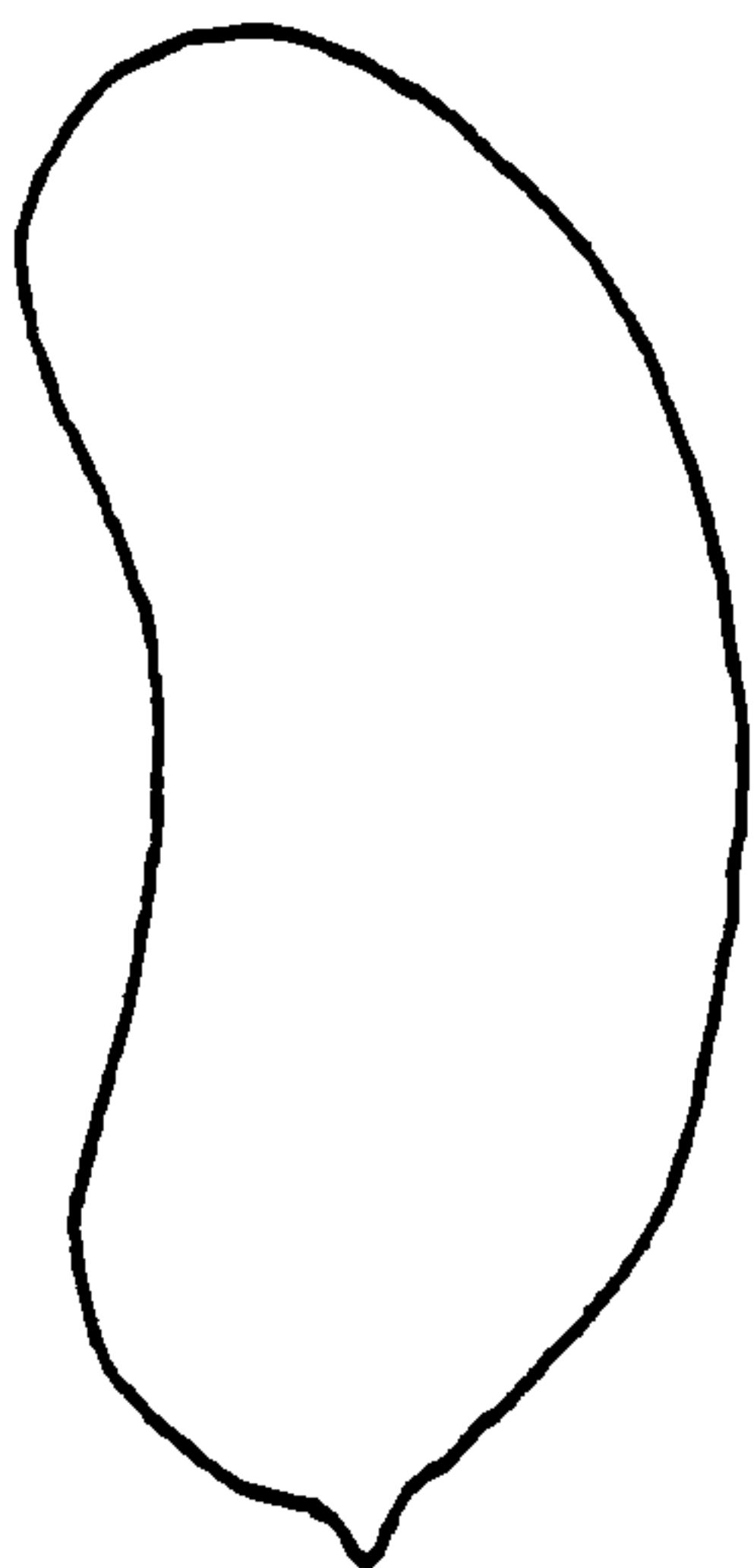
pyriform



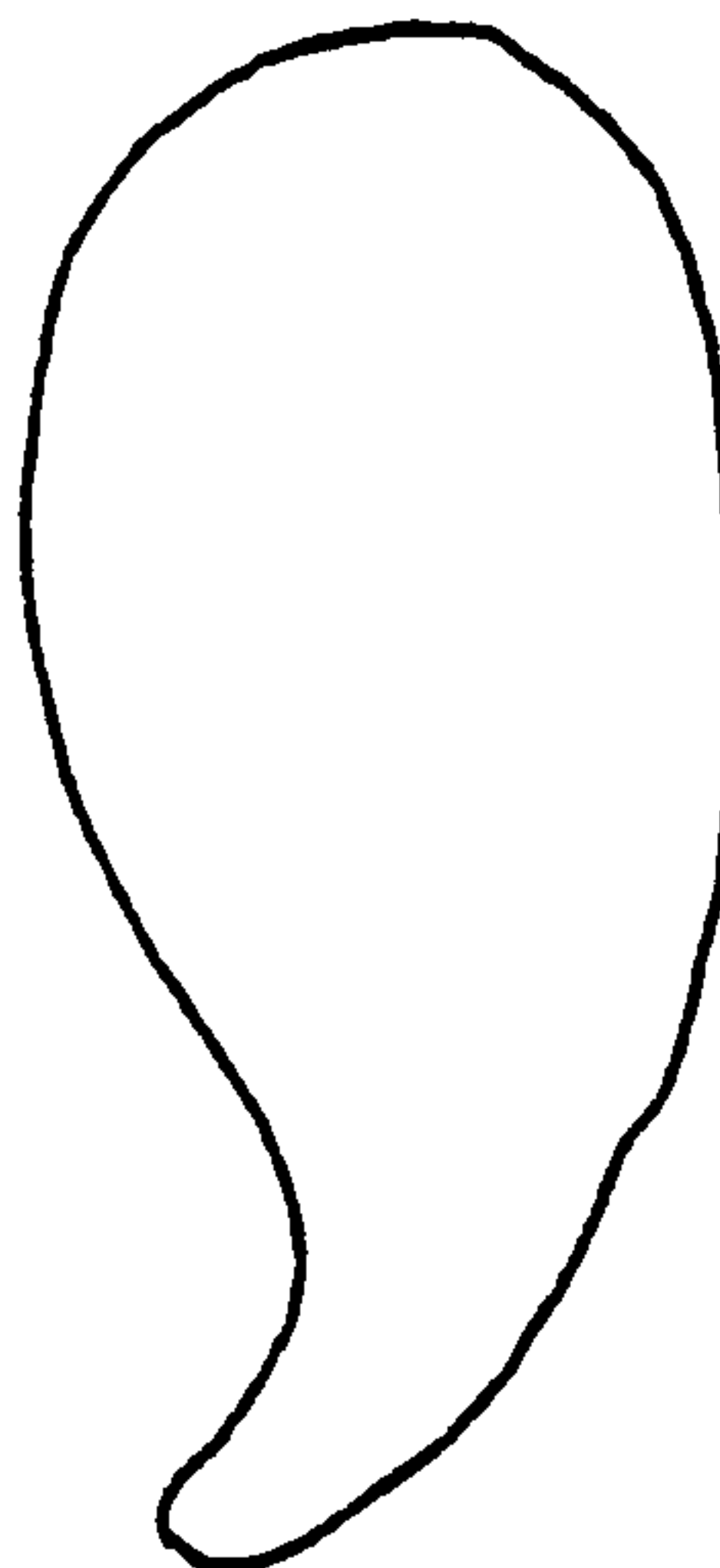
amygdaliform



fusiform

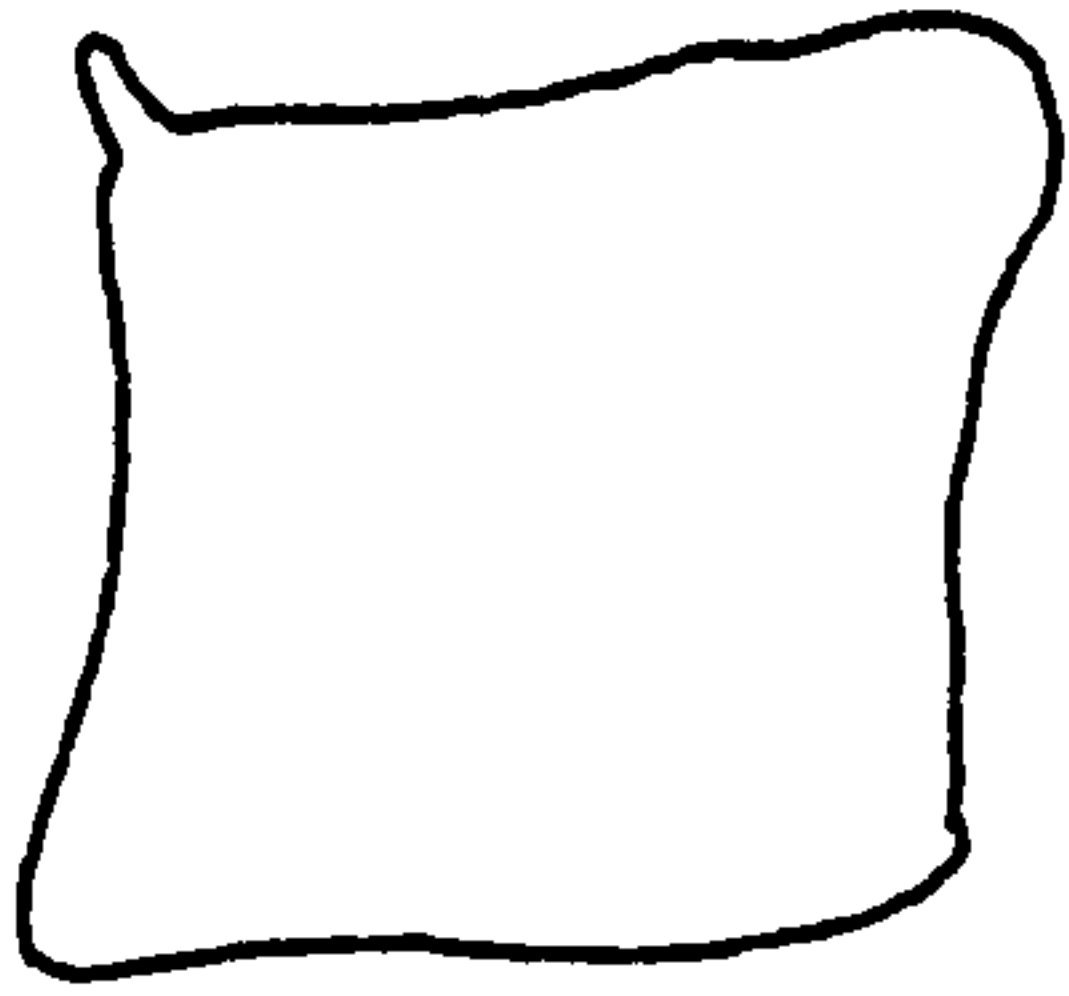


phaseoliform

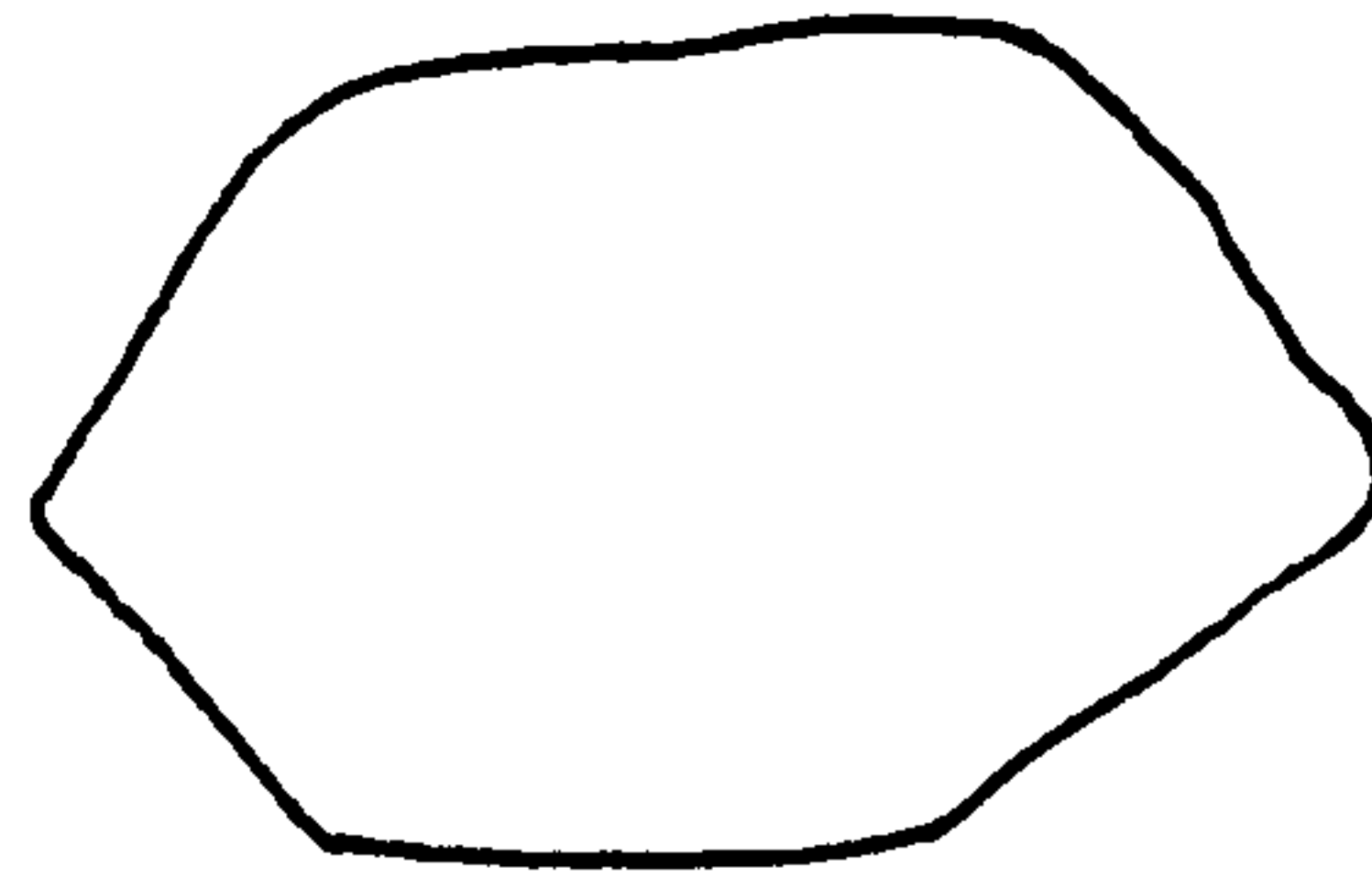


pip-shaped

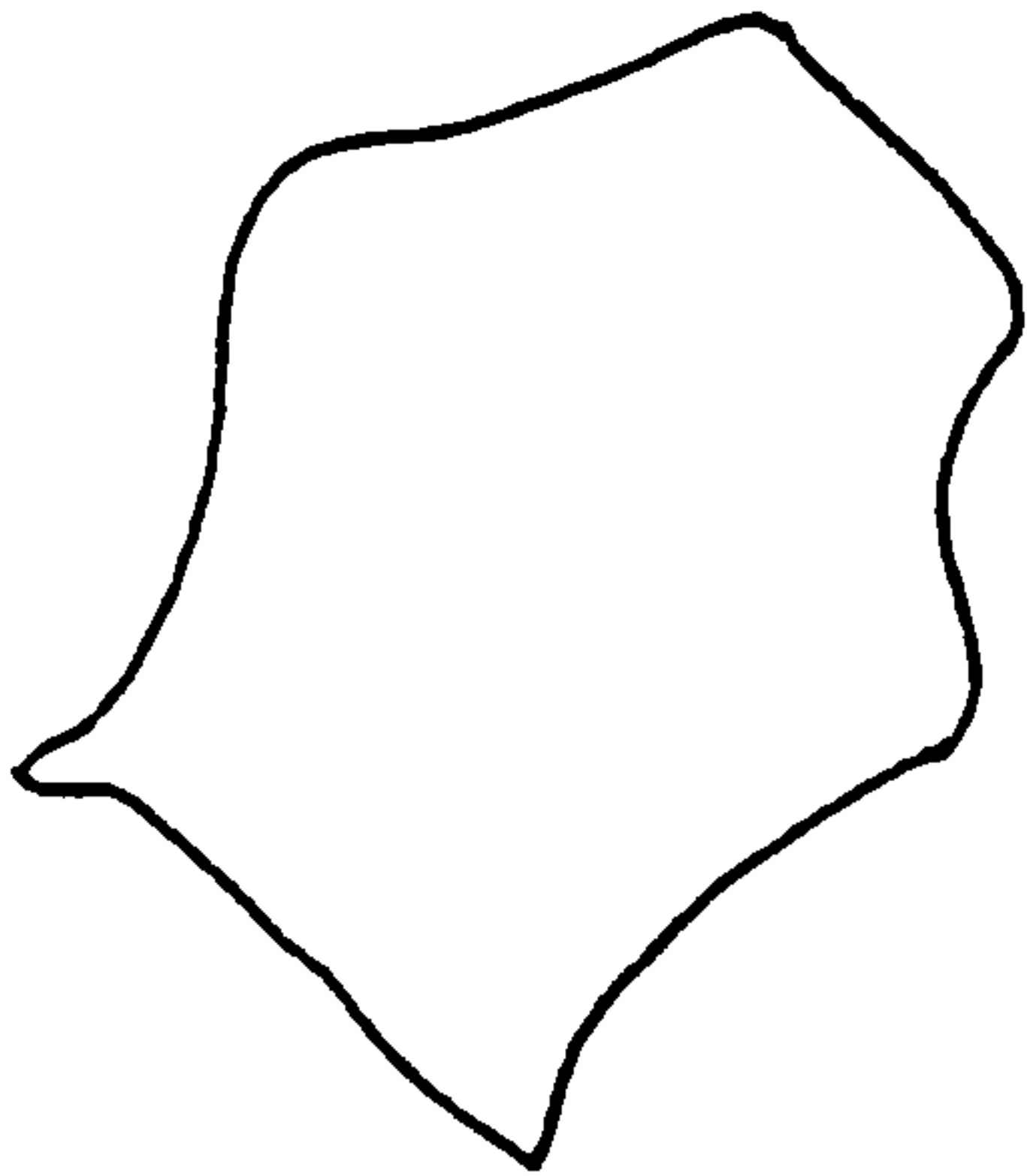
Figure 7: Examples of Ellipsoid Spores



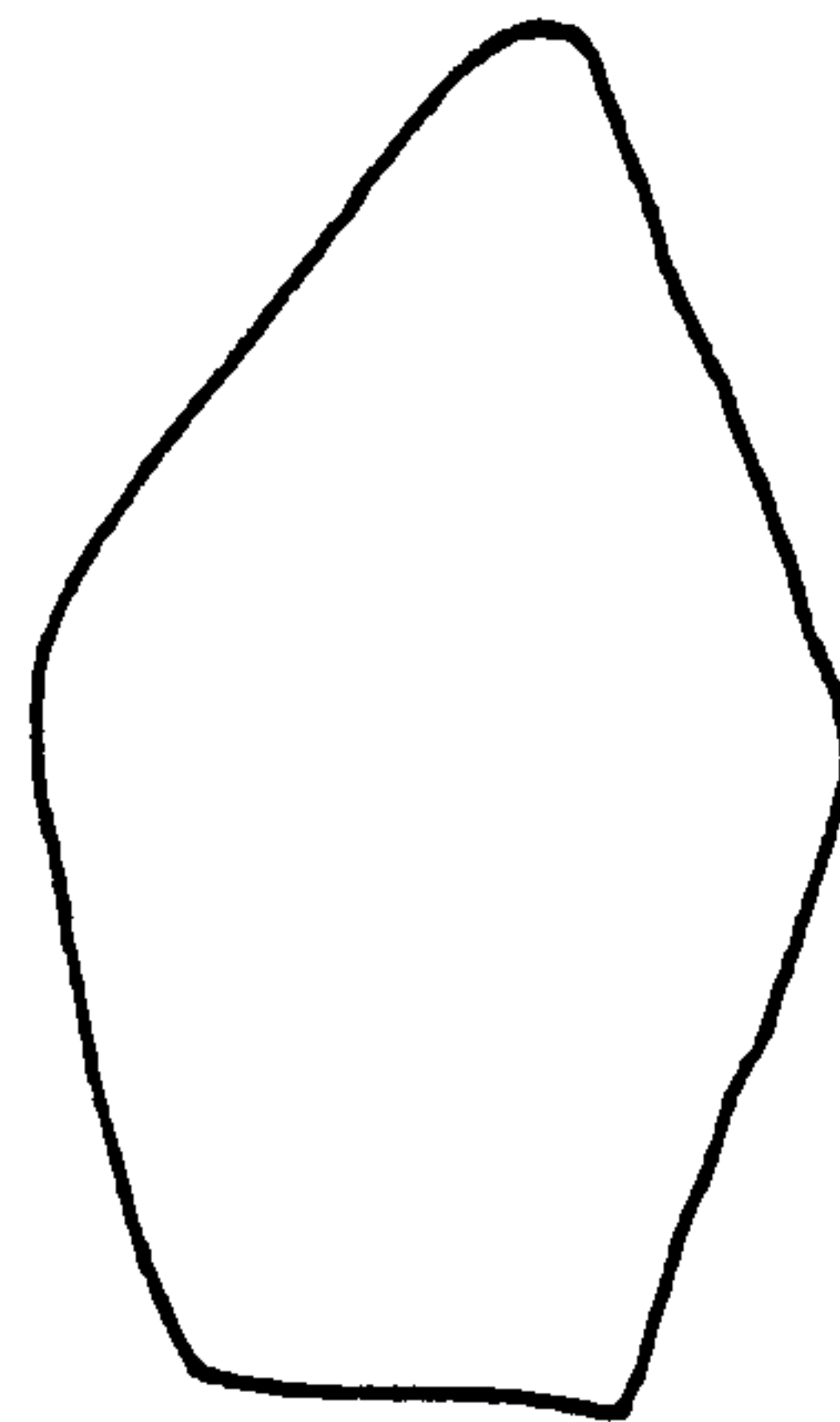
rhomboid



hexagonal



prismatic



pentagonal

Figure 8: Examples of Angular Spores

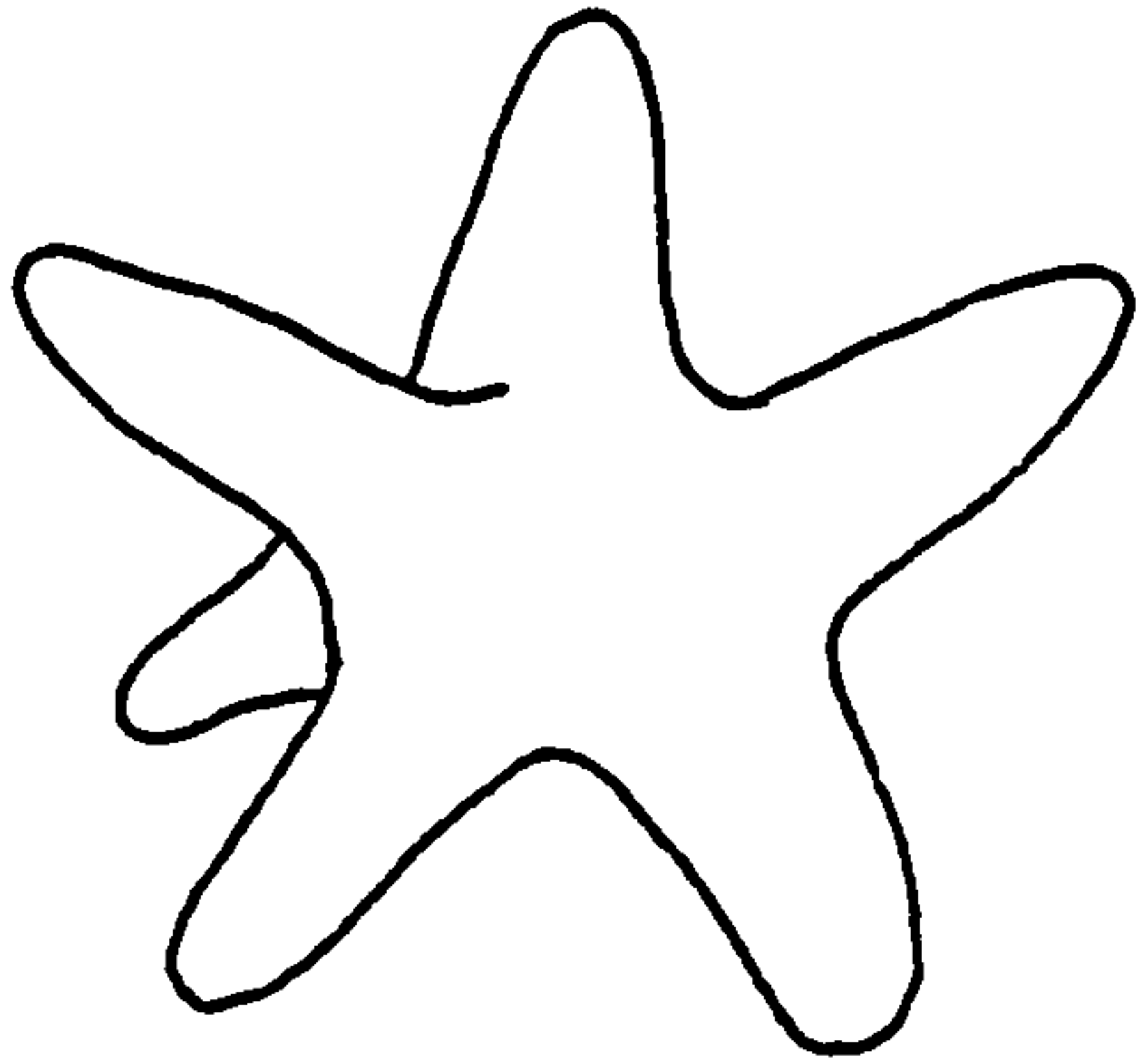
2.2.2.1.1.11.4. Various.

All spores which cannot be fitted into the first three categories come under this heading. They comprise stellate, nodulose, naviculate, bacilliform, cylindrical, etc (Figure 9). This helps in resolving a genus which contain a great number of poisonous species (Inocybe).

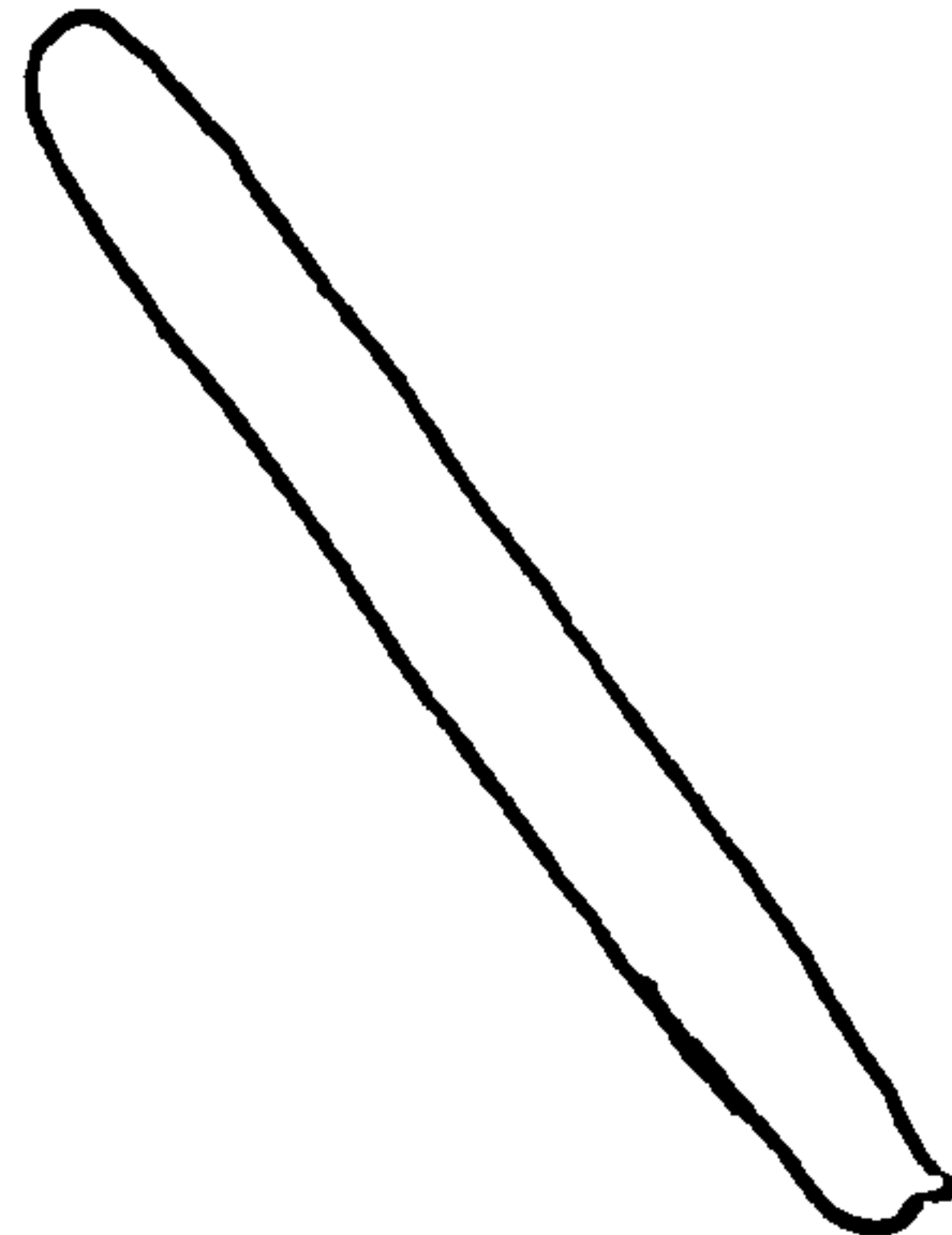
2.2.2.1.1.12. Spore Surface. (8)

- States :
- smooth
 - rough
 - ridged

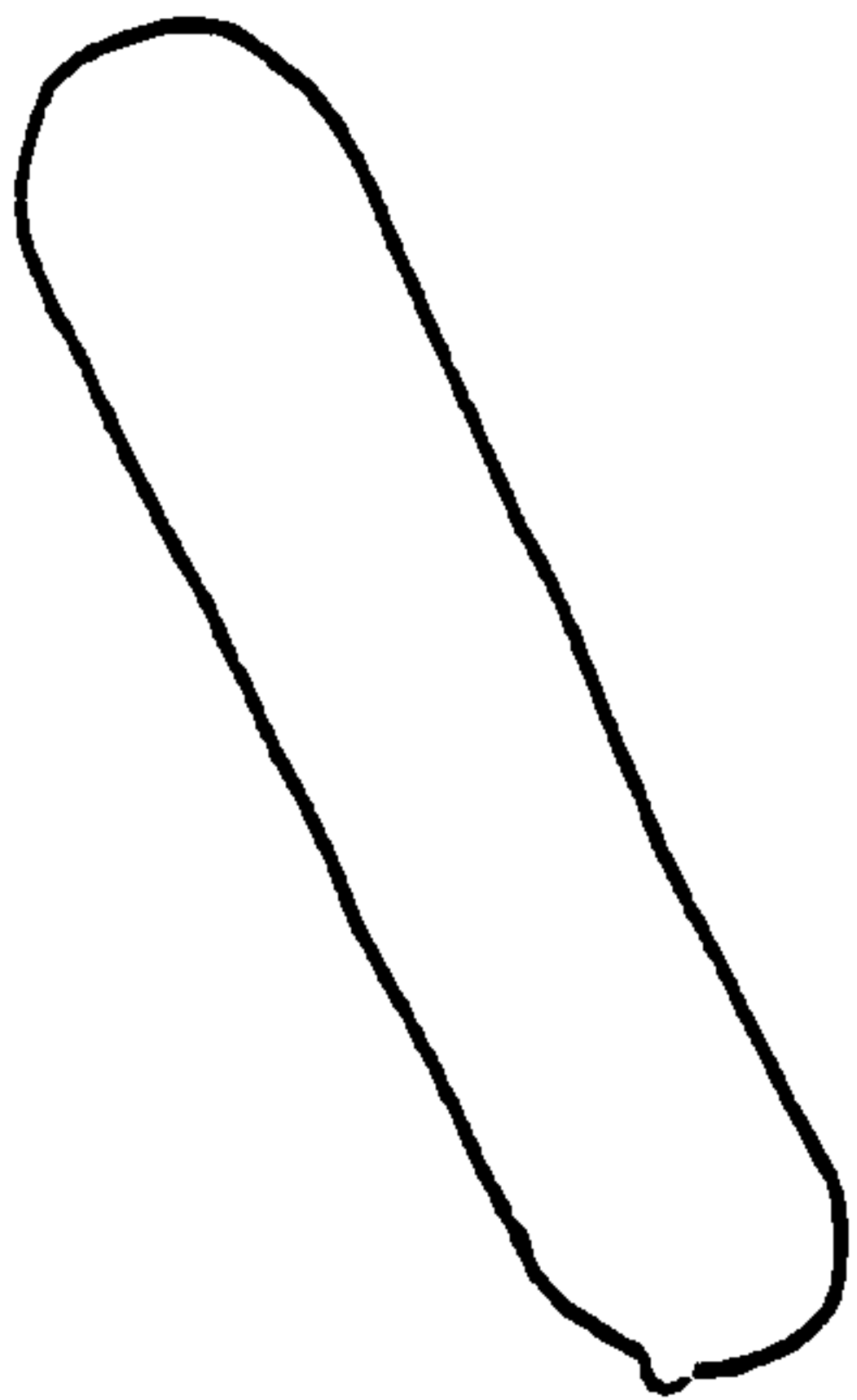
The spore surface can be smooth or ornamented in different ways. The ornamentation is not always easy to see and a high magnification is often necessary to observe this character (at least x400). Lack of ornamentation seen at lower magnification is not proof of the absence of such, and a higher magnification must be used before the state "smooth" is recorded. The ornamentation can either be a roughened surface (punctate, verrucose, spinose) or ridged (reticulate, striate). This observation may be enhanced using a staining reagent such as Melzer's as the mount (see appendix 6.1.1.).



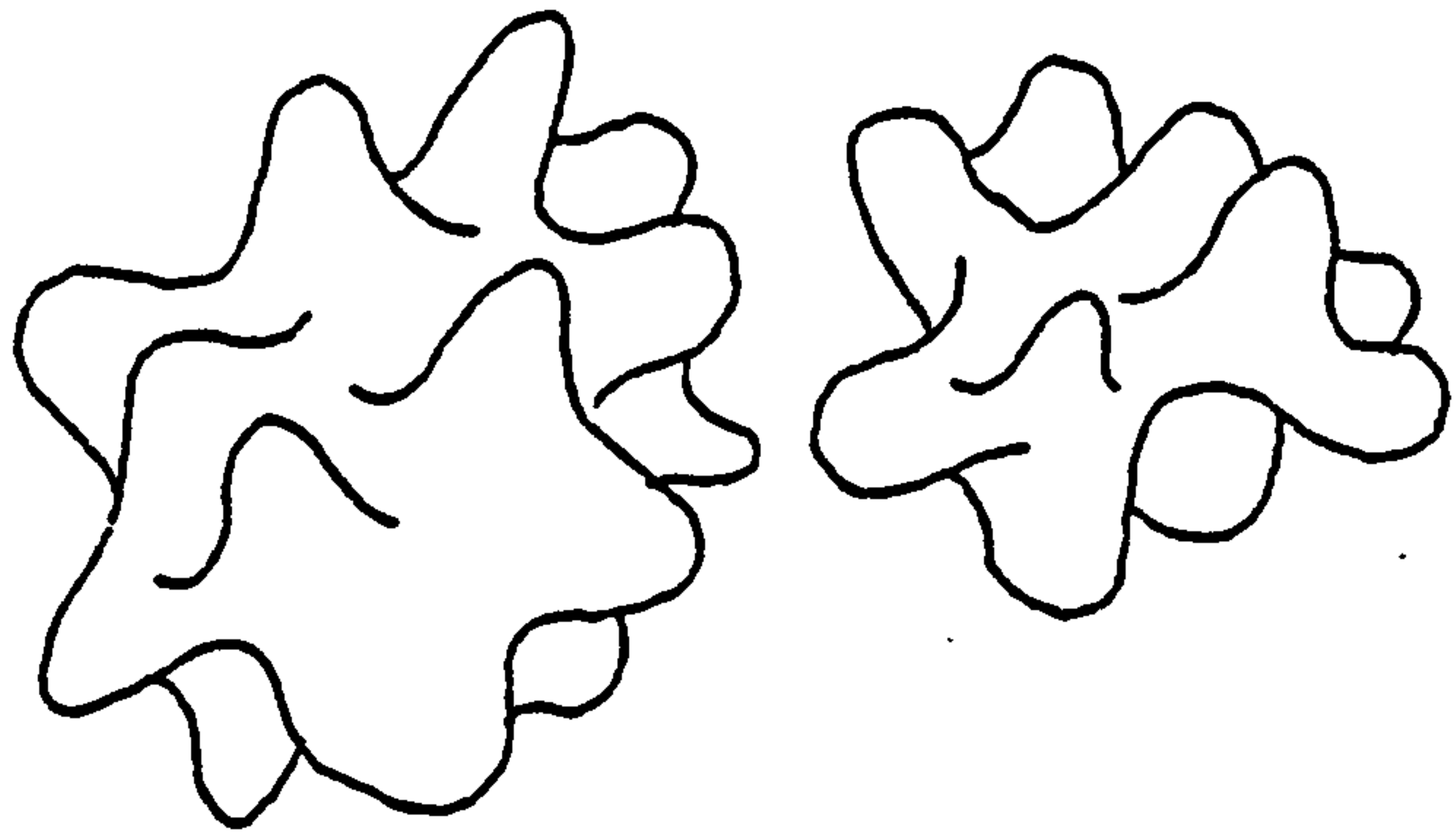
stellate



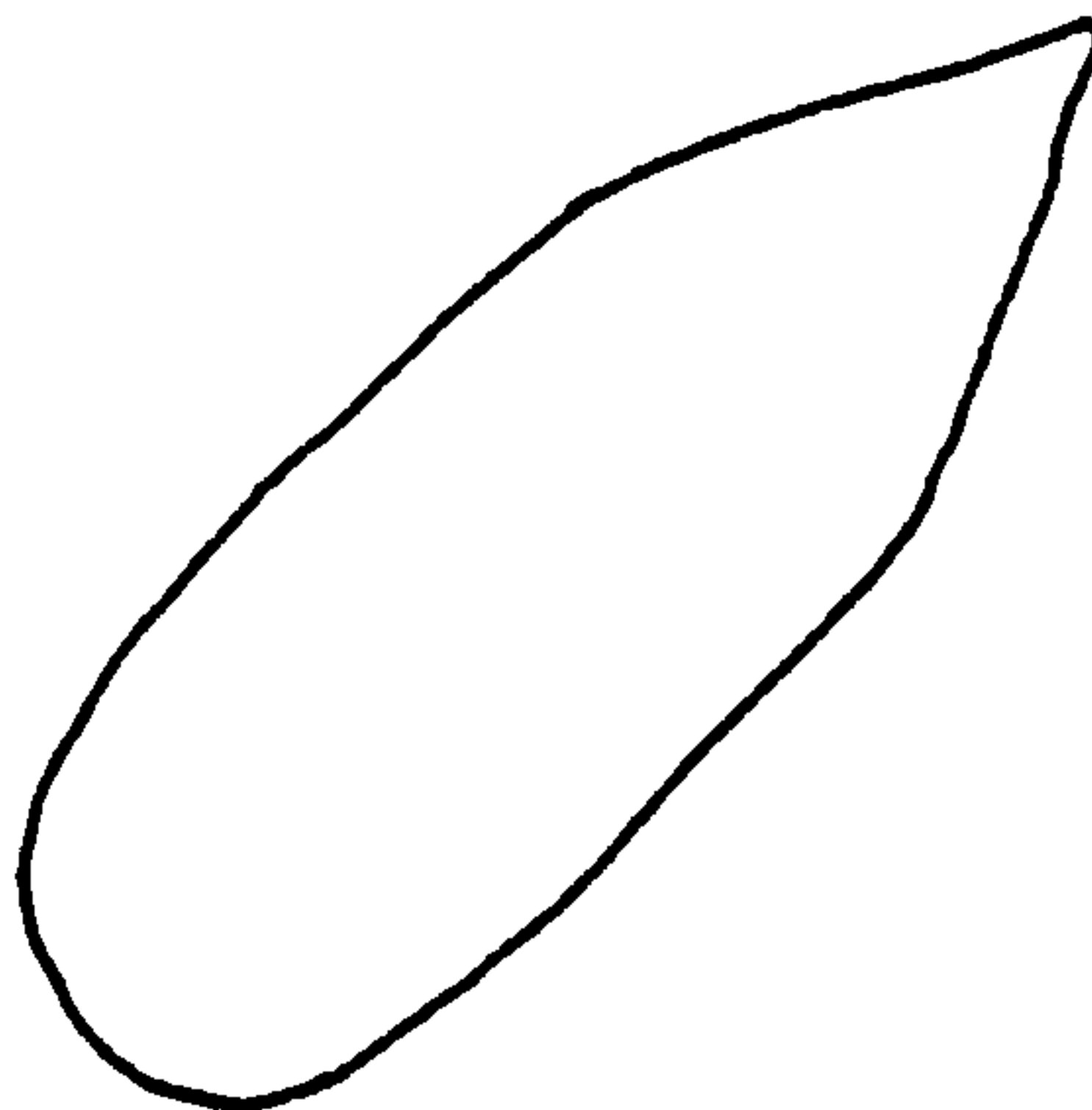
bacilliform



cylindrical



nodulose



naviculate

Figure 9: Various Shapes of Spores

2.2.2.1.1.13. Spore Wall Thickness. (9)

States : - thin
 - thick

In many cases, the spore wall is simple, or so it seems under standard investigating techniques. That is to say it cannot be recognised as double or complex, and for this reason it is usually referred to as being "thin". However, in some dark-spored families, the spore wall often consists of two or three layers which may be readily distinguishable in dilute ammonia, sodium hydroxide or Melzer's reagent (see Appendix 6.1.1.). The innermost layer is called the endosporium and the external layer the episporium. The endosporium is sometimes made up of two layers, while a further external layer may envelope the whole spore (perisporium) (Figure 10). The spore wall is determined as "thick" when it is not seen as simple. This character is readily observed and can be used by a relatively untrained microscopist. Sometimes, however a certain experience is needed to make an unambiguous determination. Because it is impossible to give an unequivocal description which will distinguish a thick from a thin walled spore, a mistake could genuinely be made by the analyst. To safeguard against such a misidentification, a near-miss match identification is

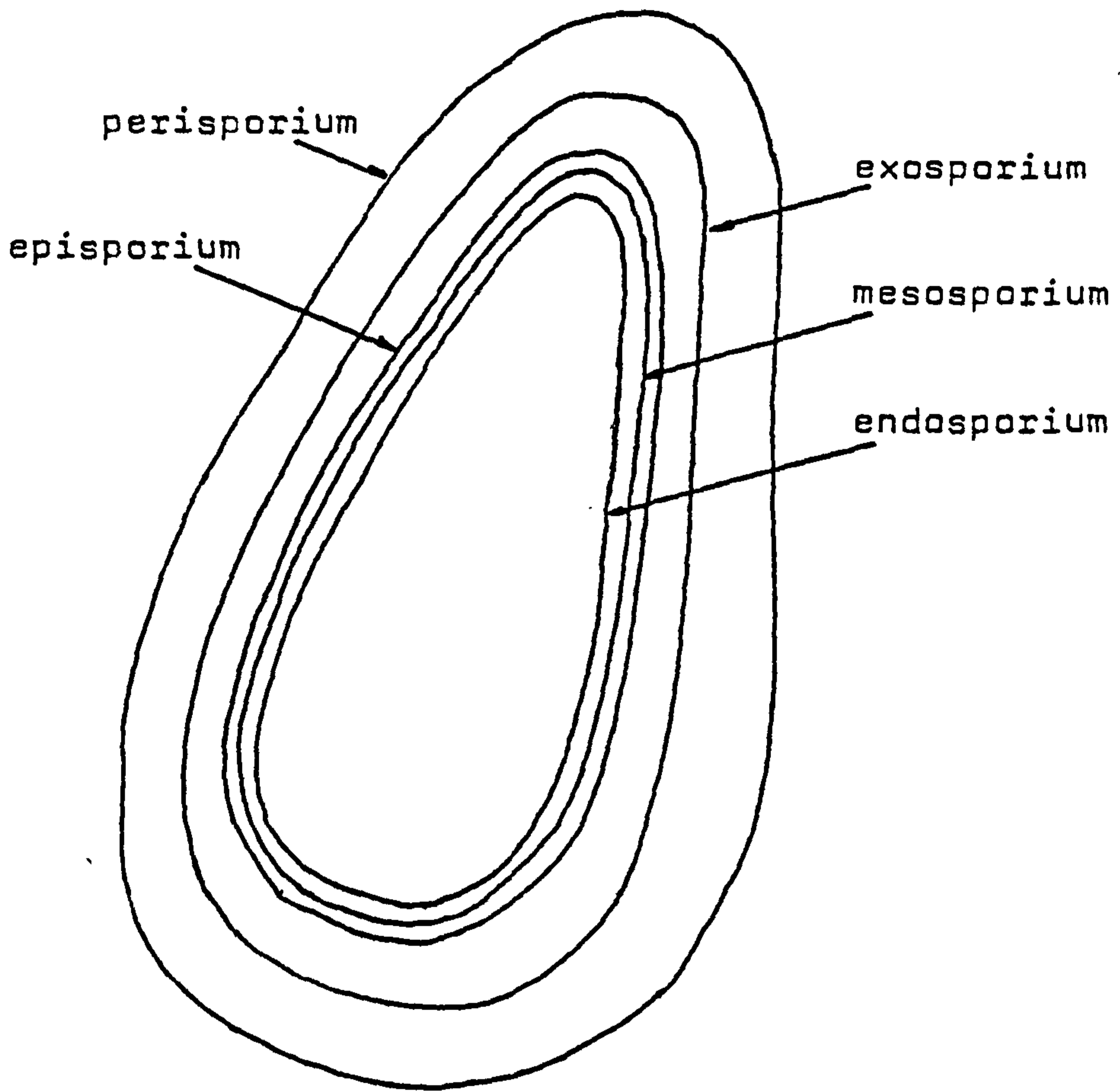


Figure 10: Spore Wall Structure

possible and details are discussed under the appropriate heading.(2.2.2.2.1.)

From the discussion it is obvious that this character is mainly useful in splitting dark spored rather than clear spored species and is therefore of minor diagnostic value.

2.2.2.1.1.14. Germ Pore. (10)

States : - present
 - absent

The spore wall is continuous in many species, or modified at the apex in others. This apical modification may be either a germ pore or a callus, the apex of the spore often appearing as a paler coloured spot flattened or indented, which modifies the curvature of the apical zone. This modification is more or less convex. (Figure 11).

When a callus or germ pore is present, the character "germ pore" shows the state "present". Species whose spores have a continuous wall have the state "absent".

Fayod in 1889 stressed the taxonomic importance of the germ or apical pore. In light coloured spores it is

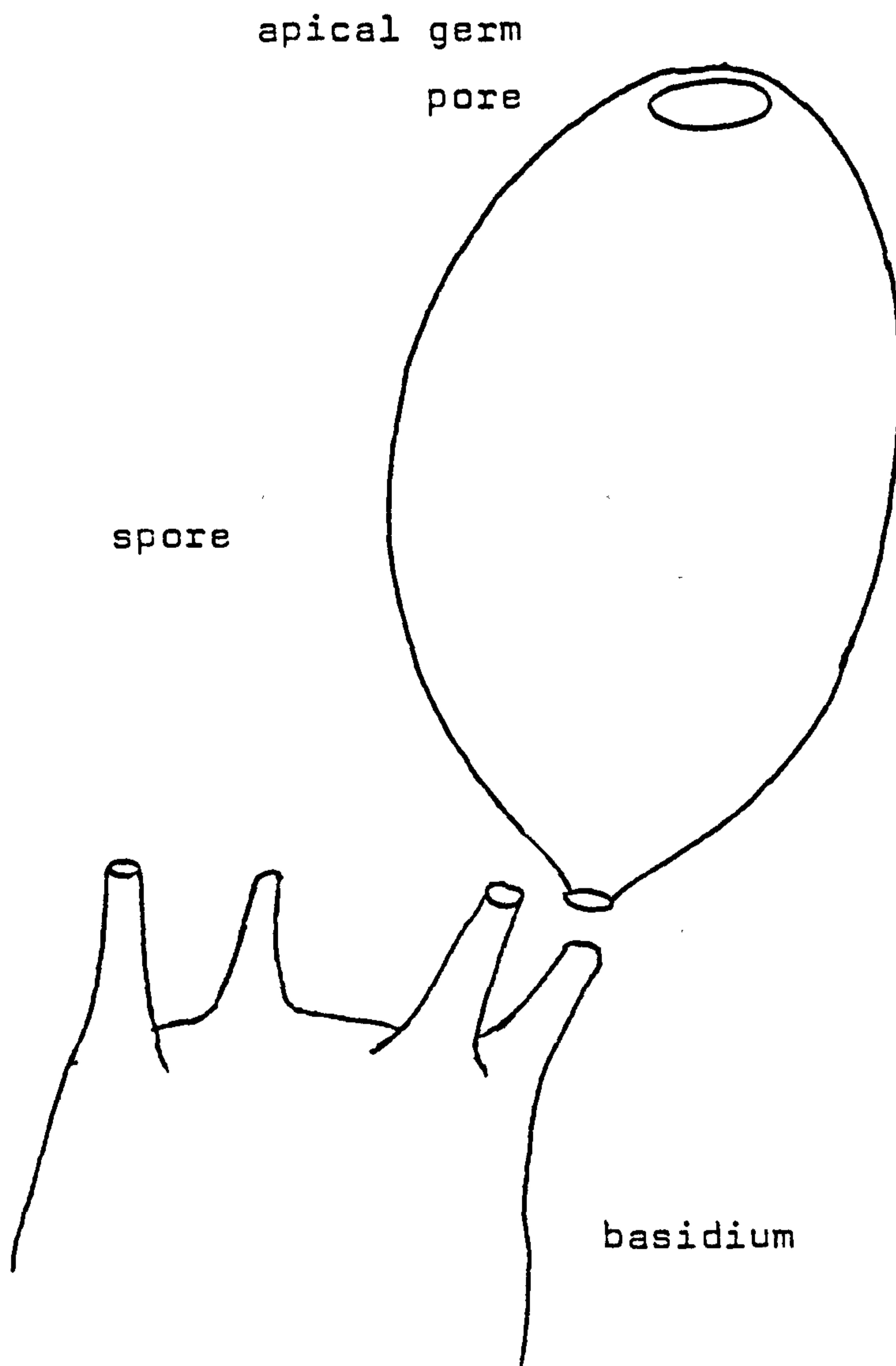


Figure 11: Germ Pore

unfortunately not always easy to observe with standard microscopical techniques unless the spores have been treated with dilute Potassium hydroxide and stained. Singer (1975) recommends an aqueous solution of cresyl blue or aceto carmine as a means of detecting the germ pore.

2.2.2.1.1.15. Clamp Connections. (19)

States : - present
 - absent

Hyphae which make up the trama of the basidiocarp and its hymenial structure are usually divided into shorter units by crosswalls or septa. A specialised type of hyphal branching called "clamp connection" is often associated with septa. It is seemingly involved with the movement of the nuclei within the hyphae. This type of connection is best observed at the base of specialised cells (basidia, cystidia) because any hyphal modification may have this type of connection. Most are small and quite difficult to observe and staining is sometimes advised (e.g. with aqueous cresyl blue). A negative state should not be recorded unless a careful search had been made of several hyphae and, if possible, of several carpophores.

Although this is a relatively difficult feature to observe and might not be used often in emergency toxicology cases, it represents a very useful character which splits genera into groups of species throughout the taxa included in the key. It is therefore retained for its use by more experienced microscopists. (see Figure 5).

2.2.2.1.1.16. The Basidium or the Ascus: Number of Spores.

(20).

States : - 2 spores
 - 4 spores
 - others

The hymenium of fungi is partly made up of specialised reproductive cells.

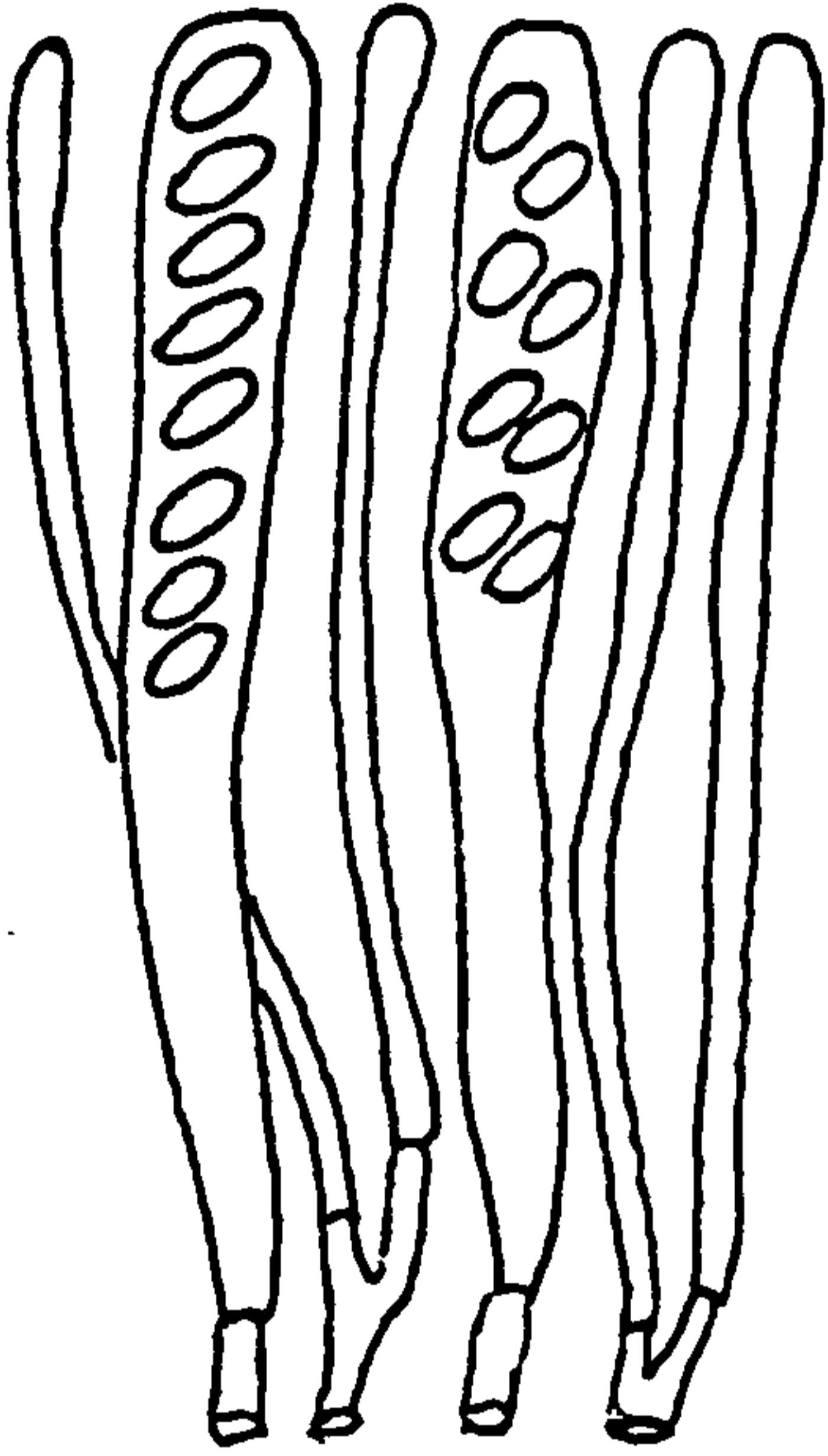
The basidia are single celled, thin walled spore bearing structures of the Basidiomycetes. They usually bear four spores. However a number of species have been described which possess only two spores per basidium and a few rarities have one or three spored basidia. This character is fairly easy to observe and very selective and as such is of limited value as a general diagnostic character. All the taxa but four included in the data bank are Basidiomycetes and only five show a state other than "4 spores".

The remaining four taxa belong to the Ascomycetes, a large mostly microscopical class of fungi of no relevance to this thesis. Their reproductive cells, the asci, are single celled thin walled club shaped spore enclosures. Each ascus usually encloses 8 spores which are shot out when mature. Observation of this microscopical character will therefore be recorded under the state description "others" and will be readily distinguished from that of the Basidiomycetes. (Figure 12; location of basidia can be seen in Figure 5).

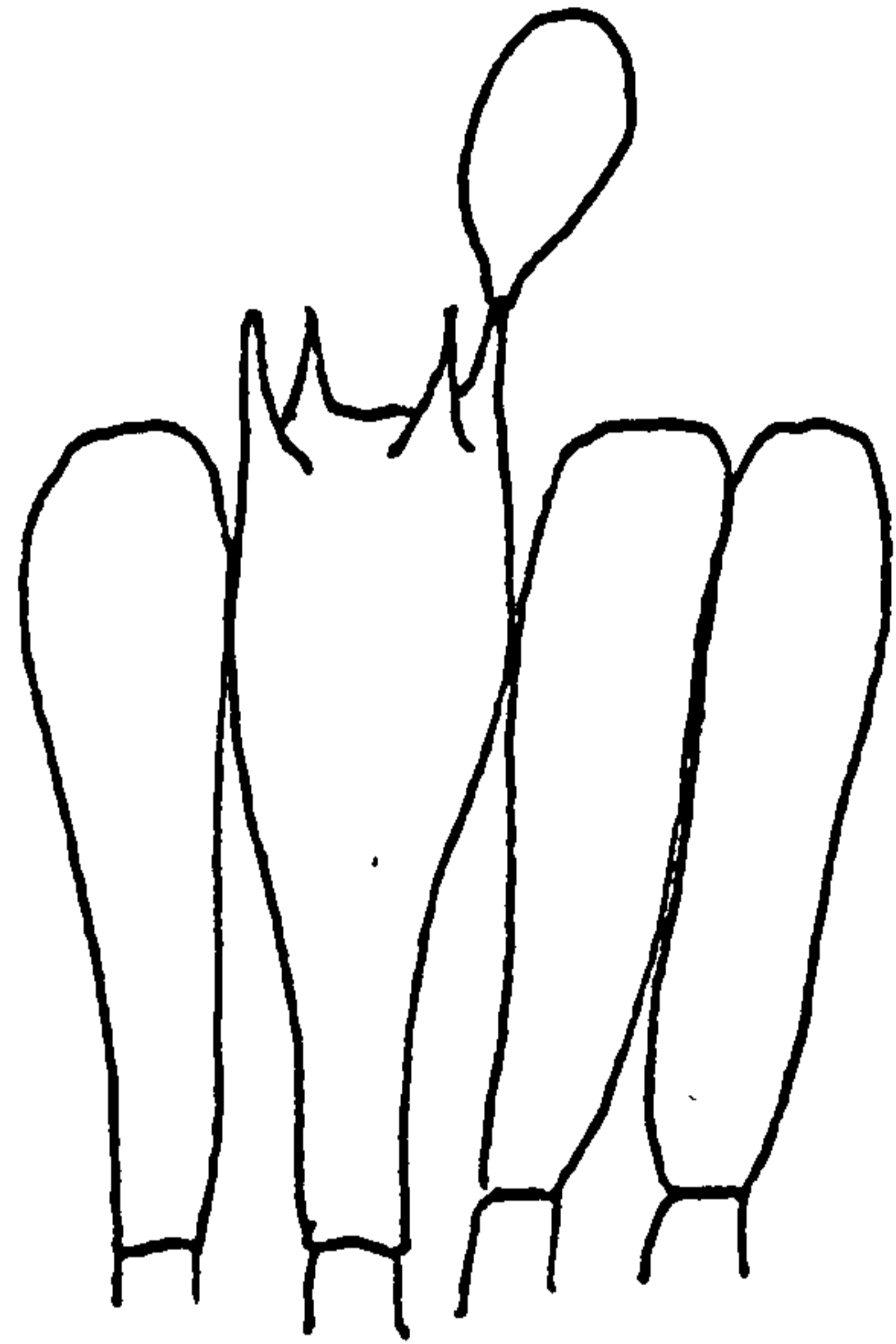
2.2.2.1.1.17. Basidia/Asci Sizes. (21).

- States :
- small (up to 35μ)
 - medium (35 to 55μ)
 - large (over 55μ)

Basidia are normally clavate to broadly clavate when mature and their length is two to four times their width. Only in a few limited cases are the basidia long and narrow, a feature which separates one genus from all the other Agaricales, i.e. the Hygrophoraceae. This helps in the identification of some very common British species of mushrooms and is important enough to warrant its inclusion in the programme although all the species concerned are only mildly poisonous and rarely come to



Asci containing
8 spores



basidium with 4
sterigma and 1 spore

Figure 12: Examples of Ascus and Basidium
Morphology

the attention of toxicological centres.

The dimensions of the asci have no relevance in the identification of the four Ascomycete species included in the key because they all show the same state, i.e. medium. The result is that this character is only of limited interest and remains here only for completeness.

2.2.2.1.1.18. The Cystidia. (22, 23 and 24)

The last microscopical characters used in the identifying process refer to specialised sterile cells forming the hymenium of the basidiocarps : the cystidia.

A cystidium is defined, for our purpose, as a differentiated sterile terminal cell of the hyphae in the hymenium. This is a restrictive definition because it concerns only the hymenial surface. Strictly speaking, other cystidial types can be found on the cuticle or the stem of the basidiocarp. The latter are not used in the present key and therefore not included in the definition. The functions of cystidia are unknown, but their morphology is usually quite distinctive and varied and makes them a useful character for identification.

Cystidia can be further defined by their position on the hymenial surface. They can be found on the face of the

gills only (as seen in the transverse section of a gill, Figure 5) where they are referred to as facial cystidia or pleurocystidia, or, they can be found only on the margin of the gill (seen on the portion of the gill sectioned as described in Figure 2) i.e. marginal cystidia or cheilocystidia. Some species have both types of cystidia and some have no cystidia at all. This can have a great diagnostic significance when mushroom samples (rather than for example stomach content) are available for identification.

For the above reasons, marginal and facial cystidia are used as two separate characters, but both present the same states and are discussed together.

2.2.2.1.1.18.1. Marginal and Facial Cystidia.(22 and 23)

States : - present
 - absent

Substates : when present: - cystidia sizes :
 - small (less than 35 μm)
 - medium (35 - 55 μm)
 - large (over 55 μm)

Substates (continued) : - Cystidia Shapes :

- cylindrical
- lageniform
- ampulliform
- utriiform
- metuloid
- uncertain

Cystidia may sometimes be quite remarkable and of significant diagnostic value, such as for example in the genus Inocybe (which has over 30 poisonous species), although they clearly need a certain experience of microscopy to be observed in a number of cases.

To maximise the use of this character, when cystidia are observed on either the face or the margin of the gills, it must then be further subdivided into subcharacters represented by the sizes and shapes of the cystidia. The various states of these subcharacters categorise large groups of poisonous fungi and are therefore important and useful diagnostic features.

2.2.2.1.1.18.2. Cystidia Sizes.

The cystidia sizes are continuous states and suffer the same drawbacks as discussed previously with the spore

sizes. It was found useful to divide sizes into three groups of delimited lengths defined as small (under $35\mu\text{m}$), medium ($35 - 55\mu\text{m}$) and large (over $55\mu\text{m}$). The majority of cystidia are of medium size, but when small or large cystidia are encountered, they quite obviously fall into their respective categories. If the character "cystidia" has been observed, it is an easy and routine exercise to measure their sizes. The problem usually is to locate the character and not to define it.

2.2.2.1.1.18.3. Cystidia Shapes.

Shapes of cystidia are numerous and it is a major difficulty to categorise them into a limited number of well-defined characteristic shapes. Five groups are defined as follows, but might be better described by the figures which relate to each state:

- a) cylindrical cystidia essentially possess parallel sides in which the width remains roughly the same along the entire length of the cell. This includes filiform, cylindrical and clavate cystidia (Figure 13).
- b) lageniform cystidia are cystidia having a swollen base and an apical part which tapers into a sort of beak. It includes acuminate, subulate, fusiform,



filiform



cylindrical



aciculate



claviform

Figure 13: Cylindrical Cystidia

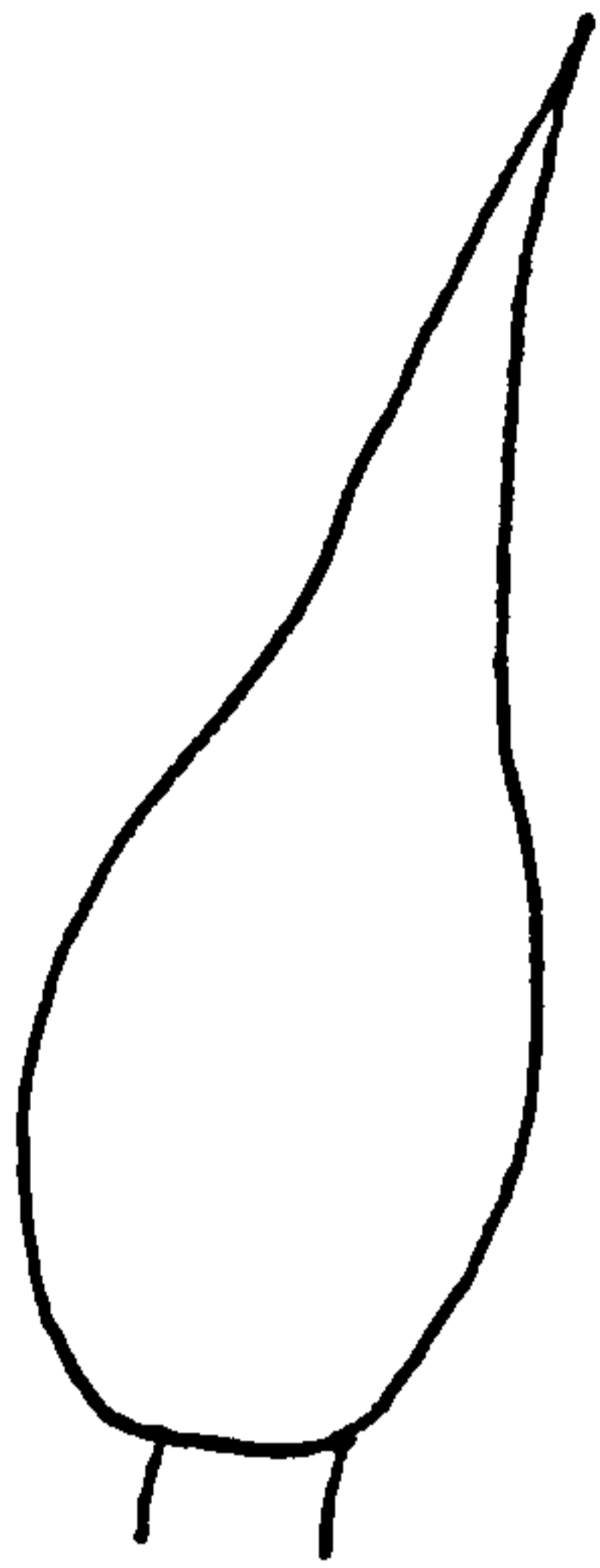
lanceolate, obclavate and digitate cystidia (Figure 14).

c) ampulliform cystidia are broad, almost as broad as they are long; they are ventricose to sub-ventricose, napiform, vesiculate and pyriform (Figure 15)

d) utriform includes all cystidia which possess a constriction in their medium part. Some liberties have been taken here with the original mycological definition of utriform, i.e. "cystidia which have a slight constriction below a large, round head, like a bladder: therefore bladder-shaped" (from Largent et al. 1977). Sphaeropedunculate, capitulate, lecythiform, etc cystidia belong to that category. (Figure 16).

e) metuloid. Although it is included in the substate "shape of cystidia", the last group "metuloid" does not define a shape as such, but a special differentiated apex of some cystidia. It is always found in association with one of the other shape states, e.g. ampulliform with metuloid or lageniform with metuloid, etc. The cystidium with a metuloid has a special thick walled apex of variable shape containing amorphous or crystalline material (Figure 17).

The information content of these characters is very important although quite difficult to observe and define



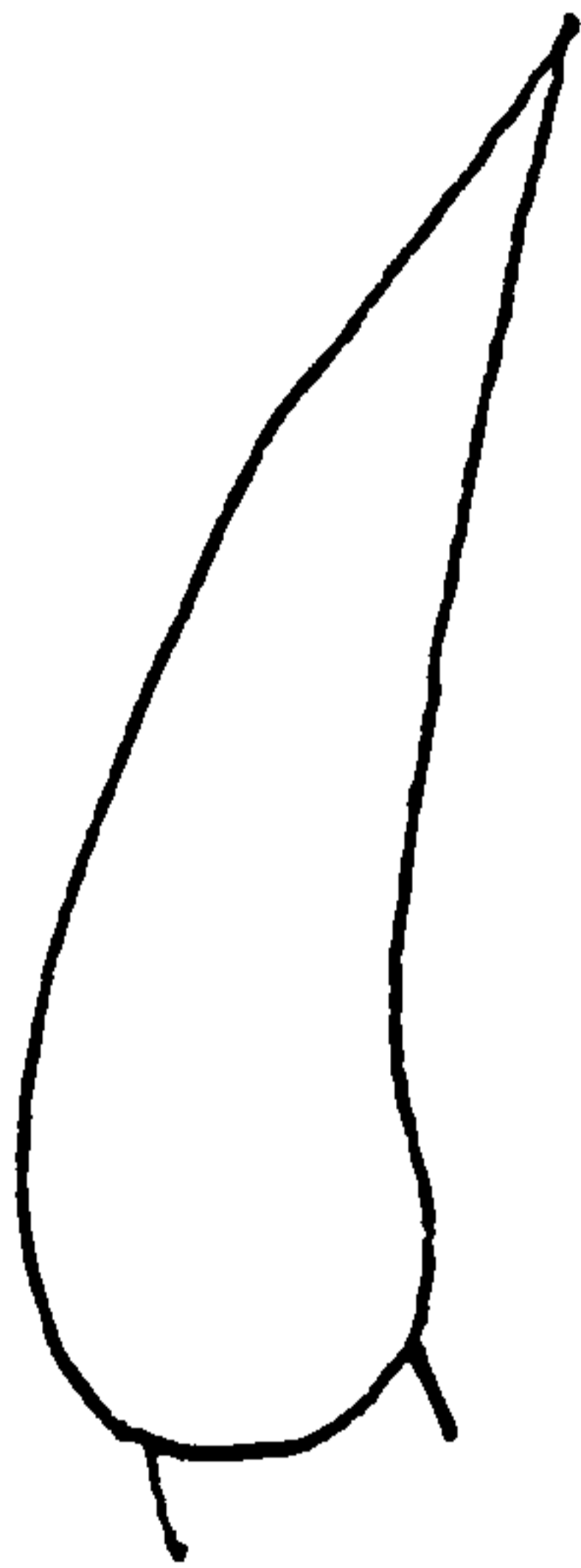
obclavate



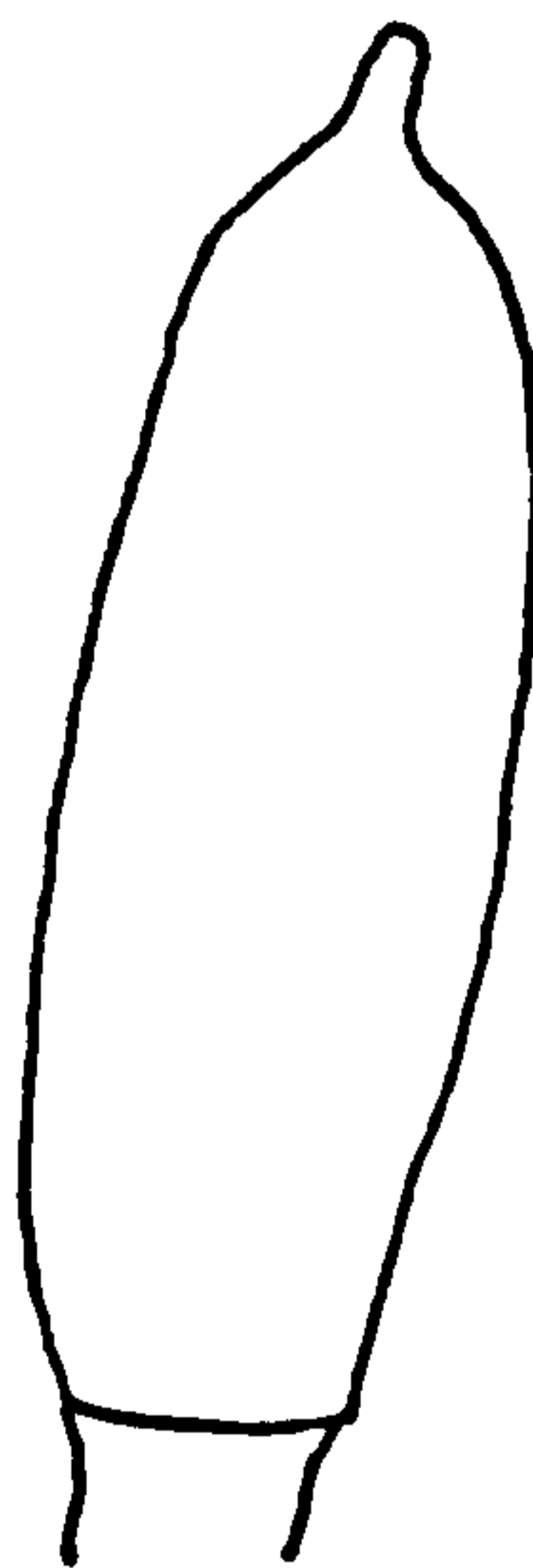
subulate



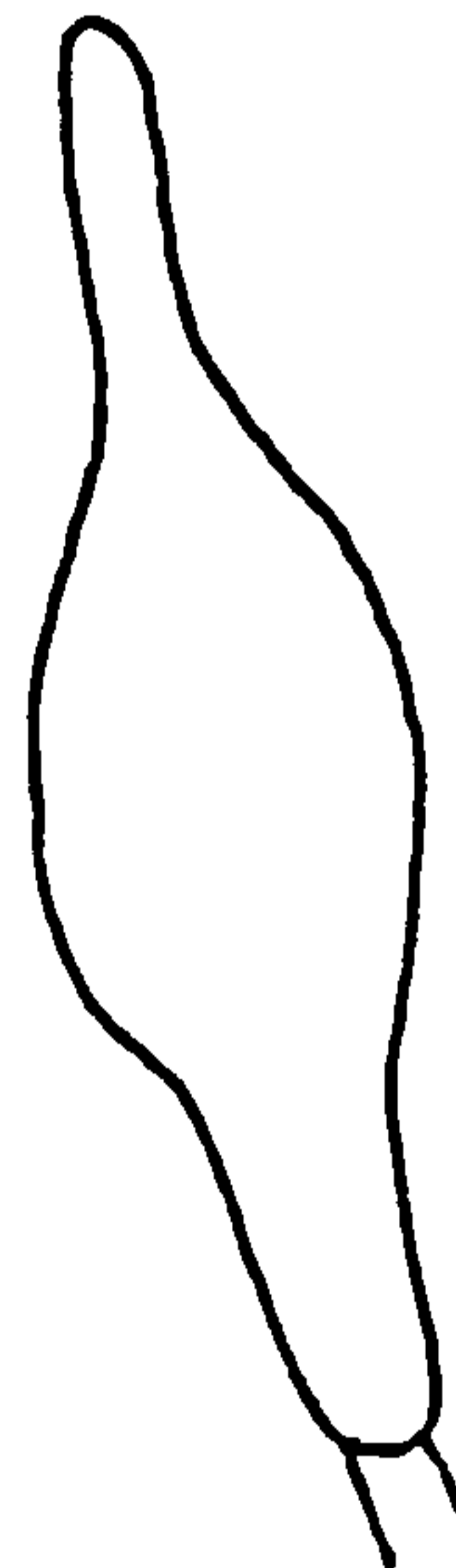
fusiform



lanceolate

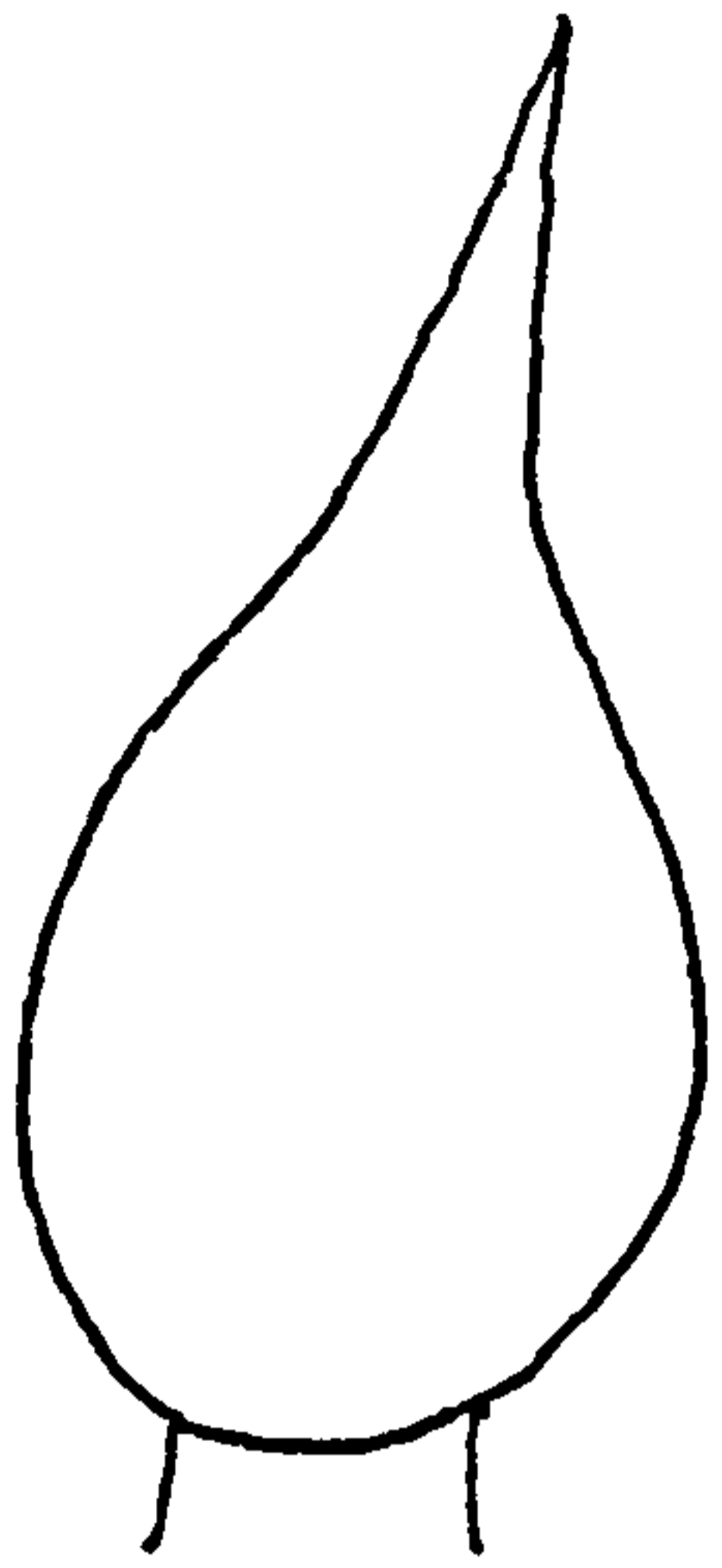


mucronate

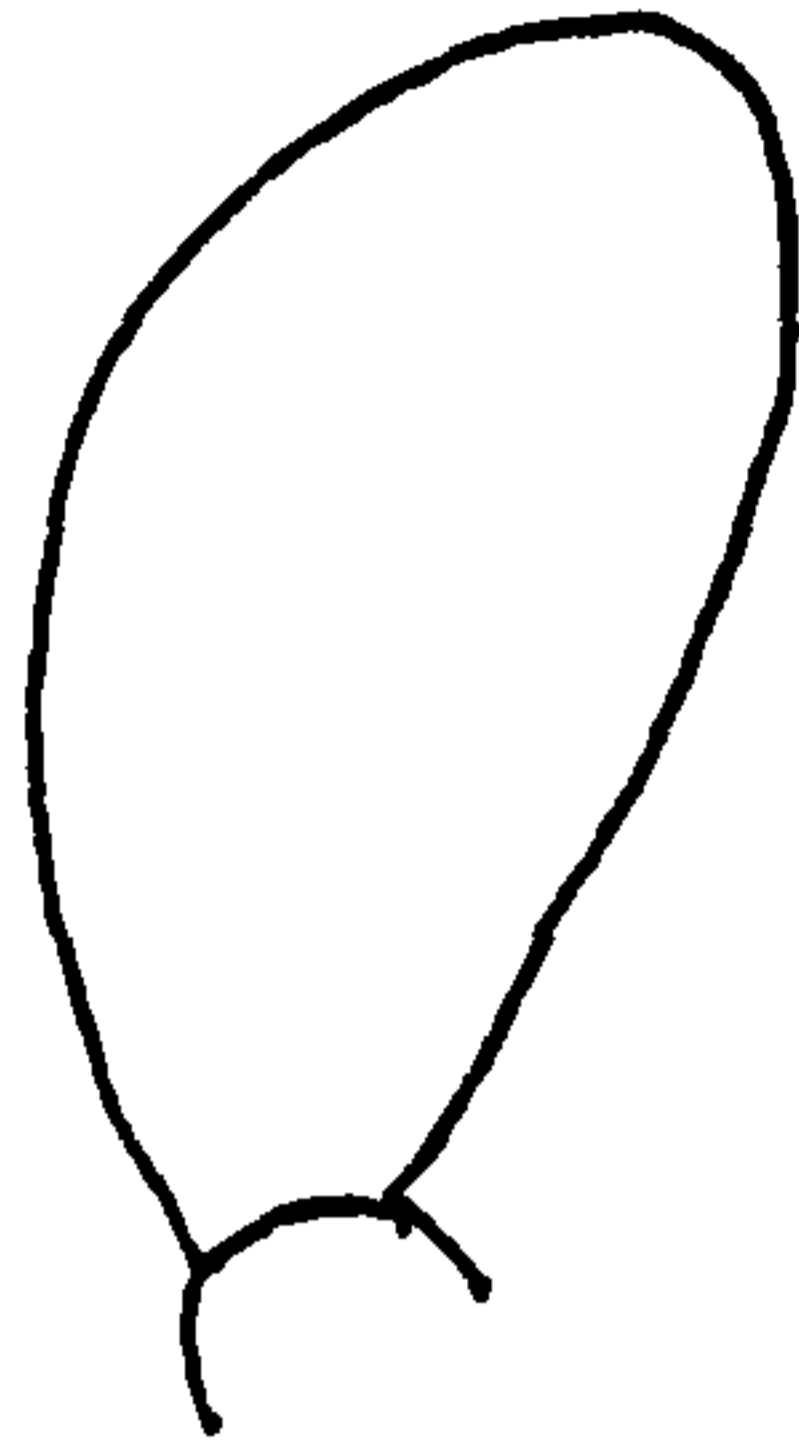


digitate

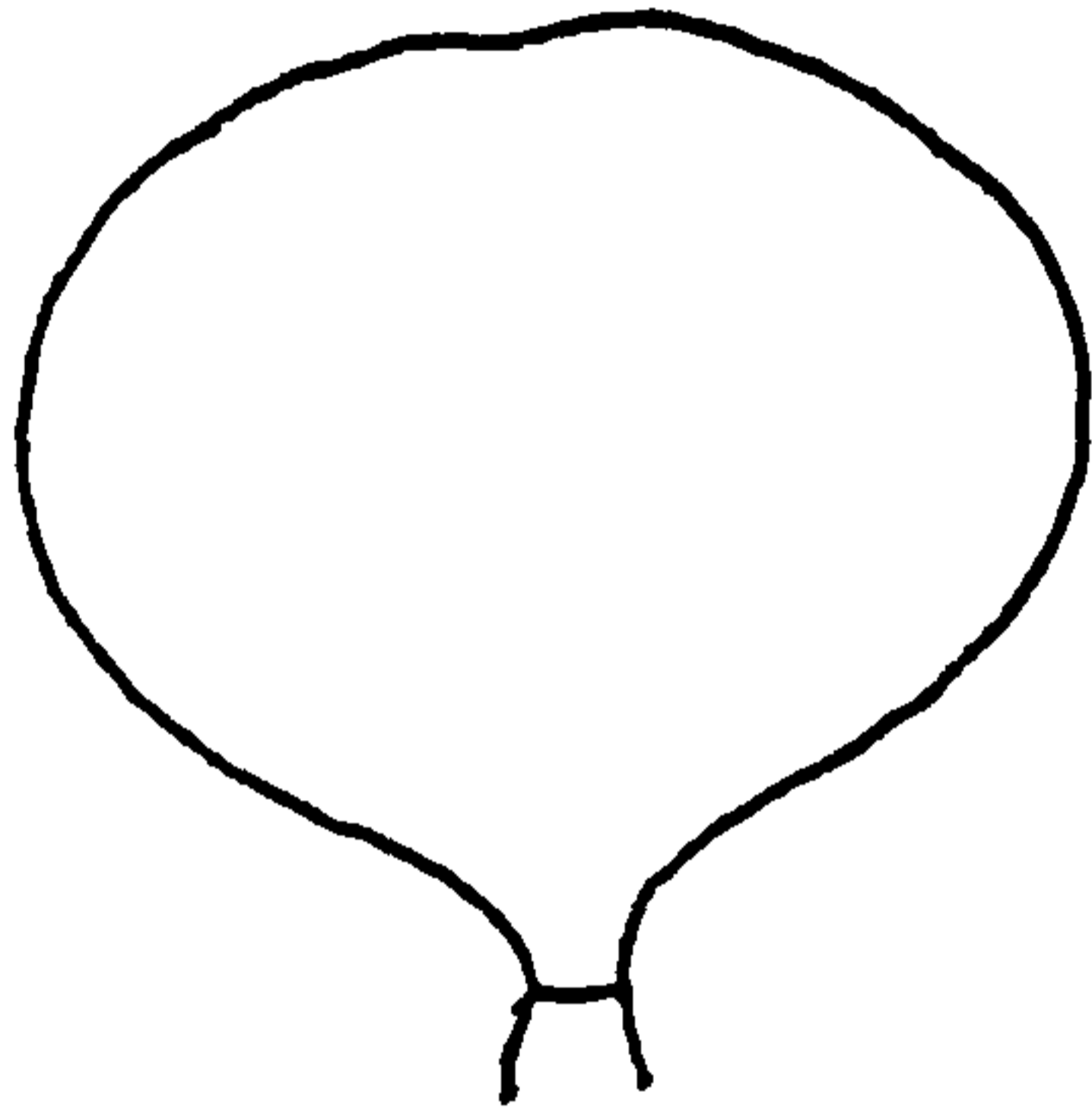
Figure 14: Lageniform Cystidia



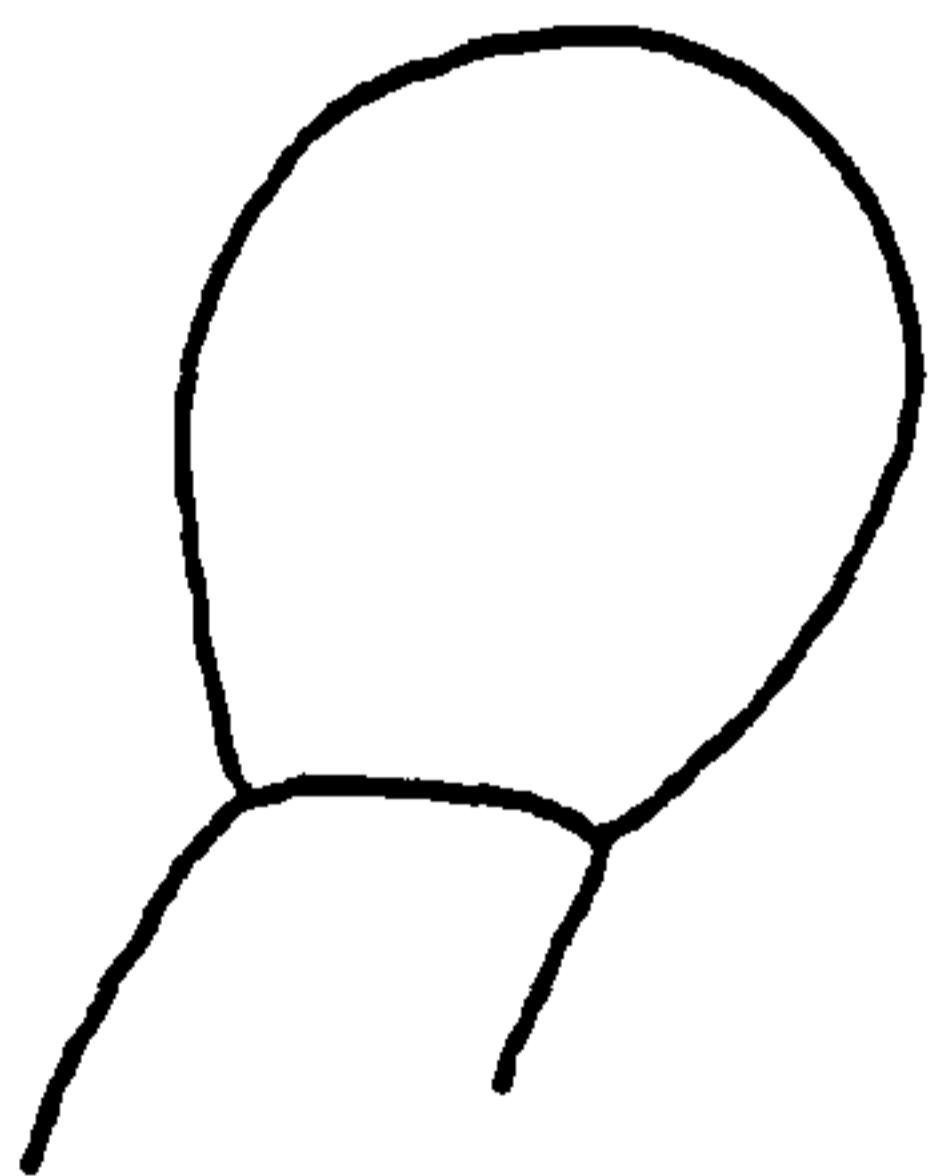
ventricose-rostrate



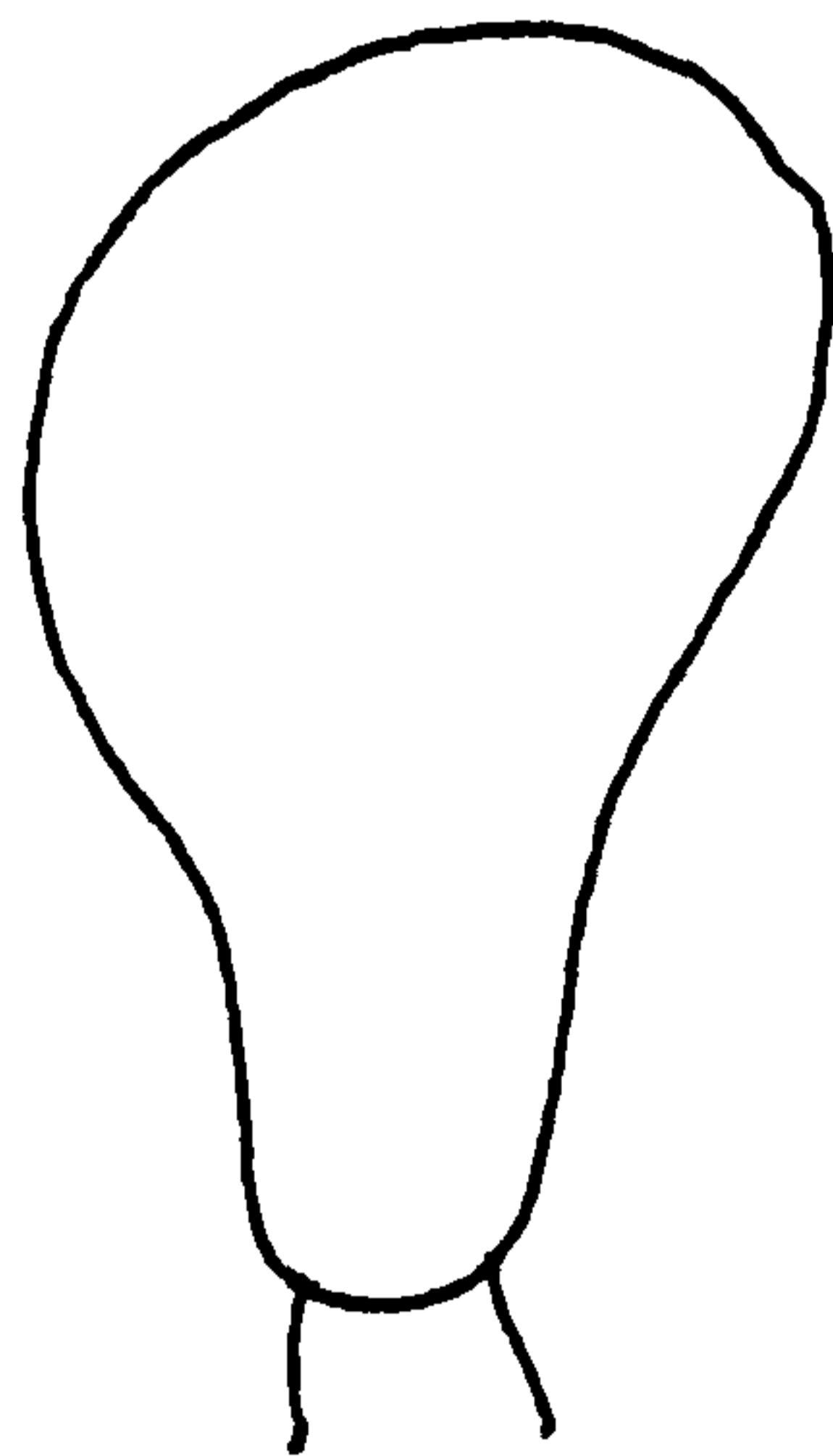
napiform



turbinate

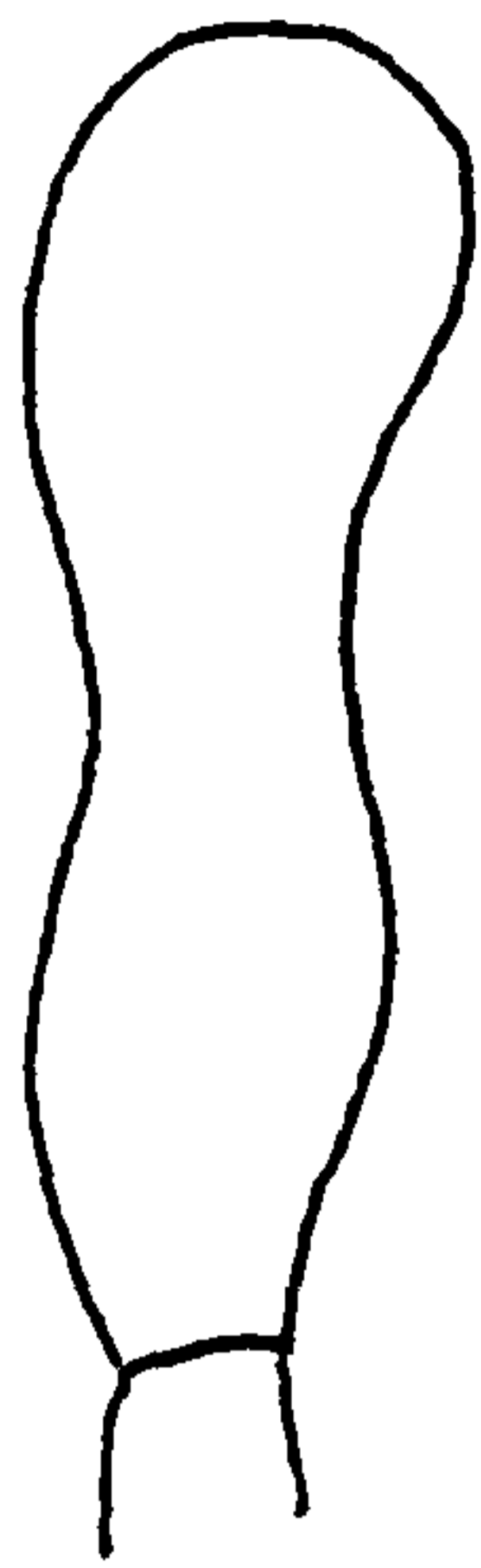


vesiculate



pyriform

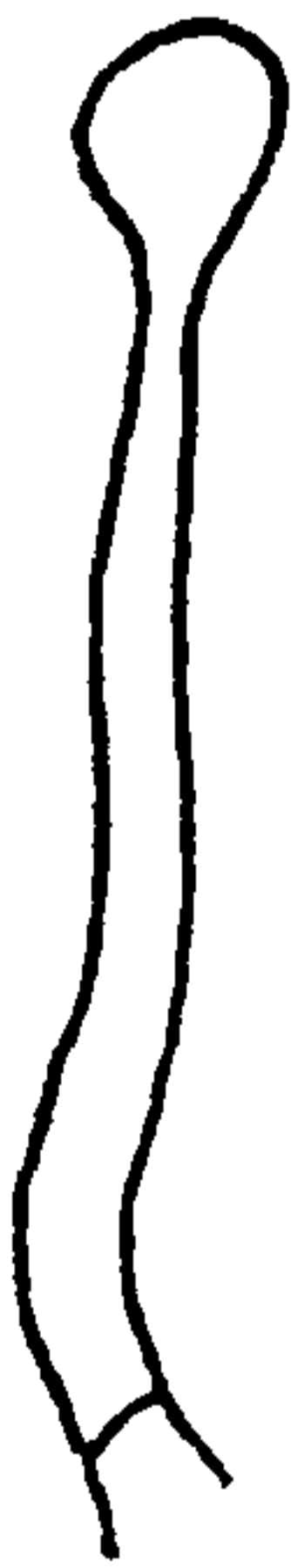
Figure 15: Ampulliform Cystidia



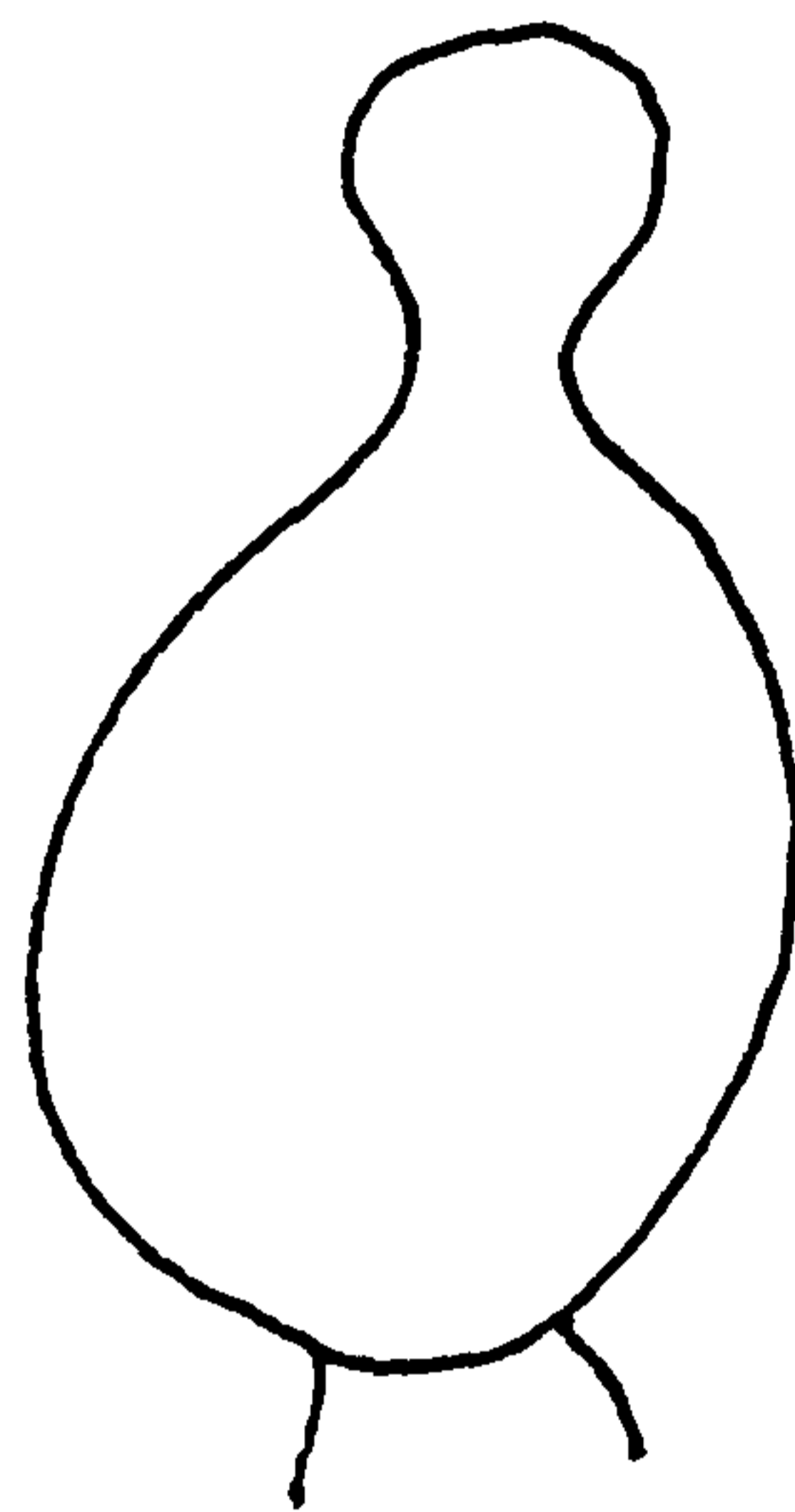
sphaeropedunculate



capitulate



tibiiform

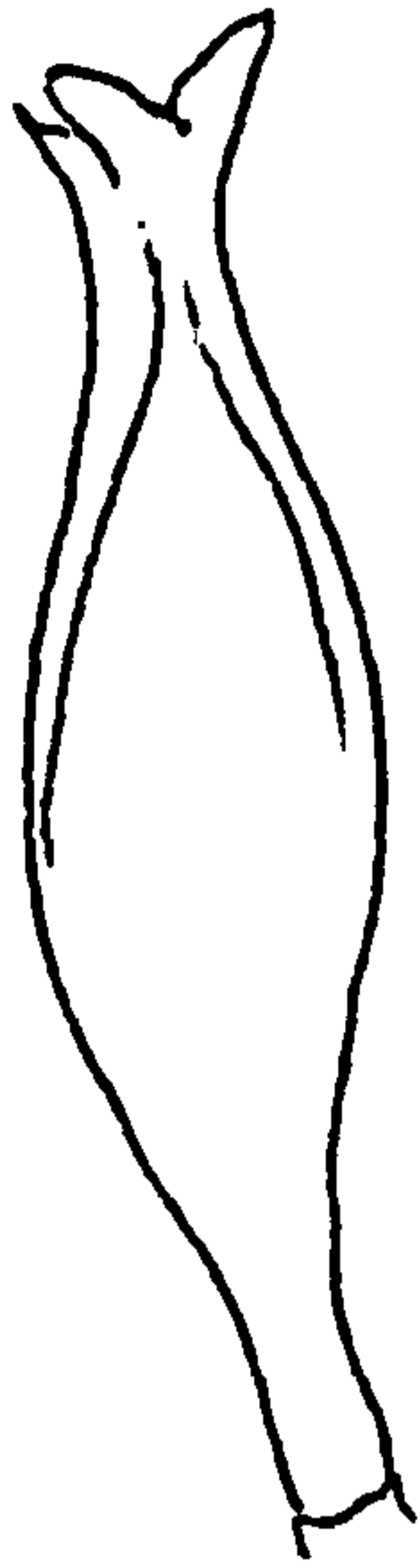


lecythiform

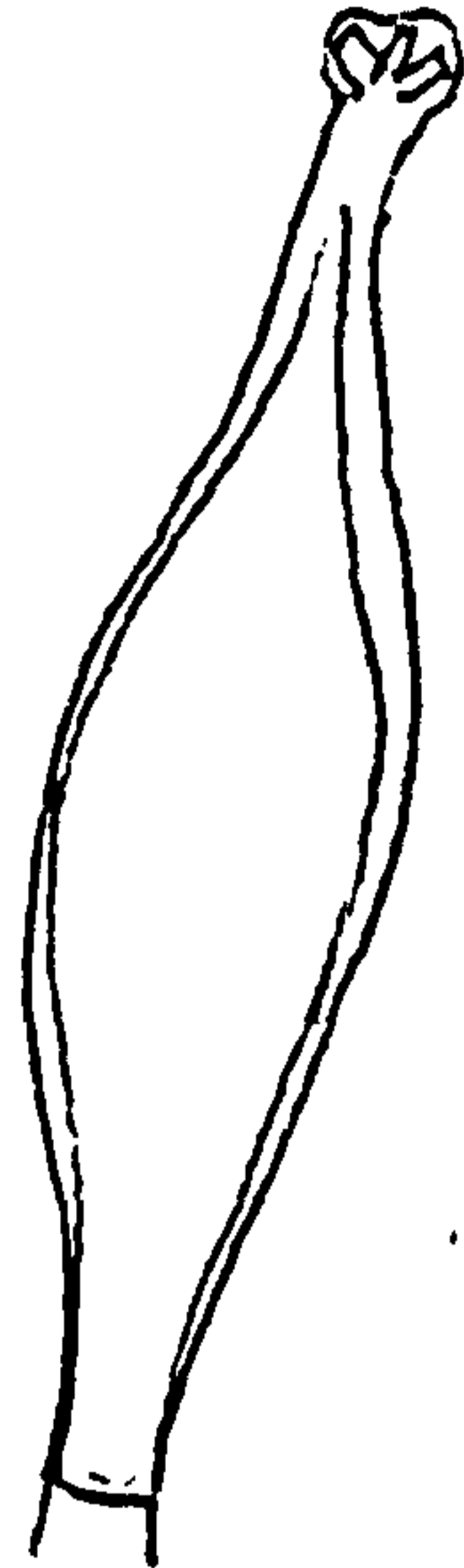


strangulated

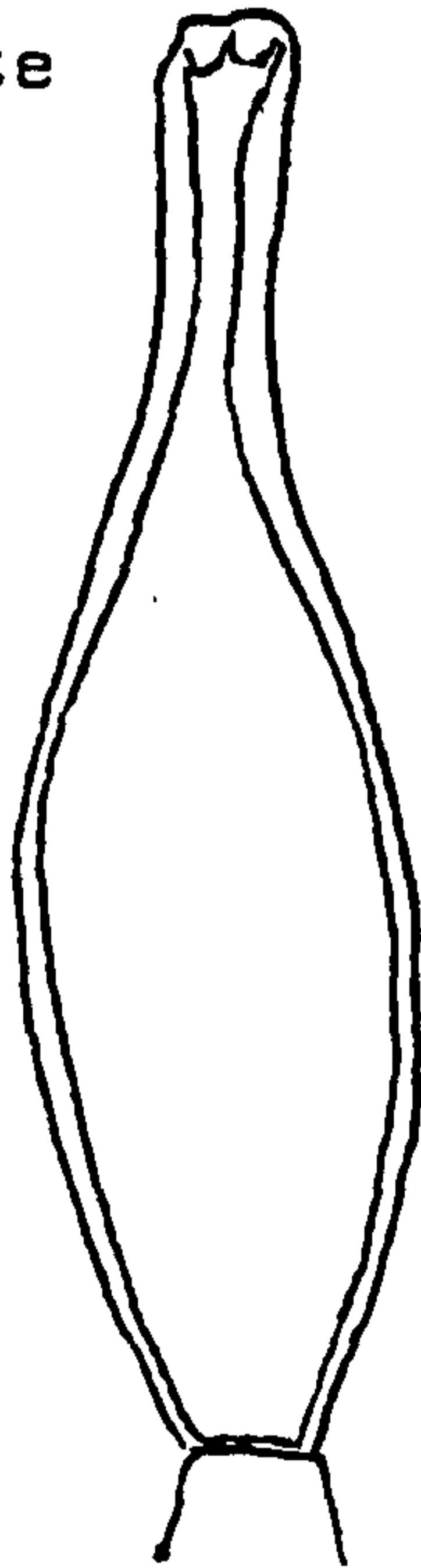
Figure 16: Utriform Cystidia



cornuate



fusoid metuloid



obclavate metuloid

Figure 17: Cystidia with Metuloid.

in a number of cases. An uncertain category has been included in the key for users who are not sure of their identification. On the other hand, a state such as metuloid is very easy to identify and provides an important split in the genus Inocybe (over 30 poisonous species).

The importance of this character seems to warrant a little training on the part of people interested in using the present identification scheme.

2.2.2.1.1.18.4. Chrysocystidia. (24)

States : - present
 - absent

The last microscopical character introduced in the diagnostic key is in fact a microchemical character. Some cystidia, in the presence of strong aqueous alkali solutions (e.g. 50 % Ammonia) stain yellow to golden. They are specific to a few genera of interest to the mycotoxicologist (Stropharia, Hypholoma). These cystidia are called chrysocystidia and show the states "present" or "absent". No distinction is made here between facial and marginal cystidia because they show the same topological distribution for the genera of interest (i.e. they are found on both the margin and the face of the gills).

This completes the set of anatomical characters included in the programmed key. They are necessary if an attempt to identify a specimen down to species level is required, but in some cases they may not be sufficient, where closely related species are involved and further detailed microscopic examinations by a trained mycologist are necessary. For example, closely related Tricholoma are not specifically identified by the present set of characters.

2.2.2.1.2. Ecological Characters.

Three characters have been considered useful here because they are likely to be known at the time of an emergency. They are available to whoever uses the key and, above all, may carry very valuable information content. These are geographical distribution, habitat and time of growth.

2.2.2.1.2.1. Geographical Distribution. (3)

- States :
- North temperate
 - South temperate
 - subtropical
 - tropical
 - English
 - Scottish

A general state definition is found to be practical and theoretically acceptable. It splits fungal distribution into four large climatic regions defined as North temperate, South temperate, subtropical and tropical.

A choice of alternative states was available. The geographical distribution could have been described by groups of countries such as Siberian, Northern European, Mediterranean, North American, Australian, etc, which theoretically would have had high diagnostic value and therefore given a very useful split in the population included in the data bank. This was found to be unpractical because the distribution of mushrooms is very well documented in Europe, but that is certainly not the case for other areas of the world. Major advances have been made in this respect during the last decade and another five to ten years of data collection may prove sufficient to consider using such state definitions.

The North temperate area includes most of Europe, most of North America and a large part of Asia. Since the key was constructed in Britain and distribution surveys were carried out in Scotland a more selective division has been introduced by taking into consideration the distribution of fungi in England and Scotland, two countries which show marked differences in their fungal

flora. Whereas England has a flora comparable to most Central and Western European countries, Scotland has one comparable to that of Northern European countries (Scandinavia, Iceland and Alpine areas). This is a very useful split at the European level where the data is fairly complete and comprehensive. The states "England" and "Scotland" could easily be replaced by, for example, "Germany" and "Sweden" or "Holland" and "Scandinavia", etc. It is obvious that this character will often show multiple states for given species i.e. widespread European species show states "North temperate", "English" and "Scottish". North temperate species which are neither English nor Scottish are in most cases North American and therefore are split by the fact that multiple answers are used. This is also reflected in the data retrieved when an identification is attempted (see 2.2.1.7.)

South temperate species are mostly Australian, New Zealand and South American species, but the data is incomplete and needs to be expanded if a comprehensive coverage is required.

Subtropical species are found mainly in South Mediterranean and Central American countries, the data for other areas being scattered and largely incomplete. When the fungal distribution is better known in

subtropical areas, this state will have to be divided into two separate states i.e. "North subtropical" and "South subtropical".

Tropical fungal floras seem to be better known than the subtropical floras due to a number of surveys by eminent mycologists (Heim, Pegler, Heinemann, etc). This state covers a large part of Africa, South Asia, Indonesia and parts of South America.

It is obvious that an overlap between climatic areas is inevitable and that this character is continuous. Climatic conditions can change from one year to another and modify these areas to varying extents. This means that a subtropical mushroom may, for instance, be recorded, exceptionally, from the South of England. The likelihood of it being involved in a case of poisoning in the South of England is however very remote due to the rarity of its occurrence. Alternatively, a mass poisoning is likely to occur in a high frequency area for the given species. This guarantees the validity of this character and its selective strength.

Apart from data about Scotland, where personal collections and research supported and complemented literature data (see Title 4.2.), all the geographical data has been gathered from the mycological literature.

2.2.2.1.2.2. Habitat. (4)

- States :
- ground in coniferous woods
 - ground in hardwoods
 - ground in grass
 - ground in dung
 - on stumps
 - others

All fungi, by their very nature, have to grow in association with other living, or once living, organisms as parasites, symbionts or saprophytes. The host-fungi association is often one of a selective nature and even sometimes of a specific nature (that is, a certain fungal species will necessarily be associated with a single specific host).

Many fungi are less specialised and a whole range of associations, from non-selective to specific can be found. It is useful to divide the host type into six selective groups representing the main types of association encountered with the higher fungi.

Mushrooms living in mycorrhizal association with trees are divided into coniferous (or softwood) and hardwood association. Multiple states are permitted and mushrooms found growing in both types (mixed) of woods or

in deciduous woods show both states of character. Most deadly poisonous mushrooms belong to one or other, or both the above categories.

Mushrooms growing in grasslands are quite common and a number of them are upsetting when ingested and provoke some gastrointestinal disturbances to the eater. Some hallucinogenic fungi are also found in this section although most of them grow on dung, the next specialised habitat considered in the key. Coprophilous fungi, as a group, are generally small and, because of their rather unappetising habitat are unlikely to find their way onto the plate of the mycophagist. Intoxications with coprophilous fungi are likely to arise from people collecting fungi for their hallucinogenic properties, since many hallucinogenic mushrooms listed in the data bank grow on dung. This fact warrants the use of this specific type of habitat as a separate state.

Other saprohitic fungi grow on dead wood or stumps. Lignicolous species are numerous and the most common British poisonous mushroom grows on such a specialised habitat (i.e. Hypholoma fasciculare) Apart from the example cited only a few poisonous species are to be found in this category and a number are not large or fleshy enough and their toxicity, if any, is unknown.

This makes this state fairly restrictive but still useful with regards to the frequency of occurrence of the species found here.

The last state, "others", is, as it implies, a mixed bag containing various other types of specialised habitat such as burnt patches, sand-dunes, heathland, marshes, other fungi, etc. They are all grouped together because only a few species are associated with them or, more likely, only very few toxic or suspect species are known to grow in connection with these habitats.

There is one major drawback when using this character in that the division between one state and another is not always clear or can even be wrong. For example, a mushroom growing in grass, five meters outside a larch forest, may in fact be associated with the larches and not with the grass (e.g. Suillus grevillei or larch bolete). Further, a dung may not be seen as such after rain, and a pasture may be seen as rich "grass" rather than "dung" in grass. Equally, a change in vegetational history of an area may not be completely reflected in the fungal distribution found there. This phenomenon is known and has been used to study the ecological history of given plant communities. Fungi growing in association with specific hosts have been found to grow decades and centuries after

the host had disappeared, thereby adapting to a new environment (Watling, 1978).

Such examples could fill pages and indicate that, in fact, the true habitat of a given mushroom could be genuinely mistaken, and hence defeat the whole purpose of identification unless provision for such a mistake could be introduced into the programme.

One such safeguard has been devised, in the key, which allows an identification despite incorrect observation of the state of the present character. That safeguard is discussed elsewhere under the heading "Near-miss match identification" (2.2.2.2.1.).

The main advantage of the character "habitat" is that, when accurately defined, it splits toxic and hallucinogenic fungi into large groups of relatively equivalent importance, i.e. the information has an important diagnostic impact. Its usefulness is also emphasised by its ease of observation, in a majority of cases, and is supported and exemplified by a major case of poisoning which occurred in Scotland in 1979 (Watling, 1980). Here, the victims were able to pinpoint the locality of collection to a coniferous wood near their camping site, a fact which helped greatly in the subsequent identification of the species involved and the clinical management of the

case (Short et al. 1980) (see 3.4.2.2. for further discussion of this case).

2.2.2.1.2.3. Time of Growth. (5)

States : - spring
 - summer
 - autumn
 - winter

The time of growth has been described in terms of the seasons, spring, summer, autumn and winter. There are two reasons for including this character in the key. The first and deciding one is that a common deadly mushroom (Amanita verna) and a few dangerous ones grow at times other than during the summer or the autumn.

Mushrooms are rare in winter or spring and keen collectors or mycophagists may be tempted to try the few species available for collection and consequently have a greater chance of being poisoned than at other times, when common and well known edible species are at hand.

The other reason for using this character is its availability to whoever is using the key.

Nevertheless it remains one of the least useful characters because the peak time for growth of most higher fungi spreads from July to October and that means a very poor selectivity in terms of information which the character conveys.

This feature also shows the same drawback as the previous character in that it can genuinely be mistaken because the time of growth varies depending on the geographical location of the fungus as well as on the climatic conditions for a given year.

To safeguard against a misidentification due to a genuine error, a possible identification based on erroneous data is made possible by a device introduced into the programme, discussed under the separate heading "Near-miss Match Identification" (2.2.2.2.1.).

Example: in extreme weather conditions, summer may start at the beginning of June which, theoretically and strictly following calendar rules, is still spring. Allowance can be made for this misidentification in the same manner as for the previous character.

2.2.2.1.3. Medical Characters.

Here, the only concern for identification purposes relates to the symptoms produced by the mushroom poisoning: the array, the time of onset and the duration of the symptoms. These are well and widely documented. Lincoff and Mitchel's classification (1977) seemed to be appropriate and is used here.

2.2.2.1.3.1. The Symptoms. (1)

- States :
- gastroenteritis after 8 hours, no fever, cytolytic hepatitis.
 - minimum 15 hours after ingestion: gastroenteritis, intense thirst, polyuria then renal failure.
 - after alcohol consumption: flushing face and neck, swelling, tingling of hands; metallic taste, palpitations, hypotension, later nausea and sweating.
 - perspiration, salivation, lacrymation, blurred vision, abdominal cramps, diarrhoea, constriction of pupils, fall in blood pressure, slow pulse.

- - dizziness, drunkenness, incoordination, staggering, muscular cramps, spasms, hyperkinetic activity, deep sleep and visions.
- mood changing, unmotivated laughter, hilarity; muscle weakness, drowsiness, hallucinations.
- gastroenteritis; vomiting
- gastroenteritis after 5 hours; fever; cytolytic and haemolytic action; headaches.

It is practical and useful to group sets of symptoms to form the several states of this character. The symptoms, grouped in a set, are typical of one single type of mushroom poisoning. Seven sets of symptoms characterise seven definite types of mushroom poisoning, while an eighth state includes all the species which give uncharacteristic and usually mild gastrointestinal upsets.

An alternative approach was to divide the character "symptoms" into as many characters as there are symptoms and signs which doctors can observe. Each character would either be present or absent, e.g. a first character "nausea" could be present or absent, a second character "dilated pupils", in the same way, could be present or

absent, and so on, with fever, salivation, lacrymation, hallucinations, hypotension, etc.

This latter approach is unpractical for three main reasons. First, and not least, although it may seem trivial, is the length and tediousness of the procedure. Secondly, if the symptoms are recorded separately, an uncharacteristic and unrepresentative symptom may be selected which would foil the whole process of identification. Finally, a number of these characters, such as gastrointestinal disorders, nausea, emesis, diarrhoea, etc when taken in isolation are common to all or most mushroom poisonings and are therefore irrelevant to identification, but when used in combination with others, as a set or as an array, are characteristic of one single type of poisoning.

Symptoms can be readily divided into those with an immediate or rapid onset and those with a delayed onset after ingestion of the mushrooms. The symptoms provoked by the most dangerous mushrooms always have a delayed onset and are dealt with first.

2.2.2.1.3.2. Delayed Onset of Symptoms.

This concerns three states of the character "symptoms",

the first one, known also as "Amanita Poisoning", the second "Cortinarius Poisoning" and the last one, "Gyromitra Poisoning".

2.2.2.1.3.2.1. Amanita Poisoning.

When gastroenteritis appears, usually after eight to twelve hours, followed by signs of cytolytic hepatitis and a characteristic absence of fever, it is indicative of what is commonly called "Amanita Poisoning" (Mitchel, 1980). No complete and accurate statistics exist on the frequency and proportion of different types of mushroom poisoning, but it is generally admitted that about 50 % of all cases and up to 95 % of all the known fatal cases of mushroom poisoning are due to Amanita poisoning. These values fall to 18 % and 75 % respectively in a ten years' survey by Bornet (1980), but admittedly the mushrooms responsible for the intoxications were unknown in 47.5 % of the cases and in 16.3 % of the cases following death!

2.2.2.1.3.2.2. Cortinarius Poisoning.

This type of poisoning exhibits a delayed action usually of the order of three days but sometimes as short as 15 hours and as long as 17 days. The symptoms start with gastroenteritis, as with "Amanita poisoning", but

are then accompanied by intense thirst and polyuria and followed by renal failure. It was given the name of "paraphalloidic poisoning" when it first came to light in 1952 (Grzymala, 1957) but is more accurately described as "Cortinarius poisoning" because the symptoms are different to those provoked by Amanita phalloides (i.e. Amanita poisoning or phalloidic symptoms) and arise only after ingestion of some Cortinarius species. Some of the more recent serious cases of poisoning in Scandinavia (Hulmi et al., 1974) and Scotland (Short et al., 1980) showed this set of symptoms.

2.2.2.1.3.2.3. Gyromitra Poisoning.

A delayed action ranging from 5 to 8 hours is characteristic of "Gyromitra poisoning". The symptoms are revealed as fever, gastroenteritis with headaches, cramps and hemolysis followed by liver damage. This type of poisoning is serious and death ensues in a great number of cases. Mushroom poisoning of this type arises from highly regarded edible fungi sold as dried mushrooms in markets of a number of European countries. Eaten fresh, these mushrooms are highly toxic, but they lose their toxicity in the drying or cooking process. Lincoff and Mitchel (1977) reported that all deaths in the United

States due to mushroom poisoning other than "Amanita poisoning" arise from this type of mushrooms (Gyromitra and Helvella species).

2.2.2.1.3.3. Rapid Onset of Symptoms.

All other types of mushroom poisonings show an early onset of symptoms starting soon and usually within two hours after ingestion. These poisonings are less serious and rarely fatal in healthy adults.

2.2.2.1.3.3.1. Coprinus Poisoning.

This is state 3 of the character "symptoms". Coprinus poisoning is conditional and appears only if alcohol has been consumed with or after Coprinus species. It shows a remarkable similarity to the disulfiram-ethanol reaction: flushing of the face and neck, swelling of hands and feet accompanied by a metallic taste, palpitations, hypotension and followed by nausea and sweating. The complete metabolism of alcohol is impaired, resulting in the accumulation of toxic levels of acetaldehyde which produce the symptoms described above (Wiseman and Abeles, 1979). This disulfiram-like effect (or Antabuse^R reaction as it is often called) appears immediately after consumption of

alcohol, even if the mushroom has been eaten during the preceding four to five days. No death has been reported from this type of poisoning.

2.2.2.1.3.3.2. Muscarine, Psilocybin and Muscimol Poisonings.

The next three sets of symptoms are becoming increasingly important and common in modern society where large minority groups gather mushrooms for their use as a "recreational drug". This increase in popularity is undoubtedly due to the legal status of the so-called "magic mushrooms", as it is not a controlled drug under the present Misuse of Drugs Act (1971). Underground circles and cultures have very quickly taken advantage of the implications of such a legal status. This is emphasised by a recent Release publication (Release, 1979).

2.2.2.1.3.3.3. Muscarine Poisoning.

The three well known characters of muscarine poisoning, perspiration, salivation and lachrymation appear very quickly after ingestion of the toxin. This is accompanied by blurred vision, abdominal cramps and followed by diarrhoea, constriction of the pupils, fall in blood pressure and slow pulse.

For a long time all mushroom poisonings were attributed to muscarine, for which the antidote is atropine. Unfortunately, this dangerous assumption still persists even in recent pharmacy text books!

In Britain, muscarine is mainly confined to small fairly widespread mushrooms which are unlikely to find their way onto the plate of the mycophagist. However, they may be picked deliberately but mistakenly by "magic mushroom" gatherers!

2.2.2.1.3.3.4. Muscimol Poisoning.

This represent the fifth set of the character "symptoms", or state 5. Amanita muscaria - the "toadstool" or "fairy tale mushroom" - gave its name to muscarine, but the main toxins of this mushroom produce a different set of symptoms, unrelated to those of muscarine. They are dizziness, drunkenness, incoordination, staggering, muscular cramps, spasms, hyperkinetic activity, deep sleep and visions. These symptoms are produced by a number of common and closely related mushrooms which are sought after for their inebriating and hallucinogenic properties by some sectors of society. Amanita muscaria was widely used until fairly recently in Siberia and a long history of religious and shamanistic

traditions surrounds this mushroom (Wasson, 1968).

2.2.2.1.3.3.5. Psilocybin Poisoning.

This is the last set of characteristic symptoms elicited by some mushrooms after ingestion (or state 6). Hallucinogenic mushrooms produce symptoms similar to those recorded for LSD following ingestion. They include mood changes, unmotivated laughter, hilarity, muscle weakness, drowsiness and hallucinations and are often described under the collective name of "psilocybin poisoning".

Hallucinogenic mushrooms have attracted the interest of research scholars of the history of religions and ethnopharmacology, as well as a fringe of an hedonistic section of modern society. They have often been described as the "Mexican hallucinogenic mushrooms" after G. Wasson discovered, participated and described (Heim and Wasson, 1958) religious rites based on these mushrooms still taking place in some remote parts of Mexico. Hallucinogenic mushrooms were later found to be widespread throughout the world and their study form a substantial part of the research carried out in the present thesis (Margot and Watling, 1980).

2.2.2.1.3.4. Discussion.

For practical reasons, these descriptions of symptoms are limited to a few characteristic signs which are present when a specific type of poisoning occurs. It does not, in any way, describe or represent the whole array of medical information which can be gained from the patient. These symptoms have been widely, fully and expertly described by many authors and in many languages (English: Lincoff & Mitchel, 1977; Rumack & Salzman, 1978; German: Flammer, 1980; French: Heim, 1963/1978; Borner, 1980; Italian: Arietti and Tomasi, 1969/1975; etc) and it is to these authorities that the clinician is referred for further details, or, better, to the bibliographic reference retrieved with the present programme when an identification is obtained. A full description is beyond the scope of this thesis and would not serve in the identification of the causal agent of a mushroom intoxication, - the primary purpose of this programme.

2.2.2.1.3.5. Onset of Symptoms. (2)

- States :
- within 2 hours
 - 3 to 8 hours
 - 8 to 12 hours
 - typically over 15 hours

Repeated reference to the delay of the onset of symptoms was done in the previous paragraphs (2.2.2.1.3.2. to 2.2.2.1.3.3.). It is a primordial diagnostic factor which can mean the life or death of a patient. To emphasise its importance, it has been included as a separate character which can be used by non medically trained users of the programme. Borner (1980) has shown from his studies over a ten years period that this character is known in 95.9 % of cases.

The onset of symptoms can be described in terms of the four selective states given above.

If symptoms appear within two hours of ingestion, all the most poisonous species of mushrooms to be found can be eliminated as being responsible for that clinical condition. This therefore makes a useful distinction between all the toxic species. The three other states conveniently divide the remaining and mostly dangerous species into three specific groups. A delay of action of three to eight hours is shown in cases of "Gyromitra poisoning" (usually 6 to 8 hours). "Amanita poisoning" is characterised by a delay of more than 8 hours but less than 15 hours (typically 10 to 12 hours) whereas "Cortinarius poisoning" shows a very long delay of action ranging from 15 hours to 17 days in extreme cases,

but typically of about 3 days. Earlier signs of gastroenteritis have been reported by Flammer (1980) in a few rare cases of "Cortinarius poisoning" but this still needs further confirmation.

2.2.2.1.3.6. Duration of Symptoms. (15)

- States :
- weeks if not death
 - 2 to 4 hours
 - up to 6 hours
 - 6 to 24 hours
 - few days

The character "duration of symptoms" would seem to be more of an academic than practical interest and has no place in an identification process where emergency toxicology is concerned.

Nevertheless, it was thought useful to include this character as a preventive as well as for a medico-legal reason. In the preventive context, the identification of the causative agent of an intoxication, whether the victim dies or not, is necessary. People could be advised via the mass media that a certain poisonous species of mushroom is growing in a given geographical location and at a given time of the year. Furthermore, a full description and a warning could be issued, thereby preventing

further intoxications.

In the case of death, the identification of the causative agent becomes the main medico-legal concern and it can be of utmost importance (for example for insurance purposes) to determine that a poisoning is accidental rather than criminal or due to pollution or other hazards of modern society.

This necessity for identification warrants the inclusion of this character in a general multipurpose identification programme.

The first state of this character, "weeks if not death" is shown in all serious cases of intoxication. Slow acting toxins, which are present in the most dangerous mushrooms, may affect the victim of a poisoning for weeks. Sooner or later, depending on the individual and the dose ingested, the outcome may be death.

The second state, "2 to 4 hours" is very selective. Only conditional poisoning of the "Coprinus poisoning" type show symptoms appearing immediately after ingestion of alcohol and disappearing very quickly : within 2 to 4 hours.

Individuals poisoned by hallucinogenic mushrooms are affected up to five or six hours after ingestion.

It represents the third state of the character "duration of symptoms". This does not take into account possible recurrence of the symptoms at later dates (flashbacks as experienced with LSD).

The fourth state (6 to 24 hours) is experienced with most of the other poisonous mushrooms . Their effects gradually reside within one day.

Finally, a few species show more serious symptoms which can last for a few, usually two to three, days. Poisoning cases with these species are fairly common and a separate state - "few days" - is very useful in that context.

The time periods over which individuals adversely respond to mushrooms, just like the delays in the onset of the symptoms, are characteristic of given toxins and therefore are valuable diagnostically.

It should always be borne in mind however, that the medical history of the patient before the poisoning occurred may affect the duration of the symptoms, or even the outcome of the poisoning. An unhealthy victim may die from exposure to a mildly poisonous mushroom or small doses of toxin. This should be common sense for anybody working in the field of toxicology or medicine and should be allowed for when using the key.

2.2.2.1.4. Chemical Characters.

Only one character has found a place here: the toxins. The chemistry of most of the higher fungi is very poorly described. Furthermore, compounds known to occur in some species have not been widely surveyed in other species. The result is that the distribution of the toxins, within identical species of different origins or between species, is unknown.

2.2.2.1.4.1. Toxins. (14)

- States :
- cyclopeptides
 - orellanine
 - muscarine
 - hallucinogens
 - muscimol/ibotenic acid
 - coprine
 - others
 - gyromitrin

At the present time, this character is only useful when fresh material is available for analysis, and will certainly not be used in cases of emergency toxicology for a while yet. The study of known toxins in the human body, their analysis and distribution is non-existent or,

at the very best, poorly documented. It is hoped that this will be a major area of research in the next few years.

Some personal research has helped to complete some data used in this programme and is discussed and described in the last part of this thesis (chapter 4).

Seven groups of known toxins form seven states of the character "toxins", whereas the eighth state, defined as "others" include all the yet unidentified toxins contained in a number of mushrooms. The state "gyromitrin" is placed last in the above list because it was added to the programme at a late stage.

2.2.2.1.4.1.1. Cyclopeptides.

The toxic cyclopeptides, characteristic of "Amanita poisoning", are responsible for the majority of deaths due to mushrooms (Mitchel, 1980; Bornet, 1980). The most lethal of these toxins are the amatoxins (Figure 18), first identified in Amanita phalloides, the "Death Cap" (Wieland and Faulstich, 1978). The amatoxins are a group of eight octapeptides that inhibit the transcription of RNA and DNA. As a consequence, protein synthesis and cellular reproduction are blocked, followed by necrosis

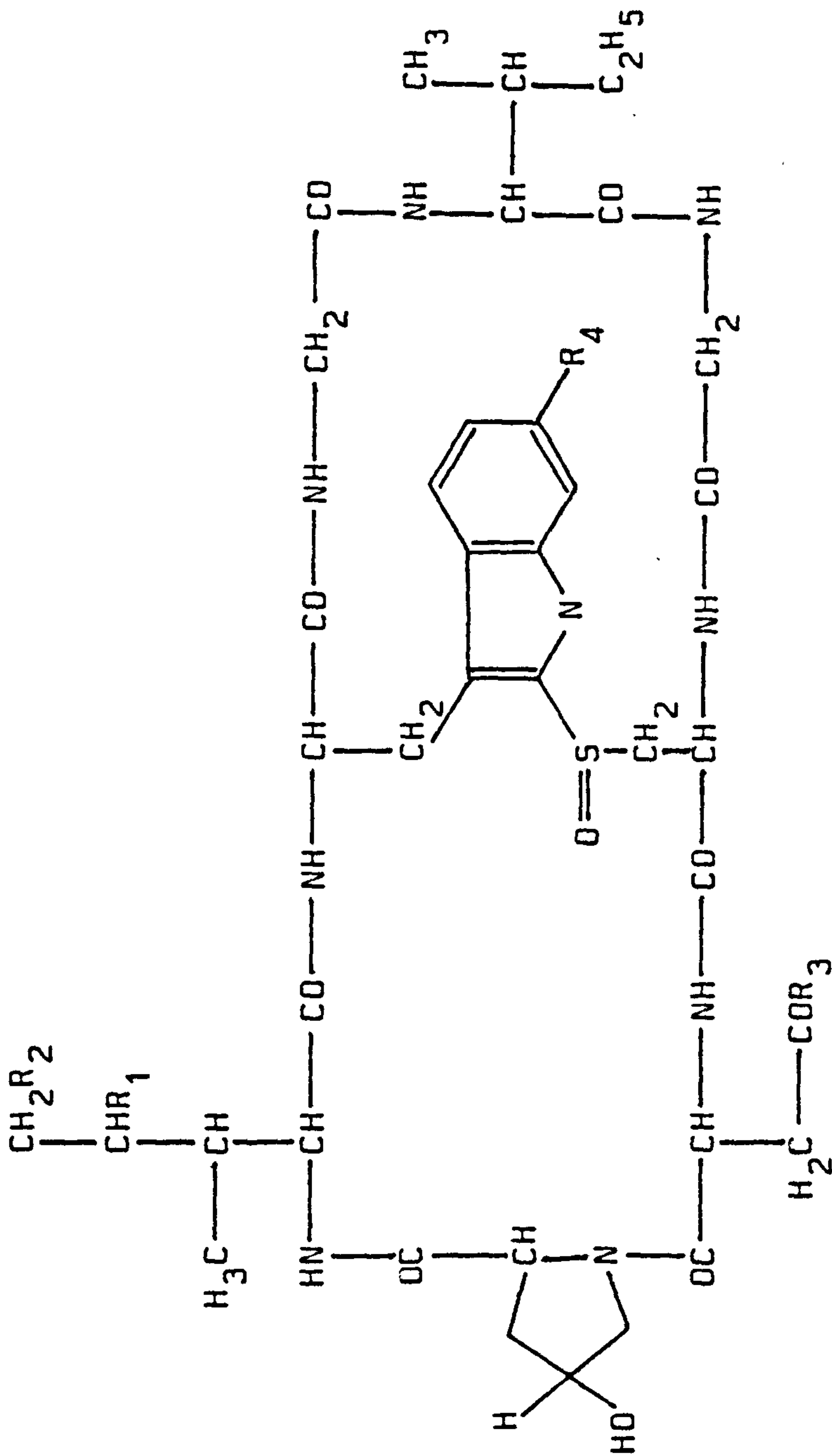


Figure 18 : Description of the General Structure of the Amatoxins

Figure 18 : Legend : respective structures of the related amatoxins

	R ₁	R ₂	R ₃	R ₄
α - amanitin	OH	OH	NH ₂	OH
β - amanitin	OH	OH	OH	OH
γ - amanitin	OH	H	NH ₂	OH
δ - amanitin	uncertain			
ξ - amanitin	OH	H	OH	OH
Amanin	OH	OH	OH	H
Amanullin	H	H	NH ₂	OH
Amaninamide	OH	OH	NH ₂	H

of the affected cells. Seven heptapeptides, the phallotoxins have also been found in Amanita phalloides (Figure 19) but they are probably not involved in cases of human poisoning because of their heat lability. More recently, new cyclopeptides have been isolated and identified in Amanita virosa (the Destroying Angel). They are the virotoxins, related to the phallotoxins (Faulstich et al., 1980), and also amaninamide (see Figure 18) which is closely related to the amanitins (Buku et al. 1980).

A number of Amanita, Galerina and Lepiota species contain amanitins (Gérault and Girre, 1977) and are therefore highly dangerous. A number of analytical techniques have been devised and used in their characterisation and are detailed in the appendix (6.2.1.)

2.2.2.1.4.1.2. Orellanine.

There is much controversy surrounding the identity of this toxin or group of toxins (Greer, 1980). Undoubtedly it differs chemically from the Amanita toxins. Furthermore its physiological action also differs, the attack being observed in different organs, primarily the kidneys (Nieminen, 1976).

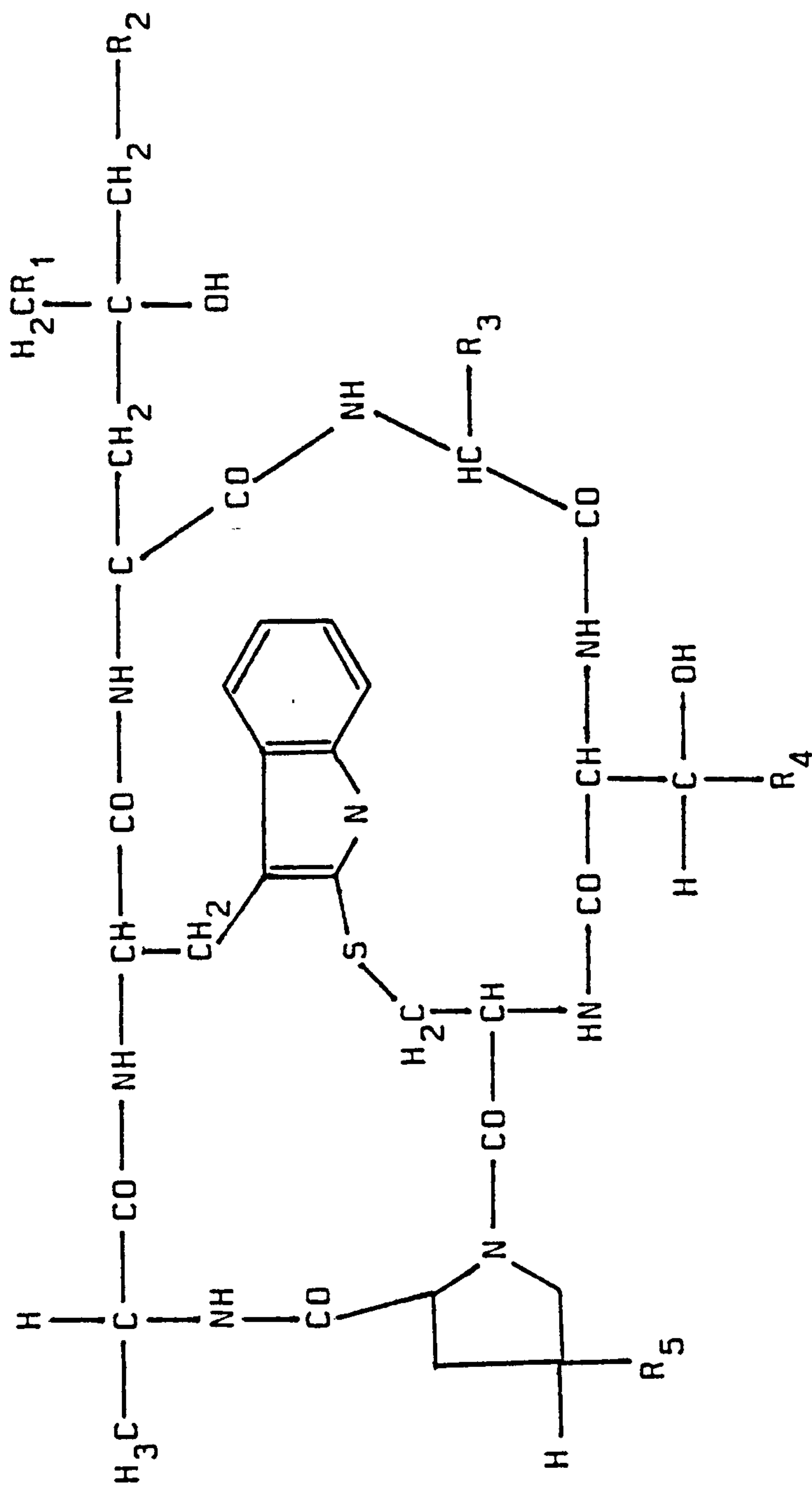


Figure 19 : General Structure of the Phallotoxins

Figure 19 : Legend : respective structures of the related phallotoxins.

	R ₁	R ₂	R ₃	R ₄	R ₅
Phalloidin	OH	H	CH ₃	CH ₃	OH
Phalloin	H	H	CH ₃	CH ₃	OH
Phallisin	OH	OH	CH ₃	CH ₃	OH
Phallacin	H	H	CH(CH ₃) ₂	CO ₂ H	OH
Phallisacin	OH	OH	CH(CH ₃) ₂	CO ₂ H	OH
Phallacidin	OH	H	CH(CH ₃) ₂	CO ₂ H	OH
Phallin B	H	H	CH ₂ C ₆ H ₅	CH ₃	H

Antkowiak and Gessner in 1979 proposed a tetrahydroxy bipyridyl structure for orelline and its bis N-oxide derivative orellanine (Figure 20), structures which are still unconfirmed (Antkowiak and Gessner, 1980; Greer, 1980).

Analytical procedures for the isolation and partial characterisation of these compounds are described in the appendix (6.2.2.)

2.2.2.1.4.1.3. Muscarine.

Muscarine, first extracted from Amanita muscaria by Schmiedeberg and Koppe in 1869 was only characterised in 1954 by Eugster and Waser (Figure 21). For a long time muscarine was thought to be the cause of all mushroom poisoning, but recent work has shown that it does not even account for the toxicity of Amanita muscaria due to its very low concentration, coupled with a low oral toxicity.

It is, nevertheless, a common toxin found in high concentrations in a number of small mushroom species belonging to the genera Inocybe and Clitocybe. Its analysis is not easy and only a few claims for the presence of muscarine in higher fungi are based on chemical

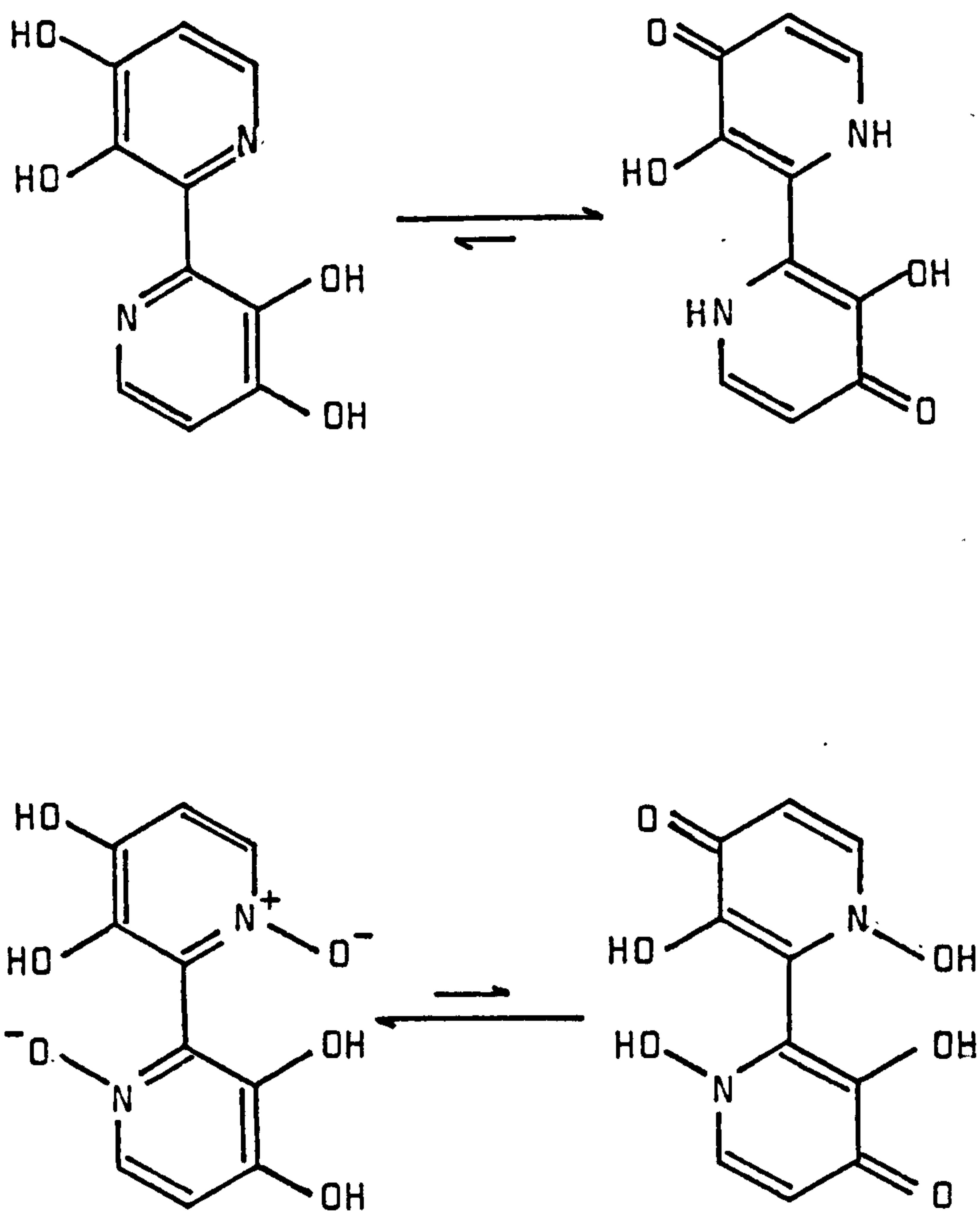


Figure 20: Proposed Structures for Orelline (top) and Orellanine (bottom) (Antkowiak & Gessner, 1979).

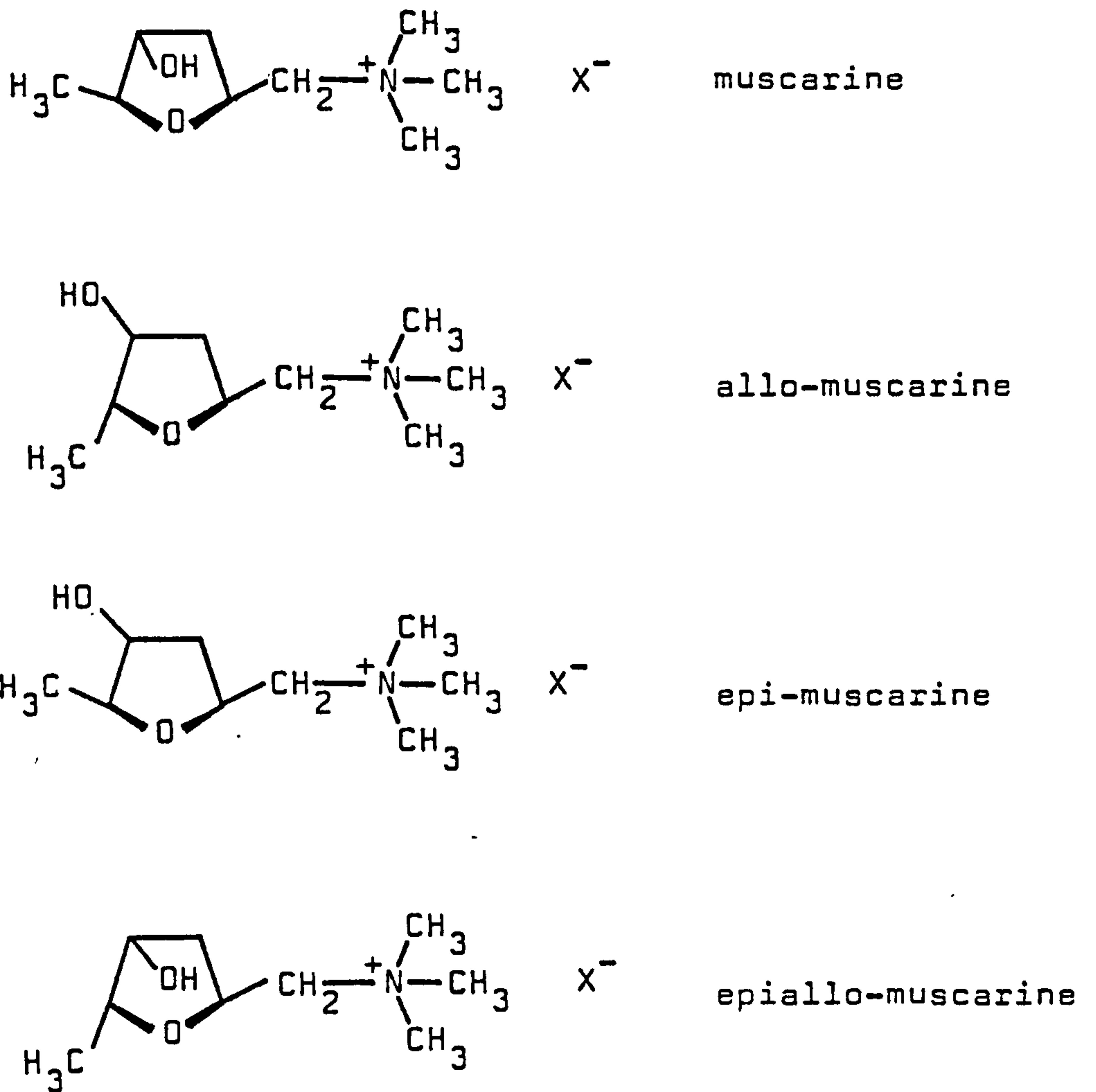


Figure 21: Structure of Muscarine and its Isomers
(Catalfomo & Eugster, 1970).

analysis (Eugster, 1969). The remaining claims are based on animal work and must therefore be considered as unsubstantiated. However, until further data are available, the present claims are used in this programme.

Detailed analytical procedures are given in the appendix (6.2.3.)

2.2.2.1.4.1.4. Hallucinogens.

Since the discovery of psilocybin and psilocin in 1958 and 1959 respectively by Hofmann and co-workers, two more indole hallucinogens of the higher fungi have been discovered: baeocystin and norbaeocystin, two analogs of psilocybin (Figure 22) (Leung and Paul, 1967; *ibid.*, 1968).

In the past ten years, claims for the presence of some of these compounds in different mushrooms belonging to the genera Psilocybe, Stropharia, Conocybe, Hypholoma, Panaeolus, Gymnopilus, have been numerous and often unconfirmed. The list of species known to contain psilocybin and analogs is limited (see Table 3). Suspect species have also been included in the programme despite some negative results obtained in this laboratory (4.2.3.) These latter studies show that a number of claims for

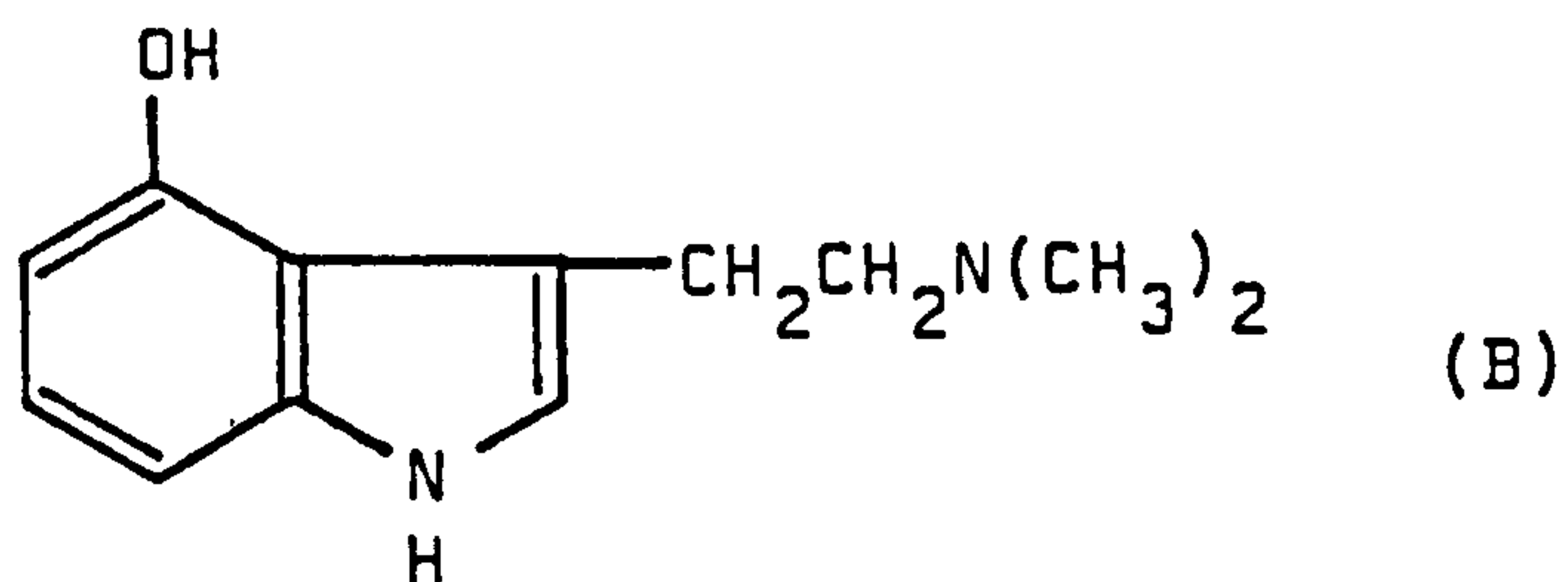
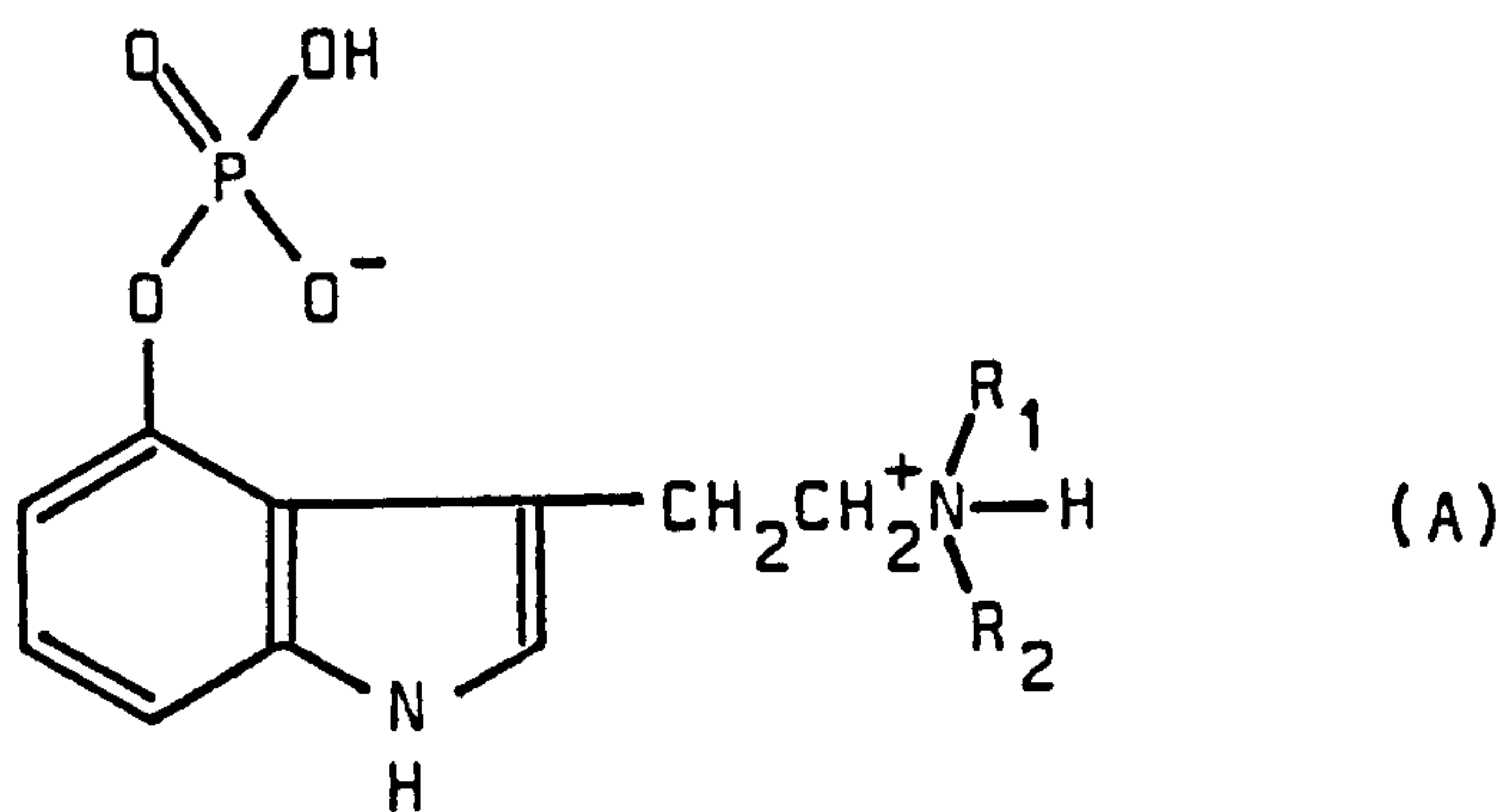


Figure 22: Structure of Psilocybin and Analogues

	R_1	R_2
with (A) psilocybin	CH_3	CH_3
baeocystin	H	CH_3
nor-baeocystin	H	H
(B) psilocin		

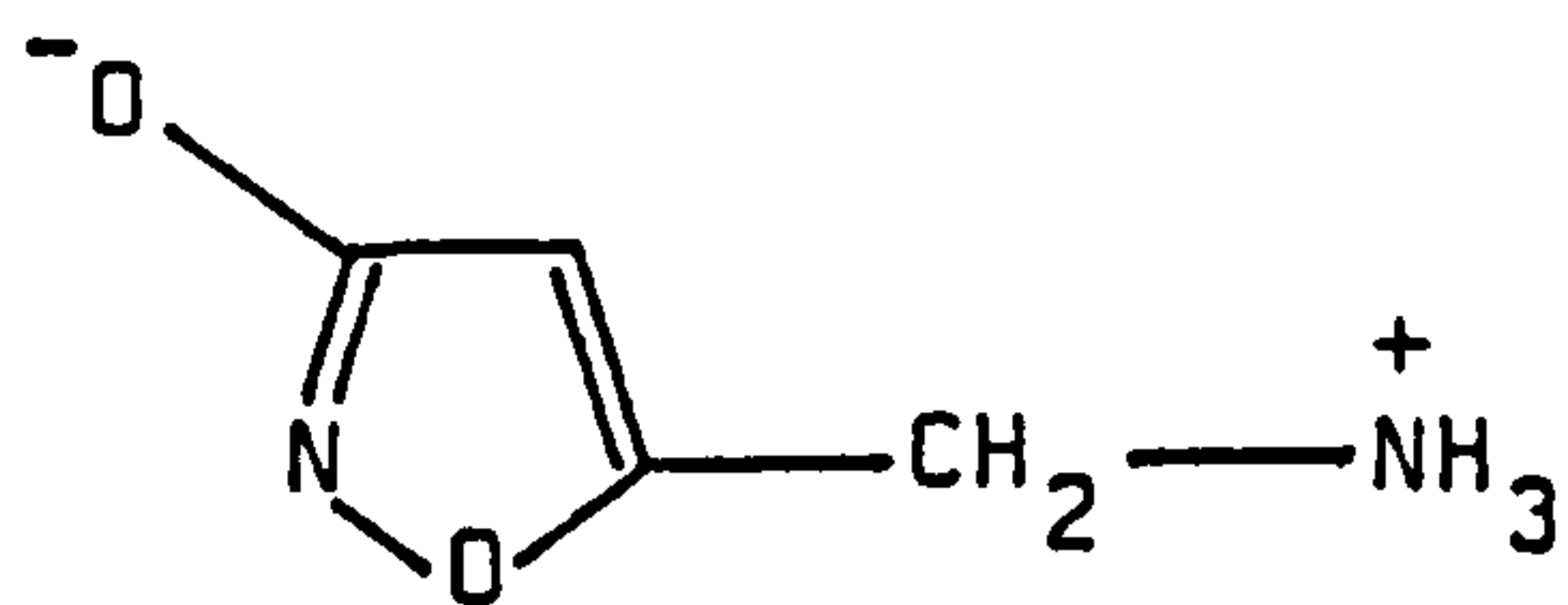
the presence of these compounds may be unfounded, or even, in the case of Panaeolus species, that other indole hallucinogens may be involved. Certainly, results presented by Ola'h (1969) could not be confirmed, thus supporting previously recorded doubts (Singer, 1975).

However, definite cases of intoxications, showing symptoms of psilocybin intoxication with Panaeolus species have occurred and have been investigated at Strathclyde University. It seems other, yet unidentified, hallucinogens may be at work here.

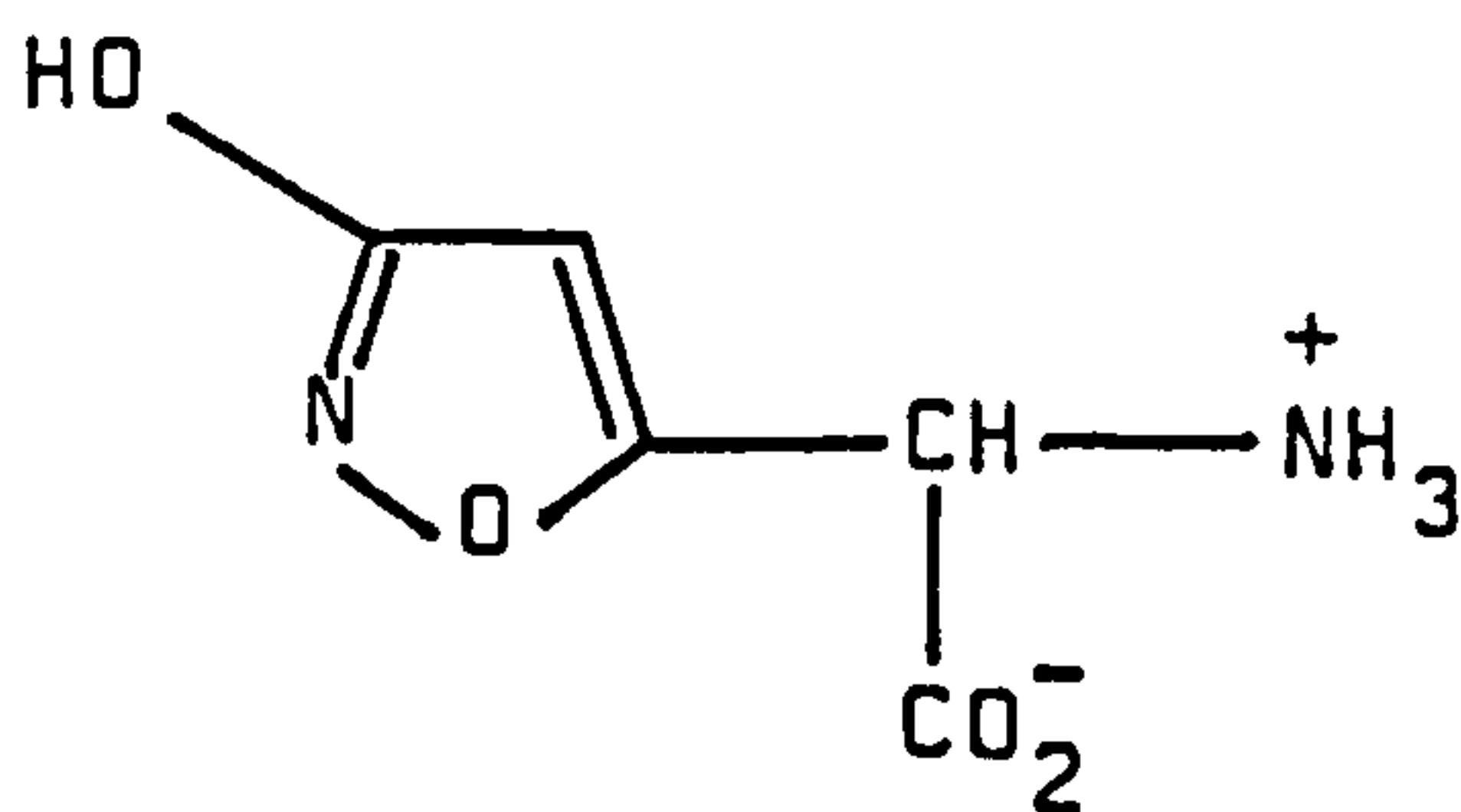
A thorough investigation of suspect species is necessary, and taxonomic positions should be made clear before further claims are made in the future. Furthermore, voucher specimens should be deposited in recognised centres.

2.2.2.1.4.1.5. Muscimol and Ibotenic Acid.

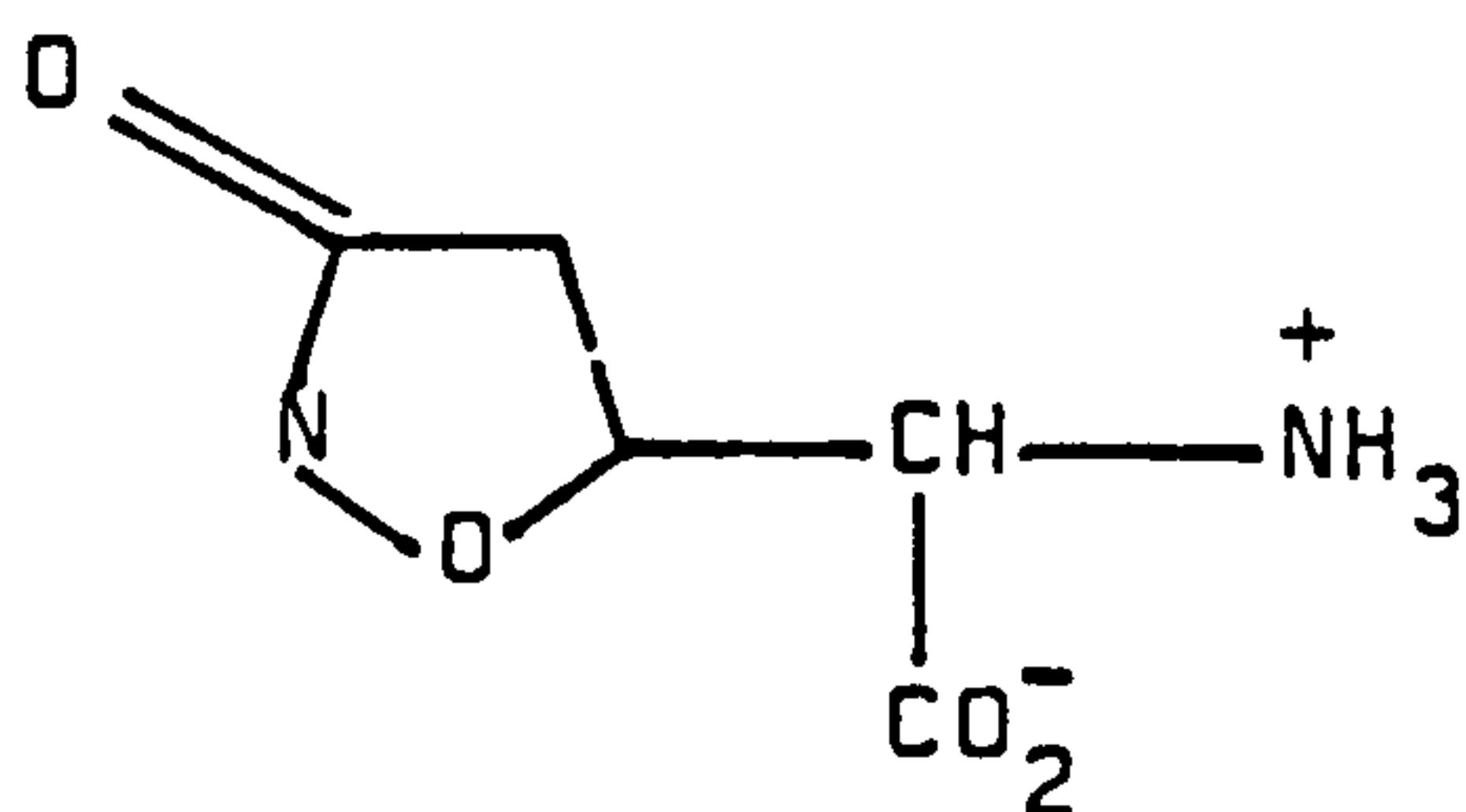
A number of compounds are included in this "state", which possess an isoxazole derivative structure (Figure 23) and which account for most of the symptoms shown by the, now famous, Amanita muscaria. They were isolated and characterised by



Muscimol



Ibotenic Acid



Tricholomic Acid

Figure 23: Structures of Isoxazole Derivatives
Found in the Higher Fungi.

the Eugster team in Zürich from Amanita muscaria (Eugster, 1969). These compounds, muscimol and ibotenic acid, do not explain the whole range of physiological action experienced during intoxication with this mushroom. Eugster's work is the closest, yet known, to a complete investigation of the chemistry of one fungus. Ibotenic acid and muscimol are important compounds used by pharmacologists to obtain an understanding of the mechanism of γ -amino butyric acid (GABA) in the brain.

Mushrooms containing these isoxazole derivatives are well known and easily recognised by even the least experienced amateurs which means that accidental poisonings are unlikely. Most intoxications will be self-inflicted in the experimentation and search for natural and uncontrolled drugs. Extensive descriptions of Amanita species containing these toxins are to be found in the underground literature dealing with drugs (Margot, 1977).

Analytical procedures are detailed in the appendix (6.2.5.).

The related tricholomic acid (Figure 23) has been found in Japanese Tricholoma muscarium species and may be at the origin of the toxicity of other Tricholoma species found in Europe.

2.2.2.1.4.1.6. Coprine.

Hatfield and Schaumberg (1975) identified coprine, a cyclopropane derivative of glutamine (Figure 24), which inhibits the metabolism of alcohol in humans. Its effect, with alcohol, shows a remarkable similarity to the reaction elicited by disulfiram and alcohol, the former being a drug used in the treatment of chronic alcoholism. The toxin is not dangerous and is present in only a few species of Coprinus. It is unlikely to be found in emergency cases. The species involved are succulent and considered as delicacies by many, but, if alcohol is consumed, even two or three days after the mushrooms, it may trigger a rather unpleasant reaction which may lead to panic in the victim.

The analysis for coprine is detailed in the appendix (6.2.6.).

2.2.2.1.4.1.7. Gyromitrin.

Serious intoxications, with numerous fatalities (2 - 4 % of mushroom related deaths), have occurred in Europe involving a mushroom often sold as a delicacy in its dried form : Gyromitra esculenta. Gyromitrin (Figure 25) was first identified by List and Luft (1968)

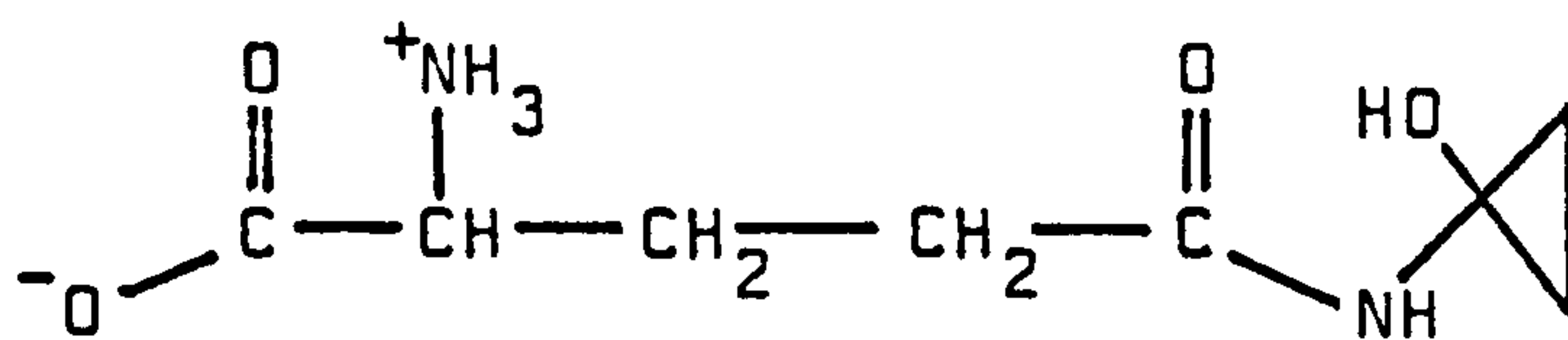


Figure 24 : Structure of Coprine

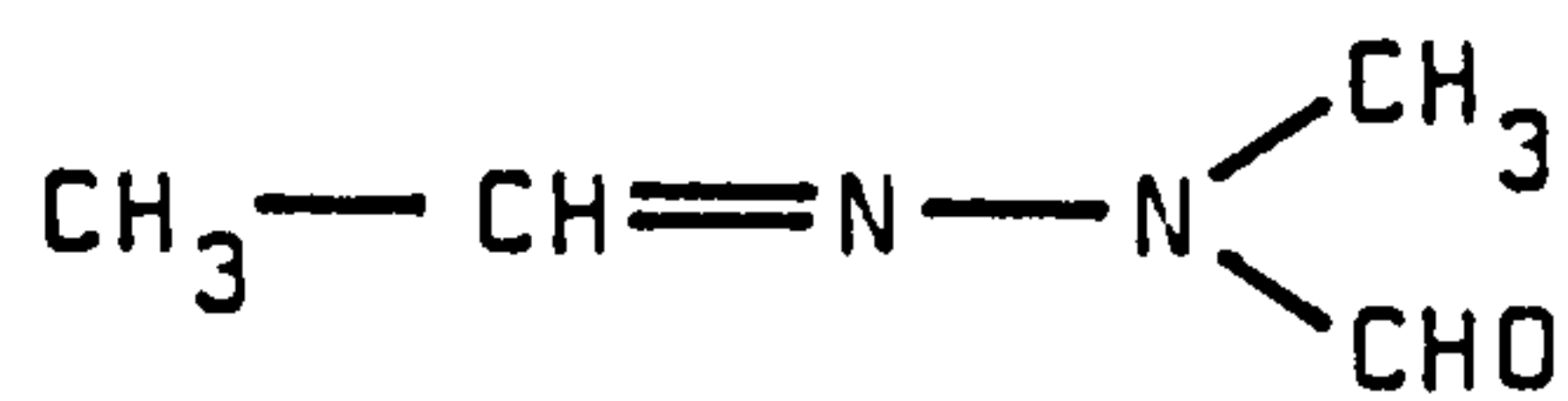


Figure 25 : Structure of Gyromitrin

and further hydrazone derivatives were later identified by Pyysalo (1975). It is suggested that these compounds undergo hydrolysis to form monomethylhydrazine, which is presumed to be the active component in human poisoning. The symptoms are consistent with symptoms produced by monomethylhydrazine poisoning (Lincoff & Mitchel, 1977).

Species containing gyromitrin are Ascomycetes and differ markedly (microscopically as well as macroscopically) from all the other higher fungi. It was found necessary to include them in the present programme because of the high incidence of poisoning due to them.

Analytical procedures are described in the appendix (6.2.7.)

2.2.2.1.4.1.8. Other Toxins.

Every year, a number of poisonings are recorded which involve tens of species of mushrooms. The symptoms produced are often described as mild to serious gastrointestinal disturbances which disappear after one or two days, without further ill effects. In most cases nothing is known about the chemistry of the species involved, not to mention the toxins! In these cases, it frequently arises that only one or two people of a much larger group

are affected.

This remains a virgin field for research!

2.2.2.1.4.1.9. Comments.

It must be noted that poisoning by old, partially decomposed mushrooms, or by mushrooms contaminated by pesticides or heavy metals (Stijve & Roschnik, 1974) are not included in the present work because the intoxications are not due to toxins of the higher fungi.

Other important questions require to be investigated in relation to the mycotoxins of the higher fungi. What, for example, is the relationship of the host to the fungi in respect of the formation of toxic secondary metabolites? Does the toxin change if the host is different? For example, Amanita muscaria grows in mycorrhizal association with either birch or coniferous trees. The influence of the host tree on the metabolic activity may explain the absence of the toxins muscimol and ibotenic acid in some of the analyses reported in the chemical literature. (Catalfomo & Eugster, 1970). These questions need further investigations and might provide answers of great taxonomic significance and of great economic value to man.

2.2.2.2. Discussion.

Twenty four characters have been chosen to be included in a multi-access, interactive identification programme. If sub-characters are taken into account, 30 features can be used for the identification of poisonous and hallucinogenic mushrooms of forensic interest. All have advantages and disadvantages which have been discussed under their appropriate heading. All the characters are well documented, at least for the European species, and provide a solid data base. Furthermore, they are all stable within wide and documented limits and, in most cases, easy to identify and observe. Finally, different combinations of characters, using only five to eight observations give, in most cases, an identification down to species or a group of closely related species, as is shown in the next two chapters.

It must be emphasised that some closely related species cannot be differentiated using the above twenty four characters and would be identified only with great difficulty even by experienced mycologists, unless fresh samples were made available. In any case, when an emergency has been dealt with, samples should be sent to the appropriate specialists who can confirm the diagnosis, use the evidence to up-date records and, possibly, bring

about modifications to the present programme for even more efficient management. Detailed records of the anamnesis and clinical data should be given with the samples.

This should result in a marked improvement on the present state of the identification of poisonous mushrooms in emergency cases. For instance, such identification has been made by a specialist in only 10.7 % of the cases reported in Switzerland between 1967 and 1977 (Bornet, 1980), and this in a country specifically organised to deal with such matters!

As mentioned previously, the states of some of the characters observed could be genuinely mistaken under certain circumstances, thus hindering a possible match with the stored data in the data bank. Steps have been taken to prevent this happening and to allow what can be referred to as a "Near-miss match identification".

2.2.2.2.1. Near-miss Match Identification.

Five characters were found, after close scrutiny, to be uncertain when observations were made near the limits of their defined states because of their continuous nature (i.e. the length and breadth of spore, see 2.2.2.1.1.10.; the wall thickness of spores, see

2.2.2.1.1.13.; and the time of growth, see 2.2.2.1.2.3.) or because of historical and ecological reasons (i.e. habitat, 2.2.2.1.2.2.) as was demonstrated in the appropriate chapters.

If the user of the key, for some reason, thinks that the determination of one or more of these five characters is doubtful, he has then the opportunity to choose whichever of these characters is uncertain as a possible miss-match with the stored data.

Example: a mushroom identification is required in a hospital after a patient has been brought in showing definite symptoms of poisoning. Spores are found in a stomach washing, their colour determined and their dimensions found to be well-defined within the ranges used in the programme. The time of growth, 19th of June, is identified as "spring", despite the closeness of summer and a spell of mild weather. The user, with this data, should be able to get a reasonable identification, but having some doubts as to the determination of the character "time of growth", he can choose this character as a possible miss-match. That is, if no "perfect match" is obtained, species differing on the character "time of growth" with the given data are then listed as "species matching closely on character time of growth".

If more than one character's data is allowed to mismatch or permitted to disagree with the data stored, when a near-miss match identification is obtained, the character(s) in disagreement with the taxon identified is (are) specified to the user. In the above hypothetical example, if the spore length and the time of growth are judged uncertain and allowed to differ with the data of the data bank, the resulting identification could be of the kind:

Species A matches closely on spore length;

Species B matches closely on time of growth;

Species C matches closely on spore length and time of
growth;

Species D matches exactly with the given data, etc.

Therefore, the characters allowed to disagree with the stored data in the identification process are well defined and the user is made aware as to which character observed disagrees with the data defining the taxon identified.

The "near-miss match identification" facility should be used when the character "habitat" is observed, unless a well characterised and historically well established habitat is known, for example, the native Scottish pine forest.

While this facility may not be relevant in most cases, it represents an important safeguard which guarantees the completeness and accuracy of an identification as far as is possible with the data at hand.

CHAPTER III : THE PROGRAMME

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III. THE PROGRAMME.

On the basis of the requirements for an identification programme (chapter II) and characters available, some of which possess single and others multiple options, a programme was built by Mr. George Farquhar of the Computer Centre of Strathclyde University. His expertise was invaluable in helping to improve and modify this programme to suit all the requirements.

3.1. Data Coding.

Before an identification is attempted, the data of all the taxa included in the data bank had to be coded. To make the matching process easy for the machine, the data is coded as a string of numbers, where the first number corresponds to the state of character 1 of the given taxon, the second number corresponds to the state of character 2, and so on. When multiple options are possible for one character a "-1" indicates that the character has multiple options. The numeral immediately following "-1" indicates the number of possible options which is, in turn, followed by the numerals corresponding to the different possible states.

The way the data is coded can be likened to a chest

of 24 drawers where each drawer pictures a character. Each drawer is then subdivided into as many compartments as there are options for the given character. These pigeon-holes are filled corresponding to the states shown by the taxon defined.

When the identification is attempted, the data observed is coded in a similar way, the characters identified are like drawers being opened and their contents compared with the same level drawers of the different taxa stored in the data bank. When the content of these drawers are identical (i.e. match exactly) a possible identification is obtained. If a near-miss match is allowed, all the drawers' contents should be identical except possibly the drawer(s) allowed to mismatch (i.e. provided the suspect data was wrongly characterised).

3.2. Flow Chart. (Figure 26)

The computer is an extremely powerful, fast machine, but it can do only what it is told to do, and exactly as it is told to do it. It gets its direction from the stored programme. The flowchart is a pictorial representation of that programme which serves as a means of recording, analysing and communicating problem

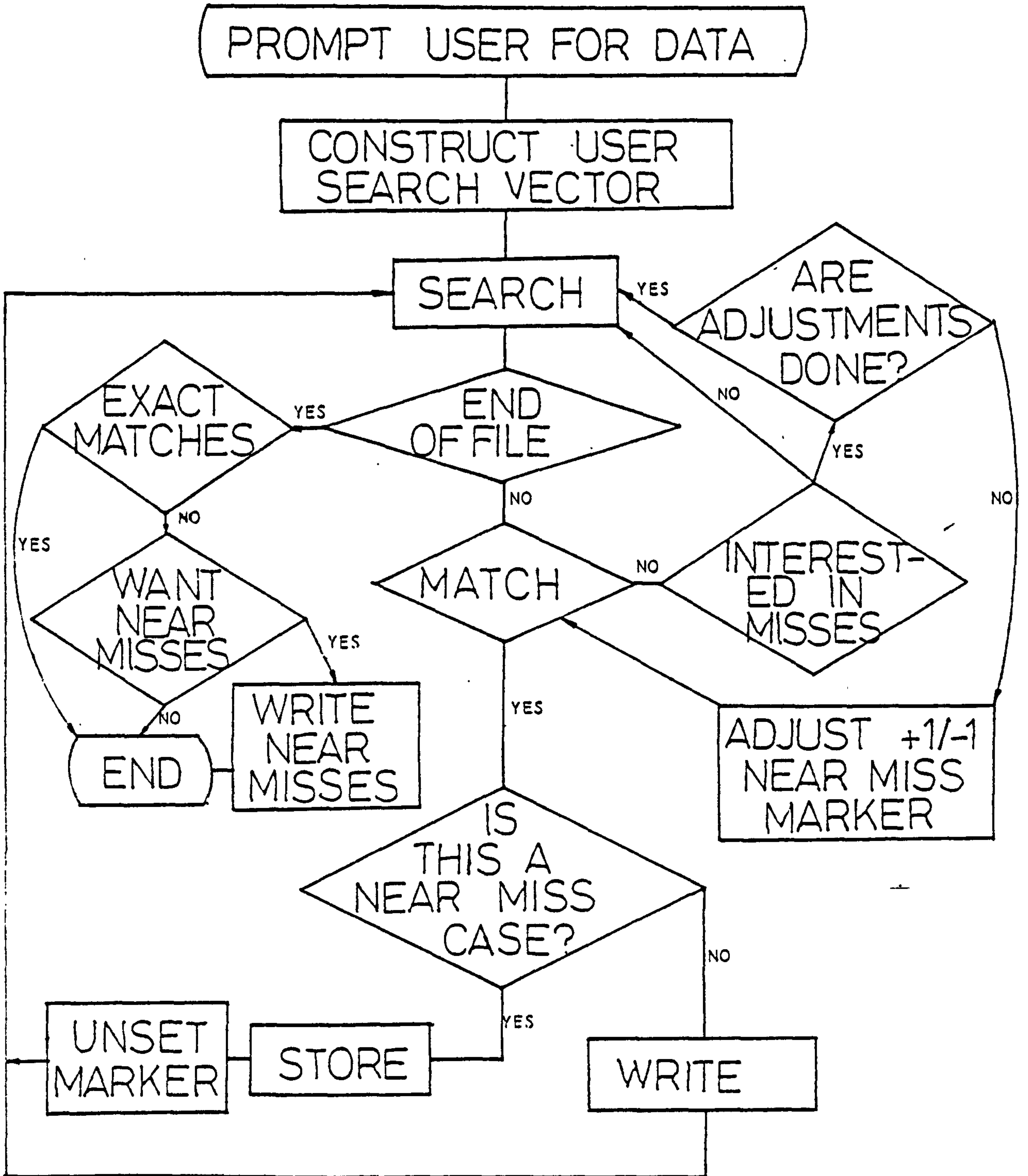


Figure 26 : Flow Chart of the Computer Programme for the Identification of Poisonous Mushrooms.

information.

Following the flowchart (Figure 26), the first step in the identification process is to prompt the user for data (i.e. open drawers and fill them up according to the states observed). The whole data thus obtained constitutes the user search vector.

This search vector is then compared with the corresponding vector of each taxon of the data bank. When the "SEARCH" is operated for the first taxon, the computer effects the comparison. If a match is found, which is not a near-miss match case, the taxon is given as a possible identification (i.e. "WRITE"). It then searches the second taxon, etc until the end of the data file. If there is no match, and no interest shown by the user in near-miss matches, as before, it searches the next taxon, and so on to the end of the file. All the exact matches, i.e. taxa identified, are printed. Ideally, only one taxon should appear!

On the other hand, if the user is interested in near-misses, the user search vector is modified ("ADJUSTED") for the character(s) allowed to disagree, so that other states for this character(s) are considered and the matching process ("SEARCH") is attempted again, but with the adjusted data. If after adjustment a match

is obtained, the taxon so identified is stored until the end of the file before being printed.

Each taxon identified is printed ("WRITE"), usually on the screen of a visual display unit (VDU), or for a permanent record, on the chart of a recorder. It is always accompanied by all the useful data included in the data retrieval side of the file. (2.2.1.)

3.3. Languages.

The programme has been produced in two forms so that it can be used in a large computer, as well as in a top-desk microcomputer.

A FORTRAN language is used on a Honeywell 6060 instrument at Strathclyde University. It has been kept to a standard form and is as machine independent as possible, so that it could be easily adapted to other machines.

In order to avoid any involved coding procedure of the data, which would require some training, a text in English is displayed on the VDU or printer. The user is only required to choose one option (or more as the case may be) out of a selected number presented to him and the computer does the rest of the work providing

the correct commands have originally been used.

A programme in BASIC language has been adapted along the same lines to be used on microcomputers, in our case on Apple II and Honeywell.

This should allow a wide use of this programme by large organisations with expensive computer capabilities as well as small laboratories or hospital wards with limited facilities.

3.4. Use of the Programme and Examples.

3.4.1. How it Works.

The basic set of commands is small and easy for anybody to remember.

The first command "/G" prompts the computer to search for the programme. In the meantime, it asks the user which data file he wishes to use with the programme by printing "Data File?". The user simply types the name of his file (here "KEEP") followed by a carriage return (CR).

The next command is "/F1" which prompts the user to supply data by displaying the 24 characters and asking the user "Which characters do you have data for?" (Figure 27)

./G
DATA FILE? YTEP

./F1

FUNGI IDENTIFICATION PROGRAM

1 SYMPTOMS	13. SPORE PRINT
2 ONSET OF SYMPTOMS	14. TOXINS
3 GEOG DISTRIBUTION	15. DURATION OF SYMPTOMS
4. HABITAT	16. PILEUS TRAMA
5 TIME OF GROWTH	17. GILL TRAMA
6 SPORE COLOUR UNDER M. SCOPE.	18. CUTICLE
7 SPORE SHAPE	19 CLAMP CONNECTION
8 SPORE SURFACE	20. BASIDIA/ASCI NO. OF SPORES
9 SPORE WALL THICKNESS	21 BASIDIA/ASCI SIZES
10 GERM PORE	22. MARGINAL CYSTIDIA
11 SPORE LENGTH	23. FACIAL CYSTIDIA
12 SPORE BREADTH	24. KOH CHRYSO CYSTIDIA

WHICH CHARACTERS DO YOU HAVE DATA FOR?
ENTER CHARACTER NUMBERS COMMA SEPARATED
=1,3 4, 13,5

DATA FOR 5 CHARACTERS REQUIRED

IF NO SPECIES MATCH EXACTLY THE GIVEN DATA A CLOSE MATCH
CAN BE ATTEMPTED ON THE FOLLOWING CHARACTERS:-

4. HABITAT
5 TIME OF GROWTH
9 SPORE WALL THICKNESS
11. SPORE LENGTH
12. SPORE BREADTH

PLEASE GIVE THE CHARACTER NUMBERS YOU REQUIRE (COMMA SEPARATED)
OR 0 (ZERO) IF NO CLOSE MATCHING IS TO BE ATTEMPTED
=4,5

CLOSE MATCHING REQUIRED ON CHARACTERS 4, 5,

Figure 27 : Illustration of the First

Commands to Start the Programme

and of the Text Displayed to the

User.

The user needs only to enter the numbers corresponding to the characters for which he has data, i.e. 1, 3, 4, 13 and 5 in the example of Figure 27. After a carriage return, the computer acknowledges the number of characters to be used (i.e. number of drawers to be opened) and asks if any "near-miss match" is to be attempted on one of the five characters described previously (2.2.2.2.1.). The user enters "0" if no close matching is required, otherwise, the numbers corresponding to the doubtful characters are entered, i.e. 4, 5 in the example of Figure 27.

After a carriage return, the computer takes each character identified by the user and displays the options available (i.e. takes each drawer for which the user knows the content and displays the choice of content it can have). At the end of the character display, the option observed in the sample to be identified is chosen and its number entered (multiple options can be entered where necessary). The computer then moves to the next character, until all the options of the characters chosen at the start have been correctly entered.

The programme is then "running" or "SEARCH" as described in the flow chart (3.2.) and taxa identified are retrieved on the VDU or the printer.

3.4.2. Examples.

Selected examples of cases which were either investigated or brought to the attention of the Forensic Science Unit at Strathclyde University demonstrate the use and advantages which this programme can bring to the efficient management of mycotoxicological emergencies.

3.4.2.1. Case 1.

On the 25th of July 1979, a young girl, Emily, 2½ was admitted to a London Hospital after her mother had seen her eat a mushroom from the garden lawn. No signs of poisoning were presented. Specimen from the lawn were brought to the toxicologist of the hospital who immediately contacted the Forensic Science Unit.

While on the phone, details of the spores were obtained, they were dark, thick-walled, rough, 13 - 14 μ m long and 7 - 9 μ m wide. The toxicologist could not give further characters as it was the first time he had encountered such a sample. Data for 8 characters were available:

- 1) geographical distribution : England
- 2) habitat: grass
- 3) time of growth : summer

4) spore colour under the microscope: dark

the colour was resistant in 50 % H₂SO₄

5) spore wall thickness : thick

6) spore surface : rough

7) spore length : 13 - 14 μ m

8) spore breadth : 7 - 9 μ m

A close matching on the spore wall thickness was requested in view of the inexperience of the analyst.

Out of 223 species making up the data bank, only one species matched the data perfectly : Panaeolus foenisecii.

This species is not dangerous, but can cause hallucinations and up-set if more than 5 or 6 carpophores are ingested. This diagnosis was later confirmed when the samples were sent to the laboratory for further investigations. In the meantime, the little girl had been kept under observation for 24 hours and released without showing any ill-effects.

This shows all the advantages of the present programme at its best: species identification in a matter of a few minutes based on simple information gathered by a non-specialist. (Figure 28).

/F1

FUNGI IDENTIFICATION PROGRAM

- 1 SYMPTOMS
- 2. ONSET OF SYMPTOMS
- 3 GEOG DISTRIBUTION
- 4 HABITAT
- 5 TIME OF GROWTH
- 6 SPORE COLOUR UNDER M. SCOPE
- 7 SPORE SHAPE
- 8. SPORE SURFACE
- 9 SPORE WALL THICKNESS
- 10 GERM POPE
- 11 SPORE LENGTH
- 12 SPORE BPEADTH
- 13. SPORE PRINT
- 14. TOXINS
- 15. DURATION OF SYMPTOMS
- 16. PILEUS TRAMA
- 17 GILL TRAMA
- 18. CUTICLE
- 19. CLAMP CONNECTION
- 20. BASIDIA/ASCI NO. OF SPORES
- 21. BASIDIA/ASCI SIZES
- 22. MARGINAL CYSTIDIA
- 23. FACIAL CYSTIDIA
- 24. KOH CHRYSO CYSTIDIA

WHICH CHARACTERS DO YOU HAVE DATA FOR?
 ENTER CHARACTER NUMBERS COMMA SEPARATED.
 =7, 4, 5, 6, 9, 8, 11, 12

DATA FOR 8 CHARACTERS REQUIRED

IF NO SPECIES MATCH EXACTLY THE GIVEN DATA A CLOSE MATCH
 CAN BE ATTEMPTED ON THE FOLLOWING CHARACTERS:-

- 4. HABITAT
- 5. TIME OF GROWTH
- 9 SPORE WALL THICKNESS
- 11. SPORE LENGTH
- 12 SPORE BREADTH

PLEASE GIVE THE CHARACTER NUMBERS YOU REQUIRE (COMMA SEPARATED)
 OR NONE IF NO CLOSE MATCHING IS TO BE ATTEMPTED

=9
 CLOSE MATCHING REQUIRED ON CHARACTERS 9,

3 GEOGRAPHICAL DISTRIBUTION

- 1. N TEMPERATE
- 2. S. TEMPERATE
- 3. SUBTROPICAL
- 4. TROPICAL
- 5. ENGLISH
- 6. SCOTTISH

OPTION NUMBERS?
 ENTER DATA COMMA SEPARATED

=5
 OPTIONS ACCEPTED --
 5. ENGLISH

4. HABITAT

- 1 GROUND IN CONIF WOODS
- 2 GROUND IN HARDWOODS
- 3 GROUND IN GRASS
- 4. GROUND IN DUNG
- 5. ON STUMPS
- 6. OTHERS

OPTION NUMBERS?
 ENTER DATA COMMA SEPARATED

=3
 OPTIONS ACCEPTED --
 3. GROUND IN GRASS

5 TIME OF GROWTH

- 1. SPRING
- 2 SUMMER
- 3 AUTUMN
- 4. WINTER

OPTION NUMBERS?
 ENTER DATA COMMA SEPARATED

=2
 OPTIONS ACCEPTED --
 2 SUMMER

6. SPORE COLOUR UNDER MICROSCOPE

- 1. CLEAR
- 2. PALE BROWN
- 3. PALE PINK
- 4. DARK

OPTION NUMBER?

=4

OPTION 4 -- DARK

SPORES IN SULPHURIC ACID

- 1. COLOUR RESISTANT
- 2. COLOUR DISSOLVES

OPTION NUMBER?

=1

OPTION 1 -- COLOUR RESISTANT

8. SPORE SURFACE

- 1. SMOOTH
- 2. ROUGH
- 3. RIDGE ORNAMENTATION

OPTION NUMBER?

=2

OPTION 2 -- ROUGH

9. SPORE WALL THICKNESS

- 1. THIN
- 2. THICK

OPTION NUMBER?

=2

OPTION 2 -- THICK

11. SPORE LENGTH(MICRONS)

- 1. LENG < 6.0
- 2. 6.0 < LENG < 8.0
- 3. 8.0 < LENG < 10.0
- 4. 10.0 < LENG < 12.0
- 5. 12.0 < LENG < 17.0
- 6. 17.0 < LENG

OPTION NUMBERS?

ENTER DATA COMMA SEPARATED

=5

OPTIONS ACCEPTED --

- 5. 12.0 < LENG < 17.0

12. SPORE BREADTH(MICRONS)

- 1. BR < 4.0
- 2. 4.0 < BR < 6.0
- 3. 6.0 < BR < 8.0
- 4. 8.0 < BR < 10.0
- 5. 10.0 < BR

OPTION NUMBERS?

ENTER DATA COMMA SEPARATED

=3.4

OPTIONS ACCEPTED --

- 3. 6.0 < BR < 8.0
- 4. 8.0 < BR < 10.0

PROGRAM RUNNING

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
 PANAEOLUS FOENISECII (PERS EX FR)MRE UK FREQ 13.5 INDOLE HALLUC. PROPOSED
 TREATMENT. PREVENT ABSOPP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUI
 POSSIB DIAZEPAM, CHLORPROMAZINE TAXON OLA'H (1969) GENRE PANAEOLUS, REY MYC
 EN HORS SERIE CHEM HOFMANN ET AL(1959) HELY CHIM ACTA 42, 1557-1572
 ED LINCOFF & MITCHEL(1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND
 Y HALLUCINOGEN. UPSETTING COSMOPOLITAN

END OF PROGRAM

*

Figure 28 : Illustration of a Species Identification
 in the Example Case 1.

3.4.2.2. Case 2.

Three patient were admitted to the Royal Infirmary in Edinburgh on the 25th of August 1979. They showed symptoms of severe renal failure some ten days after eating a dish of wild mushrooms purported to be Cantharellus cibarius (a celebrated delicacy in most of Europe). They had been collected in a coniferous wood near their camping place in the North of Scotland (Watling, 1980; Short et al. 1980). Preliminary gastrointestinal disturbances had appeared 2 to 3 days after ingestion.

The identification was made by Dr. Roy Watling, Senior Principal Scientific Officer of the Royal Botanic Garden in Edinburgh, as Cortinarius speciosissimus after a specimen had been collected from the site by the consultant medical specialist. This mushroom is a rare nordic species containing the deadly orellanine. On the basis of the elements available at the time of the emergency and using the present programme, one species would have been keyed out : Cortinarius speciosissimus with two other species Cortinarius gentilis and Cortinarius limonius also suspected of containing orellanine and a number of other species of Cortinarius which may also contain orellanine, but so far, no firm evidence is available

(Greer, 1980). With only five characters used for identification, i.e. symptoms, onset of symptoms, geographical distribution, time of growth and habitat, these species cannot be distinguished. It is quite remarkable that they are so closely related that only an experienced mycologist can distinguish between them, provided he has access to a good specimen!

It is also worth noting, that all these species are suspected to contain the same toxin, require the same treatment and have the same medical and chemical reference literature which is all a medical consultant needs for any clinical treatment. (Figure 29).

3.4.2.3. Case 3.

This is an example where a near-miss match is necessary. It is hypothetical since that facility has not been required in cases handled so far. The hypothesis is based on thorough field observations.

Two teenagers are admitted to hospital showing dizziness, drunkenness, incoordination, staggering, visions, cramps. One hour before the symptoms developed, they had eaten some mushrooms collected on a hill-side pasture near Pitlochry. Five characters can be used

01

FUNGI IDENTIFICATION PROGRAM

- | | |
|-------------------------------|--------------------------------|
| 1 SYMPTOMS | 13. SPORE PRINT |
| 2 ONSET OF SYMPTOMS | 14. TOXINS |
| 3. GEOG. DISTRIBUTION | 15. DURATION OF SYMPTOMS |
| 4. HABITAT | 16. PILEUS TRAMA |
| 5 TIME OF GROWTH | 17. GILL TRAMA |
| 6 SPORE COLOUR UNDER M. SCOPE | 18. CUTICLE |
| 7 SPORE SHAPE | 19. CLAMP CONNECTION |
| 8 SPORE SURFACE | 20. BASIDIA/ASCI NO. OF SPORES |
| 9. SPORE WALL THICKNESS | 21. BASIDIA/ASCI SIZES |
| 10. GERM PORE | 22. MARGINAL CYSTIDIA |
| 11. SPORE LENGTH | 23. FACIAL CYSTIDIA |
| 12 SPORE BREADTH | 24 KOH CHRYSO CYSTIDIA |

WHICH CHARACTERS DO YOU HAVE DATA FOR?
ENTER CHARACTER NUMBERS COMMA SEPARATED
=1,2,3,4,5

DATA FOR 5 CHARACTERS REQUIRED

IF NO SPECIES MATCH EXACTLY THE GIYEN DATA A CLOSE MATCH
CAN BE ATTEMPTED ON THE FOLLOWING CHARACTERS:-

- 4. HABITAT
- 5. TIME OF GROWTH
- 9. SPORE WALL THICKNESS
- 11. SPORE LENGTH
- 12. SPORE BPEARATH

PLEASE GIVE THE CHARACTER NUMBERS YOU REQUIRE (COMMA SEPARATED)
OR 0 (ZERO) IF NO CLOSE MATCHING IS TO BE ATTEMPTED
=0
NO CLOSE MATCHING REQUIRED

1 SYMPTOMS

- 1 GASTROENTERITIS AFTER 3HRS. NO FEVER, CYTOLITIC HEPATITIS
- 2. MIN 15 HOURS AFTER INGESTION: GASTROENTERITIS INTENSE THIRST, POLYURIA THEN RENAL FAILURE
- 3 AFTER ALCOHOL CONSUMPTION. FLUSHING FACE AND NECK, SWELLING, TINGLING OF HANDS, METALLIC TASTE, PALPITATIONS, HYPOTENSION, LATER NAUSEA AND SWEATING
- 4. PERSPIRATION, SALIVATION, LACRYMATION, BLURRED VISION, ABDOMINAL CRAMPS, AFTER DIARRHOEA, CONSTRICTION OF PUPILS, FALL IN BLOOD PRESSURE, SLOW PULSE
- 5. DIZZINESS, DRUNKENNESS, INCOORDINATION, STAGGERING, MUSCULAR CRAMPS, SPASMS, HYPERKINETIC ACTIVITY, DEEP SLEEP AND VISIONS
- 6. MOOD CHANGING, UNMOTIVATED LAUGHTER HILARITY, MUSCLE WEAKNESS DROWSINESS, HALLUCINATIONS
- 7. GASTROENTERITIS; VOMITING
- 8. NAUSEA, GASTROENTERITIS, FEYER, CRAMPS, HEADACHES, CYTOLITIC OR HEMOLYTIC ACTION

OPTION NUMBER?

=2

OPTION 2

2 ONSET OF SYMPTOMS

- 1 WITHIN 2HRS.
- 2. 3 - 7 HRS.
- 3. 8 - 14 HRS.
- 4 AFTER 15 HRS.

OPTION NUMBER?

=4

OPTION 4 -- AFTER 15 HRS.

Figure 29: Example of Identification. Case 2.

GEOGRAPHICAL DISTRIBUTION

- 1 N TEMPEPATE
- 2 S. TEMPERATE
- 3 SUBTROPICAL
- 4 TROPICAL
- 5 ENGLISH
- 6 SCOTTISH

OPTION NUMBERS?
 ENTER DATA COMMA SEPARATED
 =5
 OPTIONS ACCEPTED --
 6 SCOTTISH

4 HABITAT

- 1 GROUND IN CONIF. WOODS
- 2 GROUND IN HARDWOODS
- 3 GROUND IN GRASS
- 4 GROUND IN DUNG
- 5 ON STUMPS
- 6 OTHERS

OPTION NUMBERS?
 ENTER DATA COMMA SEPARATED
 =1
 OPTIONS ACCEPTED --
 1 GROUND IN CONIF WOODS

5. TIME OF GROWTH

- 1 SPRING
- 2. SUMMER
- 3. AUTUMN
- 4. WINTER

OPTION NUMBERS?
 ENTER DATA COMMA SEPARATED
 =2
 OPTIONS ACCEPTED --
 2. SUMMER

PROGRAM RUNNING

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
 COPTINARIUS SPECIOSISSIMUS KUHN. & ROMAG. ORELLANINE TREATMENT. SYMPTOMAT
 TAXON. ORTON PD (1958) COPTINARIUS II THE NATURALIST, APRIL, 119
 CHEM. ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21, 1931-34
 MEDIC. GERVAULT & GIRPE (1977) BUL. SOC. MYCOL. FRANCE 93, 373-405
 DEADLY EU & N EU

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
 COPTINARIUS GENTILIS (FR)FR UK FREQ 0.4 SUSPECT ORELLANINE PROPOSED TREA
 TMENT SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCH
 STUT NY CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21, 1931-34 MED FL
 R (1980) DIFFERENTIALDIAGNOSE DER PILZVERGIFTUNGEN, FISCHER, STUTT, NY
 DEADLY EU
 **MORE(YES OR NO)?
 =YES

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
 COPTINARIUS LIMONIUS (FR EX FR)FR ORELLANINE SUSPECT PROPOSED TREATMENT
 SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER,
 STUT NY CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21, 1931-34 MED FL
 (1980) DIFFERENTIALDIAGNOSE DER PILZVERGIFTUNGEN, FISCHER, STUTT, NY
 SUSPECT DEADLY. EU

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
 COPTINARIUS CALLISTEUS (FR)FR UK FREQ 0.4 SUSPECT ORELLANINE TREATMENT.
 SYMPTOMATIC TAXON MOSEP M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER,
 STUT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934
 MED GERVAULT & GIRPE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
 **MORE(YES OR NO)?
 =YES

Figure 29: continued.

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
CORTINARIUS CINNAMOMEUS (L EX FR) UK FREQ 2.0 SUSPECT ORELLANINE
TREATMENT SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB/2
FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979)TETRAHEDRON L 21,1931-1934
MED GERALT & GIRRE (1977)BUL SOC MYCOL FR 93,373-405 DEADLY? EU N. AM

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
CORTINARIUS PHOENICEUS (BUL)MRE SUSPECT ORELLANINE TREATMENT.SYMPTOMATIC
TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB/2,FISCHER, STUTT.
CHEM ANTKOWIAK & GESSNER (1979)TETRAHEDRON L 21,1931-1934 MED GERALT &
GIRRE (1977)BUL SOC MYCOL FR 93,373-405 DEADLY? EU
**MORE(YES OR NO)?
=YES

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
CORTINARIUS SANGUINEUS (WULF EX FR)FR UK FREQ 0.4 SUSPECT ORELLANINE
TREATMENT:SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB,2
FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979)TETRAHEDRON L 21,1931-34
MED GERALT & GIRRE (1977)BUL SOC MYCOL FR 93,373-405 DEADLY? EU

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
CORTINARIUS SEMISANGUINEUS (FR)GILL UK FREQ 4.0 SUSPECT ORELLANINE
TREATMENT:SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB,2
FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979)TETRAHEDRON L 21,1931-34
MED GERALT & GIRRE (1977)BUL SOC MYCOL FR 93,373-405 DEADLY? EU;N. AM
**MORE(YES OR NO)?
=YES

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
CORTINARIUS TRAGANUS (FR EX FR)FR UK FREQ 1.2 SUSPECT ORELLANINE TREAT-
MENT:SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB,2
FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979)TETRAHEDRON L 21,1931-34
MED GERALT & GIRRE (1977)BUL SOC MYCOL FR 93,373-405 DEADLY? EU

POSSIBLE SPECIES(EXACT MATCH ON SUPPLIED DATA) --
CORTINARIUS SANIOSUS (FR)FR UK FREQ 0.4 SUSPECT ORELLANINE TREATMENT:
SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB,2 FISCHER
STUTT CHEM ANTKOWIAK & GESSNER (1979)TETRAHEDRON L 21,1931-34 MED
GERALT & GIRRE (1977)BUL SOC MYCOL FR 93,373-405 DEADLY? EU

END OF PROGRAM

Figure 29 (end).

here (the same as in case 2). But how would a pasture be described using the habitat's options? "grass", "dung" or are there some tree in the vicinity associated with the mushrooms? Therefore the "near-miss match" facility is used with the character "habitat" and the option chosen is "grass".

No exact match is found, but three species, Amanita muscaria, A. pantherina and A. gemmata are identified as a "close match on habitat" (Figure 30) i.e. these are possible identifications, provided the state of the character "habitat" is not "grass". Field observations (detailed in chapter 4) have shown the presence of Amanita pantherina in hill-side pastures near Pitlochry (reference collections 121 and 244, see chapter 4) amongst Salix repens (a dwarf shrub of the willow family, i.e. hardwoods). An area which could be described by many as "grassy"!

It is obvious that in the case of an emergency, the results are rapid and accurate enough to allow clinicians to proceed with treatment (here again identical for the three species). Further investigations, in the field, on stomach washing and on the remains of the meal, etc, can be carried out afterwards for a more specific identification.

7F1

FUNGI IDENTIFICATION PROGRAM

- | | |
|-------------------------------|--------------------------------|
| 1 SYMPTOMS | 13. SPORE PRINT |
| 2 ONSET OF SYMPTOMS | 14 TOXINS |
| 3 GEOG DISTRIBUTION | 15. DURATION OF SYMPTOMS |
| 4. HABITAT | 16. PILEUS TRAMA |
| 5 TIME OF GROWTH | 17. GILL TRAMA |
| 6 SPORE COLOUR UNDER M. SCOPE | 18. CUTICLE |
| 7 SPORE SHAPE | 19. CLAMP CONNECTION |
| 8 SPORE SURFACE | 20. BASIDIA/ASCI NO. OF SPORES |
| 9 SPORE WALL THICKNESS | 21. BASIDIA/ASCI SIZES |
| 10 GERM PORE | 22. MARGINAL CYSTIDIA |
| 11 SPORE LENGTH | 23. FACIAL CYSTIDIA |
| 12. SPORE BREADTH | 24 KOH CHRYSO CYSTIDIA |

WHICH CHARACTERS DO YOU HAVE DATA FOR?
ENTER CHARACTER NUMBERS COMMA SEPARATED
=1, 2, 3, 4, 5

DATA FOR 5 CHARACTERS REQUIRED

IF NO SPECIES MATCH EXACTLY THE GIVEN DATA A CLOSE MATCH
CAN BE ATTEMPTED ON THE FOLLOWING CHARACTERS:-

- 4. HABITAT
- 5. TIME OF GROWTH
- 9. SPORE WALL THICKNESS
- 11. SPORE LENGTH
- 12. SPORE BREADTH

PLEASE GIVE THE CHARACTER NUMBERS YOU REQUIRE (COMMA SEPARATED)
OR 0 (ZERO) IF NO CLOSE MATCHING IS TO BE ATTEMPTED

=4
CLOSE MATCHING REQUIRED ON CHARACTERS 4,

1 SYMPTOMS

- 1. GASTROENTERITIS AFTER 8HRS. NO FEVER, CYTOLITIC HEPATITIS
- 2 MIN 15 HOURS AFTER INGESTION: GASTROENTERITIS INTENSE THIRST, POLYURIA THEN RENAL FAILURE
- 3 AFTER ALCOHOL CONSUMPTION. FLUSHING FACE AND NECK, SWELLING, TINGLING OF HANDS; METALLIC TASTE, PALPITATIONS, HYPOTENSION, LATER NAUSEA AND SWEATING
- 4. PERSPIRATION, SALIVATION, LACRYMATION, BLURRED VISION, ABDOMINAL CRAMPS, AFTER DIARRHOEA, CONSTRICTION OF PUPILS, FALL IN BLOOD PRESSURE, SLOW PULSE
- 5 DIZZINESS, DRUNKENNESS, INCOORDINATION, STAGGERING, MUSCULAR CRAMPS, SPASMS, HYPERKINETIC ACTIVITY, DEEP SLEEP AND VISIONS
- 6 MOOD CHANGING. UNMOTIVATED LAUGHTER HILARITY, MUSCLE WEAKNESS DROWSINESS, HALLUCINATIONS
- 7 GASTROENTERITIS; VOMITING
- 8 NAUSEA, GASTROENTERITIS, FEVER, CRAMPS, HEADACHES, CYTOLITIC OR HEMOLYTIC ACTION

OPTION NUMBER?

=5

OPTION 5

2 ONSET OF SYMPTOMS

- 1. WITHIN 2HRS.
- 2. 3 - 7 HRS.
- 3. 8 - 14 HRS.
- 4. AFTER 15 HRS.

OPTION NUMBER?

=1

OPTION 1 -- WITHIN 2HRS.

Figure 30 : Example of an Identification Using

the "Near-miss Match" Facility. Case 3.

3 GEOGRAPHICAL DISTRIBUTION

- 1. N. TEMPERATE
- 2. S. TEMPERATE
- 3. SUBTROPICAL
- 4. TROPICAL
- 5. ENGLISH
- 6. SCOTTISH

OPTION NUMBERS?

ENTER DATA COMMA SEPARATED

=6

OPTIONS ACCEPTED --

- 5 SCOTTISH

4 HABITAT

- 1. GROUND IN CONIF. WOODS
- 2. GROUND IN HARDWOODS
- 3. GROUND IN GRASS
- 4. GROUND IN DUNG
- 5. ON STUMPS
- 6. OTHERS

OPTION NUMBERS?

ENTER DATA COMMA SEPARATED

=3

OPTIONS ACCEPTED --

- 3. GROUND IN GRASS

5 TIME OF GROWTH

- 1. SPRING
- 2. SUMMER
- 3. AUTUMN
- 4. WINTER

OPTION NUMBERS?

ENTER DATA COMMA SEPARATED

=2

OPTIONS ACCEPTED --

- 2. SUMMER

PROGRAM RUNNING

THE FOLLOWING SPECIES MATCHES CLOSELY ON HABITAT

AMANITA PANTHERINA (FR.) SECR. UK FREQ 2.4 MUSCINOL/IBOTENIC ACID PROP. TREATMENT: PREVENT ABSORPTION, INCR. EXCRETION, SYMPTOMATIC (PHYSOSTIGMIN)
 TAXON. JENKINS DT (1977) BIBLIO. MYCOLOGICA 57, 1-126
 CHEM. CATALFOMO & EUGSTER (1970) BULL. NARC. 22:4, 33-41
 MEDIC. WASER PG (1967) ETHNOPHARMAC. SEARCH .. ED. EFRON US PHSP 1645, 41
 INEBRIANT. DANGEROUS EU; N. AM

THE FOLLOWING SPECIES MATCHES CLOSELY ON HABITAT

AMANITA MUSCARIA (L EX FR) HOOKER UK FREQ 12.3 MUSCINOL/IBOTENIC ACID TR TMENT: PREVENT ABS., INCREASE EXCRETION, SYMPTOMATIC (PHYSOSTIGMINE)
 TAXON. JENKINS DT (1977) BIBLIOTHECA MYCOLOGICA 57, 1-126
 CHEM. CATALFOMO & EUGSTER (1970) BULL. NARC. 22:4, 33-41
 MEDIC. WASER PG (1967) ETHNOPHARMAC SEARCH... ED EFRON, US PHSP 1645, 413-3
 INEBRIANT, DANGEROUS EU; ASIA; N. AM.

THE FOLLOWING SPECIES MATCHES CLOSELY ON HABITAT

**MORE (YES OR NO)?
 =YES

AMANITA GEMMATA (FR) BERT. EUROPE. EDIBLE, US: SUSPECT IBOTENIC A/MUSCINOL S. AM: DEADLY! TREATMENT: SYMPTOMATIC IF SYMPTOMS ONSET > 8 HOURS + WATER, ELEC-TROLYTES, BLOOD BALANCE, HYPERTONIC GLUC. INF. PENICILLIN, THIOCTIC ACID, EXCH TRANSF. TAXON JENKINS DT (1977) BIBLIOT. MYCOL 57, 1-126 CHEM CHILTON & OTT (1976) LLOYDIA 39:2, 3, 150-157 MED. SEE AMANITA PHALLOIDES FOR S. AM. EU; N.

NO SPECIES MATCH SUPPLIED DATA EXACTLY

END OF PROGRAM

Figure 30 (end).

3.4.2.4. Identification of Mixtures.

Where mixtures are concerned, the problem is more complex and good sense has to be applied if an optimum result is to be obtained.

If only one species in the mixture is toxic, the problem is relatively simple - i.e. one type of medical and chemical characters is observed - the ecological characters should also be known, at least in relation to the time of growth and the geographical distribution. If microscopical characters are present, different features from different species will be present and mixed. If all the data are used together, they may describe a non-existent and quite extraordinary mushroom! The study, therefore, has to be limited to a single type of microscopical characters, the obvious choice being the 8 characters based on the spore (i.e. spore colour under the microscope, shape, surface, wall thickness, germ pore, length, breadth and print): they can all be observed on one single spore which clearly comes from a single species. Where mixtures are concerned, different spores are observed and each type of spore should be studied separately with the other characters (medical, chemical or ecological) and relevant results obtained

Mixtures of two or more toxic species may create more difficult problems which can hardly be solved at the present time, even with a team of specialists because the cumulative effects of different toxins administered together are not known. A separate study of different spores, with possibly some discernable clinical symptoms, may lead to useful results, but no experimental data is available so far and further discussion would be purely speculative.

Microscopical characters other than the spores could be used in this context in a minority of cases, when a particularly striking feature is observed, such as a special type of cystidia. The choice of characters to be used in such cases is to be left to the best judgement of the user of the key.

CHAPTER IV : DATA COLLECTION

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IV. DATA COLLECTION.

4.1. Poisonous and Hallucinogenic Mushrooms: a World List.

The world list of poisonous and hallucinogenic mushrooms (Table 3) has been compiled from literature sources of all types, ranging from major works mentioned earlier in this thesis to the most obscure underground publications. A number of comments are required when one sees such a list of 432 names (Table 3).

Firstly, some obvious and well known poisonous mushrooms seem to be absent from the list. This is exemplified by Lepiota morgani, Lepiota helveola and Entoloma lividum. These names are widely used in the literature, but are invalid or illegal taxonomical names as described by British authorities (Dennis et al. 1960). Their correct and/or accepted names are used, i.e. Chlorophyllum molybdites, Lepiota brunneoincarnata and Entoloma sinuatum respectively.

The second comment stems from the size of the list. Most textbooks mention five to ten deadly poisonous mushrooms and possibly another 40 to 50 species being in some degree dangerous out of approximately 10'000 mushroom species described. These latest figures would

seem perfectly realistic to the present author and certainly the number of poisonous species would not go much above the hundred mark if all the listed species were to be thoroughly investigated. This discrepancy can be readily explained if one takes into account that the literature survey relates to "mycophagic" as well as notoriously "mycophobic" countries. Some species have only once been reported to be poisonous and this as long ago as the end of the last century. Such reports are subsequently repeated in modern books without any further checks. Most of the time voucher specimens have not been kept and it is impossible to be certain that the mushroom identification, made at the time, was accurate. When one sees how dubious some of the identifications were in Switzerland between 1966-1976, a country well organised in that field (Bornet, 1980), it would not be surprising to learn that a number of these records of toxicity are unfounded.

Mushrooms are notoriously difficult to digest, and some individuals can easily suffer from indigestion with them. Some mushrooms are known to contain antibiotics (e.g. Armillaria mellea) and susceptible people have been known to suffer from allergic reactions. By contrast, if, for example, one suffers an allergic reaction to strawberries, nobody will start thinking,

never mind publishing, that strawberries are toxic. Unfortunately, this is what has happened with mushrooms. In mycophobic countries, because of the special historical status that mushrooms have occupied as being "evil, putrid, etc" (Wasson, 1968), it is easy to understand that they have readily been assumed to be toxic.

Other species, still reported to be poisonous by some, such as Amanita citrina, have been shown to be innocuous. A. citrina was believed to be a form of the related Death Cap (Amanita phalloides) until extensive experiments by Chauvin (1922) in the 1920's showed otherwise, but its smell and flavour of old potatoes do not make it palatable.

Some other species are claimed to be poisonous mainly because of their foul taste and/or smell. Cultural, or rather gastronomical differences, can be seen even within European communities. Species reported as suspect in Britain are used as spices and condiments in other countries (Italy, Hungary, etc). This is true for many acrid or peppery Russula and Lactarius, which with the exception of a few notorious species are not poisonous. In this respect, literature from one country sometimes indicates a species as good and edible, whereas it is reported as poisonous and suspect in another (e.g. Amanita

excelsa has a two "knife and fork" crosses in Kühner and Romagnesi (1953), a standard and celebrated French reference book, whereas in British guides it ranges from harmless but best avoided, to poisonous!

Another comment relates to the hallucinogenic species. Since Hofmann et al. (1958) discovered psilocybin in a Mexican mushroom, Psilocybe mexicana, literally hundreds of mushrooms have been claimed to be hallucinogenic, often without any supporting chemical evidence other than the blueing flesh of the specimens described. These claims, when critically examined, might prove to be correct, but until then, doubts remain. Some analyses on newly described Australian species demonstrate this point (Margot & Watling, 1980). Out of four blueing species of Psilocybe submitted for analysis, only two proved positive for the presence of psilocybin (see 4.2.3.) Recently, critical reviews by Guzmán (1978) on the genus Psilocybe have also shown that many hallucinogenic species, described as new separate species in the last twenty years, are in fact synonymous (e.g. Psilocybe zapotecorum = P. candidipes = P. bolivarii).

Lastly, a number of species are excellent when consumed, provided they are first cooked, dried or parboiled and the water discarded. This is important

because they contain heat labile and/or water soluble toxins. This is the case of Amanita rubescens, Gyromitra esculenta, Morchella species, etc, all regularly eaten by the present author and his friends without ill-effects!

Many other species listed have been eaten in reasonable quantities by the author over the past four years without ever experiencing even the slightest tummy up-set. (Agaricus silvicola, Amanita rubescens, A. vaginata, Clitocybe clavipes, C. nebularis, Coprinus atramentarius, Gyromitra esculenta, Hygrophoropsis aurantiaca, Morchella esculenta, Pholiota squarrosa (tough!), Suillus granulatus, Tricholomopsis rutilans).

It is quite clear from what precedes that the size of the above world list of poisonous or suspect mushrooms is grossly inflated, mainly because of prejudice, and that extensive toxicological work is required to reduce that list to truly poisonous species.

On the other hand, some yet undescribed toxic species may well come to light in the next few years as floras of Africa, Asia and the Australasian continent are completed.

TABLE 3 : World List of Mushrooms Known or Suspected to be Poisonous with an Indication of the Symptoms they Elicit, their Toxins (Where Known) and their Distribution in Britain, when Found in Britain.

Abbreviations: - Symptoms: AP = Amanita Poisoning CG = Cortinarius Poisoning

MU = Muscarinic IP = Ibotenic acid/Muscimol Poisoning

HA = Hallucinogenic GE = Gastroenteritic

GP = Gyromitrin Poisoning CP = Coprine Poisoning

Toxins : CY = Cyclopeptides OR = Orellanine CO = Coprine

GY = Gyromitrin MC = Muscarin IM = Ibotenic Acid/Muscimol

PS = Psilocybin and Analogs. A question mark indicates that the toxin is suspected or unknown.

A symptom followed by a question mark indicates a doubtful toxicity.

Distribution : B = British E = English and Welsh S = Scottish

Names and Authors : as accepted by British Authorities (Dennis, Orton and Hora, 1960)

Species Symptoms Toxin Distribution

<u>Agaricus</u> <u>albolutescens</u> Zeller	GE	?	
<u>A.</u> <u>hondensis</u> Murr.	GE	?	
<u>A.</u> <u>perdicina</u> Pilát	GE	?	
<u>A.</u> <u>phaelepidotus</u> (Møll.) Møll.	GE	?	E
<u>A.</u> <u>placomycetes</u> Peck	GE	?	
<u>A.</u> <u>silvicola</u> (Vitt.) Peck	GE	?	B
<u>A.</u> <u>xanthodermus</u> Gen.	GE	?	B
<u>Amanita</u> <u>alliiodora</u> Pat.	AP?	CY?	
<u>A.</u> <u>ameghinoi</u> (Speg.) Sing.	?	?	
<u>A.</u> <u>aspera</u> (Fr.) Gray	?	?	E
<u>A.</u> <u>bingensis</u> (Beeli) Heim	AP?	CY?	
<u>A.</u> <u>bisporigera</u> Atk.	AP	CY	
<u>A.</u> <u>brunnescens</u> Atk.	GE?	?	
<u>A.</u> <u>chlorinosma</u> (Peck apud Aust.) Lloyd	GE?	?	

Species Symptoms Toxin Distribution

<u>Amanita citrina</u> Gray	HA?	PS?	B
<u>A. cokeri</u> (Gilb. & Kühn.) Gilb.	IP?	IM?	
<u>A. cothurnata</u> Atk.	IP	IM	
<u>A. crenulata</u> Peck	IP?	IM?	
<u>A. echinocephala</u> (Vitt.) Qué1.	?	?	E
<u>A. excelsa</u> (Fr.) Kum.	?	?	B
<u>A. flavoconia</u> Atk.	GE?	?	
<u>A. flavorubescens</u> Atk.	GE?	?	
<u>A. frostiana</u> (Peck) Sacc.	IP?	IM?	
<u>A. gemmata</u> (Fr.) Gill.	?	?	B
<u>A. muscaria</u> (L. ex Fr.) Hook.	IP	IM	B
<u>A. ocreata</u> Peck	AP	CY	
<u>A. pantherina</u> (DC ex Fr.) Secr.	IP	IM	B
<u>A. parvicolvata</u> (Peck) Gilb.	GE?	?	

Species	Symptoms	Toxin	Distribution
<u>Amanita phalloides</u> (Vaill. ex Fr.) Secr.	AP	CY	B
A. <u>porphyria</u> (Alb. & Schw. ex Fr.) Secr.	?	?	B
A. <u>preissei</u> (Fr.) Sacc.	GE?	?	
A. <u>regalis</u> (Fr.) Sacc.	IP	IM	S
A. <u>robusta</u> Beeli	AP?	CY?	
A. <u>rubescens</u> ((Pers.) Fr.) Gray	hemolytic	?	B (only raw)
A. <u>smithiana</u> Bas	IP	IM	160
A. <u>solitaria</u> (Bull. ex Fr.) Secr.	IP?	IM?	B
A. <u>spinosa</u> Bouriquet	AP?	CY?	
A. <u>spretata</u> Peck	GE?	?	
A. <u>suballiacea</u> (Murr.) Murr.	AP	CY	
A. <u>tainaomy</u> Heim	AP?	CY?	
A. <u>tenuifolia</u> Murr.	AP	CY	
A. <u>vaginata</u> (Bull. ex Fr.) Vitt.	hemolytic	?	B (only raw)

Species	Symptoms	Toxin	Distribution
<u>Amanita</u> <u>verna</u> (Bull. ex Fr.) Vitt.	AP	CY	B
<u>A.</u> <u>virosa</u> Secr.	AP	CY	B
<u>A.</u> <u>vittadini</u> (Mor.) Vitt.	?	?	
<u>A.</u> <u>volvata</u> (Peck) Martin	GE?	?	
<u>Anellaria</u> (<u>Panaeolus</u>) <u>phalaeoarum</u> (Fr.) Quéf.	HA	PS?	
<u>A.</u> <u>semiovata</u> (Sow. ex Fr.) Pears. & Dennis	HA?	PS?	B
<u>Boletus</u> <u>calopus</u> Fr.	MU	MC?	B
<u>B.</u> <u>eastwoodiae</u> (Murr.) Sacc. & Trotter	GE?	?	
<u>B.</u> <u>kumaeus</u> Heim	HA	?	
<u>B.</u> <u>luridus</u> Schaeff. ex Fr.	MU	MC?	B
<u>B.</u> <u>manicus</u> Heim	HA	?	
<u>B.</u> <u>miniatoolivaceus</u> Frost	GE?	?	
<u>B.</u> <u>nigerrimus</u> Heim	HA	?	
<u>B.</u> <u>pulcherrimus</u> Thiers & Halling	GE?	?	

Species	Symptoms	Toxin	Distribution
<u>Boletus</u> <u>purpureus</u> Pers.	GE?	?	E
<u>B.</u> <u>queletii</u> Schulz.	GE?	?	E
<u>B.</u> <u>albidus</u> Roques	GE?	?	B
<u>B.</u> <u>reayi</u> Heim	HA	?	
<u>B.</u> <u>rufo-aureus</u> Meyr.	GE?	?	
<u>B.</u> <u>satanas</u> Lenz	GE	?	B
<u>B.</u> <u>splendidus</u> Martin	GE?	?	
<u>B.</u> <u>subvelutipes</u> Peck	GE?	?	
<u>Chlorophyllum</u> <u>molybdites</u> (Meyer ex Fr.) Mass.	GE	?	
<u>Clitocybe</u> <u>acromelalga</u> Ichimura	MU	MC?	
<u>C.</u> <u>angustissima</u> (Lasch.) Kum.	MU	MC	E
<u>C.</u> <u>candicans</u> (Pers. ex Fr.) Kum.	MU	MC	B
<u>C.</u> <u>cerussata</u> (Fr.) Gill.	MU	MC	B
<u>C.</u> <u>clavipes</u> (Pers. ex Fr.) Kum.	CP?	CO?	B

Species	Symptoms	Toxin	Distribution
<u>Clitocybe dealbata</u> (Sow. ex Fr.)Kum.	MU	MC	B
<u>C. diatreta</u> (Fr.)Kum.	MU	MC?	B
<u>C. ericetorum</u> (Bull.)Quél.	MU	MC?	B
<u>C. festiva</u> Favre	MU	MC	
<u>C. gracilipes</u> Lam.	MU	MC?	
<u>C. marginella</u> Harmaja	MU	MC?	
<u>C. nebularis</u> (Batsch. ex Fr.)Kum.	GE?	?	B
<u>C. nuoljae</u> Lam.	MU	MC	
<u>C. phyllophila</u> (Fr.)Kum.	MU	MC	B
<u>C. rivulosa</u> (Pers. ex Fr.)Kum.	MU	MC	B
<u>C. serotina</u> Lam.	MU	MC	
<u>C. suaveolens</u> (Schum. ex Fr.)Kum.	MU?	MC?	B
<u>C. truncicola</u> (Peck)Sacc.	MU	MC?	
<u>C. venenata</u> Heim	MU	MC?	

Species Symptoms Toxin Distribution

<u>Conocybe cyanopus</u> (Atk.)Kühn.	HA	PS	B(?)
<u>C. filaris</u> (Fr.)Kühn.	AP	CY	B
<u>C. mairei</u> Kühn.	?	?	B
<u>C. siligineoides</u> Heim	HA	PS	
<u>C. smithii</u> Wat.	HA	PS	
<u>Copelandia cambodginiensis</u> Ota'h & Heim	HA	PS	
<u>C. chlorocystis</u> Sing. & Weeks	HA	PS	
<u>C. cyanescens</u> (Berk.& Br.)Sing.	HA	PS	
<u>C. mexicana</u> Guz.	HA?	PS?	
<u>C. papilionacea</u> Malç. & Bertault	HA?	PS?	
<u>C. tropicalis</u> Ota'h	HA	PS	
<u>Coprinus acuminatus</u> (Romag.)Ort.	CP?	CO?	B
<u>C. africanus</u> Peg.	CP?	CO?	
<u>C. atramentarius</u> (Bull. ex Fr.)Fr.	CP	CO	B

Species	Symptoms	Toxin	Distribution
<u>Coprinus</u> <u>erethistes</u> Heim	CP	CO?	
<u>C.</u> <u>insignis</u> Peck	CP?	CO?	E
<u>C.</u> <u>micaceus</u> (Bull. ex Fr.) Fr.	?	CO?	B
<u>C.</u> <u>quadrifidus</u> Peck	CP?	CO?	
<u>C.</u> <u>romagnesianus</u> Sing.	CP?	CO?	B
<u>C.</u> <u>variegatus</u> Peck	CP?	CO?	
<u>Cortinarius</u> <u>aureifolius</u> Peck	?	OR?	E
<u>C.</u> <u>bolaris</u> (Pers. ex Fr.) Fr.	?	OR?	B
<u>C.</u> <u>brunneofulvus</u> Fr. s Bres.	?	OR?	E
<u>C.</u> <u>bulliardii</u> (Fr.) Fr.	?	OR?	E
<u>C.</u> <u>callisteus</u> (Fr.) Fr.	?	OR?	B
<u>C.</u> <u>cinnabarinus</u> Fr.	?	OR?	E
<u>C.</u> <u>cinnamomeofulvus</u> Hry	?	OR?	
<u>C.</u> <u>cinnamomeus</u> (L. ex Fr.) Fr.	?	OR?	B

Species	Symptoms	Toxin	Distribution
<u>Cortinarius cotoneus</u> Fr.	?	OR?	E
<u>C. elegantior</u> Fr. ex Fr.	?	OR?	B
<u>C. fluorescens</u> Horak	?	OR?	
<u>C. gentilis</u> (Fr.) Fr.	CG	OR?	B
<u>C. limonius</u> (Fr. ex Fr.) Fr.	CG	OR?	B
<u>C. malicorius</u> Fr.	?	OR?	B
<u>C. orellanoides</u> Hry	CG	OR?	E
<u>C. orellanus</u> Fr.	CG	OR	E
<u>C. phoeniceus</u> (Bull.) Mre	?	OR?	B
<u>C. puniceus</u> Ort.	?	OR?	E
<u>C. rubicundus</u> (Rea) Pears.	?	OR?	B
<u>C. sanguineus</u> (Wulf. ex Fr.) Fr.	?	OR?	B
<u>C. saniosus</u> (Fr.) Fr.	?	OR?	B
<u>C. semisanguineus</u> (Fr.) Gill.	?	OR?	B

Species Symptoms Toxin Distribution

<u>Cortinarius speciosissimus</u> Kühn. & Romag.	CG	OR	S
<u>C. splendens</u> Hry	CG	OR?	
<u>C. subfusipes</u> Hry	?	OR?	
<u>C. tophaceoides</u> Mos.	?	OR?	
<u>C. tophaceus</u> (Fr.) Fr.	?	OR?	E
<u>C. traganus</u> (Fr. ex Fr.) Fr.	?	OR?	B
<u>C. uliginosus</u> Berk.	?	OR?	B
<u>C. venenosus</u> Kawamura	?	OR?	
<u>Entoloma nidorosum</u> (Fr.) QuéL.	.GE	?	B
<u>E. niphooides</u> Romag.	GE	?	B
<u>E. rhodopolium</u> (Fr.) Kum.	GE	?	B
<u>E. sinuatum</u> (Bull. ex Fr.) Kum.	GE	?	B
<u>E. strictum</u> (Peck) Sacc.	GE	?	
<u>E. vernum</u> Lundell	GE	?	B
+ numerous other species of <u>Entoloma</u>	GE	?	B

Species	Symptoms	Toxin	Distribution
<u>Gyromitra</u> <u>ambigua</u> (Karst.) Harmaja	GP	GY	
<u>G.</u> <u>apiculata</u> (McKnight) Harmaja	GP	GY	
<u>G.</u> <u>brunnea</u> Underwood	GP	GY	
<u>G.</u> <u>californica</u> (Phill.) Raitviir	GP	GY	
<u>G.</u> <u>caroliniana</u> (Bosc. ex Fr.) Fr.	GP	GY	
<u>G.</u> <u>curtipes</u> Fr.	GP	GY	
<u>G.</u> <u>esculenta</u> Pers. ex Fr.	GP	GY	B
<u>G.</u> <u>fastigiata</u> (Krombh.) Rehm.	GP	GY	
<u>G.</u> <u>gigas</u> (Krombh.) Cooke	GP	GY	
<u>G.</u> <u>infula</u> (Schaeff. ex Pers.) Qué1.	GP	GY	B
<u>G.</u> <u>martinii</u> Donadini & Astier	GP	GY	
<u>G.</u> <u>megalospora</u> Donadini & Rioussset	GP	GY	
<u>Hebeloma</u> <u>crustuliniforme</u> (Bull. ex St Amans) Qué1.	GE	?	B
<u>H.</u> <u>fastibile</u> (Pers. ex Fr.) Kum.	GE	?	B

Species	Symptoms	Toxin	Distribution
<u>Hebeloma mesophaeum</u> (Pers.) Quél.	GE	?	B
<u>H. sinapizans</u> (Paulet ex Fr.) Gill.	GE	?	B
+ numerous other suspect <u>Hebeloma</u> species	GE	?	B
<u>Helvella acetabula</u> (Linn. ex St Amans) Quél.	GP	GY?	B
<u>H. crispa</u> Fr.	GP	GY?	B
<u>H. elastica</u> Bull.	GP	GY?	B
<u>H. lacunosa</u> Fr.	GP	GY?	B
<u>H. leucomelanea</u> (Pers.) Nann.	GP	GY?	B
<u>H. sphaerospora</u> (Peck) Imai	GP	GY?	
<u>Hygrocybe conica</u> (Scop. ex Fr.) Kum.	GE	?	B
<u>H. nigrescens</u> (Quél.) Kühn.	GE	?	B
<u>Hygrophoropsis aurantiaca</u> (Fr.) Mre apud Martin-Sans	MU?	MC?	B

Species	Symptoms	Toxin	Distribution
<u>Hypholoma fasciculare</u> (Huds. ex Fr.) Quél.	GE	?	B
<u>H. popperianum</u> Sing.	HA?	PS?	
<u>H. sublateralitium</u> (Fr.) Quél.	GE?	?	B
<u>Inocybe albodisca</u> Kühn.	MU	MC	E
<u>I. asterospora</u> Quél.	MU	MC	B
<u>I. brunnea</u> Quél.	MU	MC	B
<u>I. cincinnata</u> (Fr.) Quél.	MU	MC	B
<u>I. corydalina</u> Quél.	MU?	MC?	E
<u>I. decipientoides</u> Peck	MU	MC	
<u>I. dulcamara</u> (Alb. & Schw. ex Pers.) Kum.	MU	MC	B
<u>I. eutheles</u> (Berk. & Br.) Quél.	MU	MC	B
<u>I. fastigiata</u> (Schaeff. ex Fr.) Quél.	MU	MC	B
<u>I. fibrosa</u> (Sow. ex Berk.) Gill.	MU	MC	E
<u>I. flocculosa</u> (Berk.) Sacc.	MU	MC	B

Species	Symptoms	Toxin	Distribution
<u>Inocybe friesii</u> Heim	MU	MC	B
<u>I. gausapata</u> Kühn.	MU	MC	
<u>I. geophylla</u> (Sow. ex Fr.) Kum.	MU	MC	B
<u>I. griseolilacina</u> Lge	MU	MC	B
<u>I. hirtella</u> Bres.	MU	MC	E
<u>I. hystrix</u> (Fr.) Karst.	MU	MC	B
<u>I. lacera</u> (Fr.) Kum.	MU	MC	B
<u>I. lanuginella</u> (Schr.) Lge	MU	MC	B
<u>I. lanuginosa</u> (Bull. ex Fr.) Kum.	MU	MC	B
<u>I. lucifuga</u> (Fr.) Kum.	MU	MC	E
<u>I. maculata</u> Boud.	MU	MC	B
<u>I. mixtilis</u> (Britz.) Sacc.	MU	MC	B
<u>I. napipes</u> Lge	MU	MC	B
<u>I. oblectabilis</u> (Britz.) Sacc.	MU	MC	E

Species Symptoms Toxin Distribution

<u>Inocybe obscura</u> (Pers. ex Pers.) Gill.	MU	MC	B
<u>I. obscuroides</u> Ort.	MU	MC	E
<u>I. pallidipes</u> Ell. & Ever.	MU	MC	E
<u>I. patouillardii</u> Bres.	MU	MC	E
<u>I. perlata</u> (Cke) Sacc.	MU	MC	E
<u>I. posterula</u> (Britz.) Sacc.	MU	MC	B
<u>I. praetervisa</u> Qué1.	MU	MC	B
<u>I. pudica</u> Kühn.	MU	MC	B
<u>I. queletii</u> Mre & Kühn.	MU	MC	
<u>I. sambucina</u> (Fr.) Qué1.	MU	MC	B
<u>I. terrifera</u> Kühn.	MU	MC	B
<u>I. umbrina</u> Bres.	MU	MC	B
<u>I. xanthomelas</u> Bours. & Kühn.	MU	MC	B

Species Symptoms Toxin Distribution

<u>Lactarius blennius</u> (Fr.)Fr.	GE?	?	B
L. <u>chrysotheus</u> Fr.	GE?	?	B
L. <u>fuliginosus</u> (Fr.)Fr.	GE?	?	B
L. <u>glaucescens</u> Crossland	GE?	?	B
L. <u>helvus</u> (Fr.)Fr.	GE?	?	B
L. <u>lignyotus</u> Fr.	GE?	?	E
L. <u>pyrogalus</u> (Bull. ex Fr.)Fr.	GE?	?	B
L. <u>representanaeus</u> Britz.	GE?	?	B
L. <u>rufus</u> (Scop. ex Fr.)Fr.	GE?	?	B
L. <u>scrobiculatus</u> (Scop. ex Fr.)Fr.	GE?	?	B
L. <u>torminosus</u> (Schaeff. ex Fr.)Gray	GE	?	B
L. <u>trivialis</u> (Fr. ex Fr.)Fr.	GE?	?	B
L. <u>turpis</u> (Weinm.)Fr.	GE?	?	B
L. <u>uvidus</u> (Fr. ex Fr.)Fr.	GE?	?	B

Species Symptoms Toxin Distribution

<u>Lactarius</u>	<u>vellerus</u> (Fr.)Fr.	GE?	?	B
L.	<u>zonarius</u> (Bull. ex St Amans)Fr.	GE?	?	B
<u>Lepiota</u>	<u>aspera</u> (Pers. in Hof. ex Fr.)Qué1.	AP	CY	
L.	<u>brunneoincarnata</u> Chod. & Martin	AP	CY	E
L.	<u>brunneolilacea</u> Bon & Boiffart	AP	CY	
L.	<u>bucknallii</u> (Berk. & Br.)Sacc.	AP?	CY?	E
L.	<u>castanea</u> Qué1.	AP	CY	E
L.	<u>clypeolaria</u> (Bull. ex Fr.)Kum.	AP?	CY?	E
L.	<u>clypeolarioides</u> Rea	AP	CY	E
L.	<u>cortinarius</u> Lge	AP?	CY?	
L.	<u>cristata</u> (Fr.)Kum.	AP?	CY?	B
L.	<u>echinella</u> Qué1. & Bernard	AP?	CY?	E
L.	<u>felina</u> (Pers. ex Fr.)Karst.	AP?	CY?	E
L.	<u>friesii</u> (Lasch.)Qué1.	AP	CY	B

Species	Symptoms	Toxin	Distribution
<u>Lepiota fulvella</u> Rea	AP?	CY?	B
L. <u>fuscovinacea</u> Møll. & Lge	AP?	CY?	E
L. <u>grangei</u> (Eyre)Lge	AP?	CY?	E
L. <u>griseovirens</u> Mre	AP	CY	E
L. <u>heimii</u> Locq.	AP	CY	
L. <u>hetieri</u> Boud.	AP	CY	E
L. <u>kühneri</u> Huijism.	AP?	CY?	E
L. <u>ignivolvata</u> Bousset & Joss.	AP	CY	
L. <u>josserandii</u> Bon & Boiffard	AP	CY	
L. <u>langei</u> Locq.	AP	CY	
L. <u>leucothites</u> (Vitt.)Ort.	AP?	CY?	E
L. <u>lilacea</u> Bres.	AP?	CY?	E
L. <u>locanensis</u> Espinosa	AP	CY?	
L. <u>lutea</u> ss Guégen	AP?	CY?	B in glasshouses

Species	Symptoms	Toxin	Distribution
<u>Lepiota microsperma</u> Locq.	AP	CY	
L. <u>ochraceofulva</u> Ort.	AP	CY	E
L. <u>pseudofelina</u> Lge	AP	CY	B
L. <u>pseudohelveola</u> Kühn.	AP?	CY?	E
L. <u>pseudolilacea</u> Huijism.	AP?	CY?	E
L. <u>rhodorhiza</u> Romag. & Locq.	AP?	CY?	E
L. <u>rufipes</u> Morgan	AP?	CY?	
L. <u>scobinella</u> (Fr.) Gill.	AP?	CY?	E
L. <u>setulosa</u> Lge	AP?	CY?	E
L. <u>subincarnata</u> Lge	AP	CY	E
L. <u>tomentella</u> Lge	AP?	CY?	E
<u>Leucocoprinus badhamii</u> (Berk. & Br.) Mos.	AP?	CY?	B in glasshouses
L. <u>bresadolae</u> (Schulz.) Mos.	AP?	CY?	B "
L. <u>cepaespites</u> (Sow. ex Fr.) Pat.	AP?	CY?	B "

Species	Symptoms	Toxin	Distribution
<u>Lycoperdon</u> <u>marginatum</u> Vitt.	HA	?	
<u>L.</u> <u>mixtecorum</u> Heim	HA	?	
<u>Macrolepiota</u> <u>venenata</u> Bon	GE	?	
<u>Morchella</u> <u>angusticeps</u> Peck	GP?	GY?	(only raw)
<u>M.</u> <u>crassipes</u> Krombh.	GP?	GY?	"
<u>M.</u> <u>deliciosa</u> Fr.	GP?	GY?	"
<u>M.</u> <u>esculenta</u> St Amans	GP?	GY?	B "
<u>M.</u> <u>semilibera</u> DC ex Fr.	GP?	GY?	" "
<u>Nolanea</u> <u>hirtipes</u> (Schum. ex Fr.) Kum.	GE?	?	B
<u>N.</u> <u>mammosa</u> (L. ex Fr.) Qué1.	GE?	?	B
<u>N.</u> <u>murrayi</u> (Berk. & Curt.) Sing.	GE?	?	
<u>N.</u> <u>sericea</u> (Bull. ex Mérat) Ort.	GE	?	B
<u>N.</u> <u>staurospora</u> Bres.	GE?	?	B
+ numerous other <u>Nolanea</u> and <u>Leptonia</u> species	GE?	?	B

Species Symptoms Toxin Distribution

<u>Omphalotus illudens</u> Schw.	MU?	MC?	
<u>O. olearius</u> (DC ex Fr.) Sing.	MU?	MC?	
<u>O. subilludens</u> Murr.	MU?	MC?	
<u>Panaeolus africanus</u> Ota'h	HA	PS	
<u>P. ater</u> (Lge) Kühn. & Romag.	HA?	PS?	B
<u>P. campanulatus</u> (Bull. ex Fr.) QuéL.	HA?	PS?	B
<u>P. castaneifolius</u> (Murr.) Sm.	HA?	PS?	B
<u>P. fimicola</u> (Fr.) Gill.	HA?	PS?	B
<u>P. foeniseicii</u> (Pers. ex Fr.) Mre	HA	PS?	B
<u>P. microsporus</u> Ota'h & Cailleux	HA	PS?	B
<u>P. papilionaceus</u> (Bull. ex Fr.) QuéL.	HA	PS?	B
<u>P. retirugis</u> (Fr.) Gill.	HA?	PS?	B
<u>P. sphinctrinus</u> (Fr.) QuéL.	HA?	PS?	B
<u>P. subbalteatus</u> (Berk. & Br.) Sacc.	HA	PS	B

Species	Symptoms	Toxin	Distribution
<u>Paxillus involutus</u> (Batsch. ex Fr.)Fr.	hemolytic	?	B
<u>Pholiota squarrosa</u> (Müller ex Fr.)Kum.	GE?	?	B
<u>Psathyrella sepulchralis</u> Sing., Sm. & Guz.	HA?	PS?	
<u>Psilocybe aeruginomaculans</u> Höhn.	HA	PS?	
<u>P.</u> <u>aztecorum</u> Heim	HA	PS	
<u>P.</u> <u>baeocystis</u> Sing. & Sm.	HA	PS	
<u>P.</u> <u>banderillensis</u> Guz.	HA?	PS?	
<u>P.</u> <u>bolivarii</u> Guz.= <u>zapotecorum</u> Heim	HA	PS	
<u>P.</u> <u>brunneocystidia</u> Guz. & Horak	HA?	PS?	
<u>P.</u> <u>caerulescens</u> Murr.	HA	PS	
<u>P.</u> <u>caerulipes</u> (Peck)Sacc.	HA.	PS	
<u>P.</u> <u>callosa</u> (Fr. ex Fr.)Quél.	HA?	PS?	
<u>P.</u> <u>candidipes</u> Sing. & Sm.= <u>zapotecorum</u> Heim	HA	PS	
<u>P.</u> <u>collybioides</u> Sing. & Sm.	HA	PS	

Species	Symptoms	Toxin	Distribution
<u>Psilocybe coprinifacies</u> (Roll.) Pouz.	HA	PS?	
P. <u>cordispora</u> Heim	HA?	PS?	
P. <u>cubensis</u> (Earle) Sing.	HA	PS	
P. <u>cyanescens</u> Wakef.	HA	PS	B
P. <u>eucalypta</u> Guz. & Wat.	HA	PS	
P. <u>faqicola</u> Heim & Cailleux	HA	PS	
P. <u>fimetaria</u> (Ort.) Wat.	HA?	PS?	B
P. <u>galindii</u> Guz.	HA?	PS?	
P. <u>goniospora</u> (Berk. & Br.) Sing.	HA?	PS?	
P. <u>heimii</u> Guz.	HA?	PS?	
P. <u>herrerae</u> Guz.	HA?	PS?	
P. <u>hoogshagenii</u> Heim	HA	PS	
P. <u>inconspicua</u> Guz. & Horak	HA?	PS?	
P. <u>kumaenorum</u> Heim	HA	PS	

Symptoms Toxin Distribution

Species

<u>Psilocybe</u> <u>liniformans</u> Guz. & Bas	HA?	PS?	
<u>P.</u> <u>mammillata</u> Murr.	HA?	PS?	
<u>P.</u> <u>mexicana</u> Heim	HA	PS	
<u>P.</u> <u>montana</u> (Fr.) Quél.	HA?	PS?	B
<u>P.</u> <u>naematoliformis</u> Guz.	HA?	PS?	
<u>P.</u> <u>neocaledonica</u> Guz. & Horak	HA?	PS?	
<u>P.</u> <u>nova-zelandiae</u> Guz. & Horak	HA?	PS?	
<u>P.</u> <u>nothofagensis</u> Guz. & Horak	HA?	PS?	
<u>P.</u> <u>papua</u> Guz. & Horak	HA?	PS?	
<u>P.</u> <u>pellucida</u> (Sm.) Sing. & Sm.	HA	PS	
<u>P.</u> <u>plutonia</u> (Berk. & Cke) Sacc.	HA?	PS?	
<u>P.</u> <u>quebecensis</u> Oda' h & Heim	HA	PS	
<u>P.</u> <u>rzedowskii</u> Guz.	HA?	PS?	
<u>P.</u> <u>semilanceata</u> (Fr.) Kum.	HA	PS	B

Species Symptoms Toxin Distribution

<u>Psilocybe serbica</u> Mos. & Horak	HA	PS	
<u>P. silvatica</u> (Peck) Sing. & Sm.	HA	PS	
<u>P. singeri</u> Guz.	HA?	PS?	
<u>P. squamosa</u> (Pers. ex Fr.) Qué1.	HA?	PS?	B
<u>P. strictipes</u> Sing. & Sm.	HA	PS	
<u>P. stuntzii</u> Guz. & Ott	HA	PS	
<u>P. subaeruginascens</u> Höhn.	HA	PS	
<u>P. subaeruginosa</u> Cle1.	HA	PS	
<u>P. uxpapensis</u> Guz.	HA?	PS?	
<u>P. venenata</u> (Imai) Imazeki & Hongo	HA	PS?	
<u>P. veraecrusis</u> Guz. & Ortiz	HA?	PS?	
<u>P. wassonii</u> Heim	HA	PS	
<u>P. weldenii</u> Guz.	HA?	PS?	
<u>P. yungensis</u> Sing. & Sm.	HA	PS	
<u>P. zapotecorum</u> Heim	HA	PS	

Species Symptoms Toxin Distribution

<u>Ramaria aurea</u> (Fr.) QuéL.	GE	?	E
<u>R. flava</u> (Schaeff. ex Fr.) QuéL.	GE	?	
<u>R. formosa</u> (Fr.) QuéL.	GE	?	
<u>R. gelatinosa</u> (Coker) Corner	GE	?	
<u>R. lutea</u> (Vitt.) Schild	GE	?	
<u>R. mairei</u> Donk.	GE	?	E
<u>R. pallida</u> Schaeff. ex Fr.	GE	?	
<u>Russula agglutinata</u> Heim	HA?	?	
<u>R. emetica</u> Fr.	GE	?	B
<u>R. fallax</u> Kaufm.	GE?	?	B
<u>R. foetens</u> Fr.	GE?	?	B
<u>R. fragilis</u> (Pers. ex Fr.) Fr.	GE?	?	B
<u>R. kirinea</u> Heim	HA	?	
<u>R. maenadum</u> Heim	HA	?	

Species	Symptoms	Toxin	Distribution
<u>Russula</u> <u>mairei</u> Sing.	GE?	?	B
<u>R.</u> <u>nondorbingi</u> Sing.	GE	?	
<u>R.</u> <u>pseudomaenadum</u> Heim	HA	?	
<u>R.</u> <u>sardonis</u> Fr.	GE?	?	B
<u>R.</u> <u>urens</u> Romell apud Mre ex Sing.	GE?	?	
<u>Sarcosphaera</u> <u>crassa</u> (Santi ex Stendel) Pouz.	GE	?	
<u>S.</u> <u>eximia</u> (Dur. & Lév.) Mre	GE	?	
<u>Scleroderma</u> <u>cepa</u>	GE	?	E
<u>S.</u> <u>citrinum</u> Pers.	GE	?	B
<u>Stropharia</u> <u>aeruginosa</u> (Curt. ex Fr.) QuéL.	HA?	PS?	B
<u>S.</u> <u>coronilla</u> (Bull. ex Fr.) QuéL.	HA?	PS?	B
<u>S.</u> <u>cyanea</u> Tuomikoski	HA?	PS?	B
<u>S.</u> <u>ferris</u> Bres.	HA?	PS?	B
<u>S.</u> <u>hornemannii</u> (Fr. ex Fr.) Lund. & Nannf.	?	?	B

Species	Symptoms	Toxin	Distribution
<u>Suillus granulatus</u> (Fr.) Kuntze	GE?	?	B
<u>Tricholoma acerbum</u> (Bull. ex Fr.) Qué1.	GE	?	E
I. <u>albobrunneum</u> (Pers. ex Fr.) Kum.	GE	?	E
I. <u>album</u> (Schaeff. ex Fr.) Kum.	GE	?	B
I. <u>atrosquamosum</u> (Chevallier) Sacc.	GE	?	B
I. <u>bressadolanium</u> Clg	GE	?	
I. <u>helviodor</u> Pilát & Svrček	GE	?	B
I. <u>irinum</u> (Fr.) Bigel. = <u>Lepista</u>	GE	?	B
I. <u>josserandii</u> Bon	GE	?	
I. <u>luridum</u> (Schaeff. ex Fr.) Qué1.	GE	?	
I. <u>muscarium</u> Kawamura	GE	?	
I. <u>pardinum</u> Qué1.	GE	?	
I. <u>pessundatum</u> (Fr.) Qué1.	GE	?	B
I. <u>saponaceum</u> (Fr.) Kum.	GE?	?	B

Species	Symptoms	Toxin	Distribution
<u>Tricholoma</u> <u>sculpturatum</u> (Fr.) Qué1.	GE	?	B
I. <u>seijunctum</u> (Sow. ex Fr.) Qué1.	GE	?	B
I. <u>sulphurescens</u> Bres.	GE	?	E
I. <u>sulphureum</u> (Bull. ex Fr.) Kum.	GE	?	B
I. <u>ustale</u> (Fr. ex Fr.) Kum.	GE	?	B
I. <u>venenata</u> Atk.	GE	?	
I. <u>virgatum</u> (Fr. ex Fr.) Kum.	GE	?	B
<u>Tricholomopsis</u> <u>platyphylla</u> (Pers. ex Fr.) Sing.	GE?	?	B
I. <u>rutilans</u> (Schaeff. ex Fr.) Sing.	GE?	?	B
<u>Tylopilus</u> <u>felleus</u> (Bull. ex Fr.) Karst.	GE	?	B
<u>Verpa</u> <u>bohemica</u> (Krombh.) Schroet.	GE	?	
<u>Volvariella</u> <u>media</u> (Schum. ex Fr.) Sing.	GE?	?	E
V. <u>parvula</u> (Weinm.) Ort.	GE?	?	B

4.2. Survey in Scotland.

In the world list (Table 3), a column is used to indicate whether a species is found in England (E), Scotland (S) only or throughout Britain (B).

A first survey of the mushrooms found in Scotland was published in 1879 by the Rev. Stevenson. Over the years, these records have been extended, and specimens have been collected and deposited at the Royal Botanic Garden in Edinburgh as a reference or voucher collection. This was used, with the most appreciated assistance of Dr. Roy Watling, to draw up the list of Scottish records used in the above world list.

During the past four years, some areas of Scotland have been surveyed with a view to building a collection of poisonous and hallucinogenic mushrooms and related species at the Forensic Science Unit, so that further toxicological investigations could be initiated.

4.2.1. Geographical Data.

Two areas have been thoroughly surveyed, one centres in and around Glasgow and the other in North Perthshire principally around Pitlochry and Blairgowrie.

The detailed coordinate data of the collection areas are listed in Table 4, using standard 1:50'000 Ordnance Survey Maps of Britain.

Few specimens have been collected isolatedly in other areas or by other people than the present author. Collectors' name or topographical data is specified on the label of each specimen or on the collection card which accompanies each sample. Figure 31 illustrates an example of a collection card, with all its information completed. The ecological data are filled at the time of collection and additional data are subsequently recorded if the identification is doubtful, or if the sample's features vary from standard features as described in the literature for the given species.

The collection held at Strathclyde University includes, at present, 354 specimens of which 223 are of suspect or poisonous species. They are listed in Table 5 with their reference collection number and the locality they originated from. It is worth noting that some samples included here are of unknown toxicity (they have never been reported to be poisonous, but they are included because they belong to genera which have numerous poisonous species e.g. Leptonia incana, various Hypholomas, etc).

TABLE 4 ; Topographic Data of Areas Surveyed to Build up a Collection of Scottish

Poisonous and Hallucinogenic Mushrooms to be Kept at Strathclyde University.

Locality	Coordinates	Abbreviation
1) Glasgow Area	Ordnance Survey (OS) Map Sheet 64 (1976) 1: 50'000	
Strathclyde University	65600/59750 within a 50 m circle	SU
Botanic Garden (walkway)	67400/56800 to 68200/56750	BG
Gartnavel Hospital	68100/55100 to 67900/54800	GH
Pollock Park	61700/55000 to 62300/55200	PP
Mugdock Woods (along path)	75500/55400 to 77000/53800	MW
Mugdock Reservoir	76400/55400 to 75750/56800	MR
Low Craigton	75300/54250 to 76000/52900	LC
Golf Course	74600/56400 to 74000/56900	GC
Milngavie West	73700/53300 to 74500/53400	MT
	OS Sheet 71 (1976)	
Corehouse	41400/87700 to 42200/87900 and 40600/88250	CO
Hamilton High Park	54000/73700 to 53500/73400	HH

Table 4 (continued):

Locality	Coordinates	Abbreviation
2) Pitlochry-Blairgowrie Area OS Sheet 53 (1976) 1:50'000		
Dunkeld Hermitage (nature trail) 42300/01300 to 41700/00050		DH
Loch of the Lowes 44600/05400 to 44000/04500 and 44600/04750		LL
Loch Bog 42500/19000 to 41700/18500		LB
Blackcraig Forest (path) 52300/11000 to 52000/10600		BF
Standing Stones 56900/09200 to 57750/09250		SS
	OS Sheet 52 (1976) 1:50'000	
Falls of Balnaguard 51800/94250 to 50900/93900		FB
	OS Sheet 43 (1976) 1:50'000	
Killiecrankie (West of Road) 62800/91200 to 61750/91200		KC
Dalreoch 62500/06200 to 61300/07000		DA
Kindrogan Woods 62600/05900 to 62200/04700 and 63400/04350		KW

Table 4 (continued):

Locality	Coordinates	Abbreviation
Straloch	63500/03550 to 64100/03300 and 63600/03000	SL
Lin of Dee	89600/08000 to 89800/07900	LD
Glen Shee (Chair Lift)	77300/13400 within a 100 m circle	GS
Davan	63500/04150 to 63250/03900	DV

TABLE 5 : List of Suspect or Poisonous Species Collected and Kept in the Forensic Science Unit, Strathclyde University. (Abbreviations refer to Table 5)

Species	Collection No.	Locality
<u>Amanita citrina</u>	166	Comrie
<u>A. excelsa</u>	56;180;219	MT;LC;GC
<u>A. fulva</u>	8	MW
<u>A. muscaria</u>	172	Road B846
<u>A. pantherina</u>	121;244	SL;SL
<u>A. porphyria</u>	182	MW
<u>A. rubescens</u>	9;26;54	MW;PP;MT
<u>A. umbrinolutea</u>	117	SL
<u>Anellaria semiovata</u>	18;122	Loch Thom;SL
	262	SS
<u>Boletus calopus</u>	176	LC
<u>Clitocybe clavipes</u>	250	BF
<u>Clitocybe dealbata</u>	191	CO
<u>Conocybe lactea</u>	25	SU
<u>C. tenera</u>	216	Road A811
<u>Cortinarius anomalus</u>	253	LL
<u>C. aureifolius</u>	305	KW
<u>C. bicolor</u>	203	Road A8003
<u>C. cinnamomeobadius</u>	261;294	DV; LL
<u>C. cinnamomeolutescens</u>	313;329	DH;KW

TABLE 5 (continued):

Species	Collection No.	Locality
<u>Cortinarius</u> <u>cinnamomeus</u>	223; 229; 248	GC; GC; BF
	304; 328	KW; DH
<u>C.</u> <u>cinnamomeus</u>		
var. <u>conformis</u>	299	DV
<u>C.</u> <u>cinnamomeus</u>		
complex	307; 308; 309	KW; KW; KW
	310; 311; 312	KW; KW; KW
	314; 315; 316	KW; KW; KW
	317; 318; 319	KW; KW; KW
	320; 321; 322	KW; KW; KW
	323; 324; 325	KW; KW; KW
	326; 330	KW; DH
<u>C.</u> <u>croceifolius</u>	306	KW
<u>C.</u> <u>croceoconus</u>	292	BF
<u>C.</u> <u>cyanites</u>	247	BF
<u>C.</u> <u>elator</u>	220	GF
<u>C.</u> <u>flexipes</u>	249	DH
<u>C.</u> <u>hemitrichus</u>	288	Barry Links
<u>C.</u> <u>lepidopus</u>	86	DA
<u>C.</u> <u>mucosus</u>	255	LD
<u>C.</u> <u>paleaceus</u>	221	GC
<u>C.</u> <u>pallidipes</u>	303	Glenesk
<u>C.</u> <u>palustris</u>	285	Barry Links

TABLE 5 (continued) :

Species	Collection No.	Locality
<u>Cortinarius pseudosalor</u>	220	GC
<u>C. purpureobadius</u>	258	FB
<u>C. sanguineus</u>	224; 233	GC; GC
<u>C. semisanguineus</u>	264; 331; 334; 337	Merrone Woods; DH; DH; LD
<u>C. speciosissimus</u>	155; 351; 354; 353	LD; I. Alexander Rossdhu; ibid.
<u>C. sphagneti</u>	260; 280	LL; LB
<u>C. tabularis</u>	88; 183	DA; MW
<u>Entoloma clypeatum</u>	269	BG
<u>E. majaloides</u>	282	LB
<u>E. sericatum</u>	133; 301	FB; SS
<u>Gymnopilus junionus</u>	204	Kilbernie
<u>G. penetrans</u>	125; 188	SL; CO
<u>Hebeloma sinapizans</u>	267	Merrone Woods
<u>Hygrophoropsis aurantiaca</u>	58	MT
<u>Hygrocybe conicus</u>	92	DA
<u>Hypholoma capnoides</u>	144; 254	BF; DH
<u>H. elongatum</u>	283	LB
<u>H. fasciculare</u>	4; 17; 32; 106 177; 189; 197 225; 246	MW; ?; PP; DA LC; CO; Road A8003; MW; KW
<u>H. marginatum</u>	143; 242; 242 341	BF; BF; DH HH

TABLE 5 (continued):

Species	Collection No.	Locality
<u>Hypholoma radicosum</u>	291	BF
<u>H. sublateritium</u>	186;234	CO;Ross Priory
<u>Inocybe agardhii</u>	286	Barry Links
<u>I. asterospora</u>	174	MW
<u>I. brunnea</u>	298	BF
<u>I. calamistrata</u>	199	Road A8003
<u>I. dulcamara</u>	112;287	SL;Barry L.
<u>I. dunensis</u>	289	Barry Links
<u>I. fastigiata</u>	124;181;201;218 227;241;272	SL;MW;BG;A814 KW;KW;GC
<u>I. geophylla</u>	202;232;296 332	BG;GC;BF DH
var.alba	295	BF
var.lilacina	333	DH
var.lateritia	297	BF
var.violacea	340	HH
<u>I. hystrix</u>	91	DA
<u>I. lacera</u>	270;273;290	GC;GC;Barry L.
<u>I. lanuginella</u>	173;209	MW;Barrhead
<u>I. lanuginosa</u>	281	LB
<u>I. longicystis</u>	31	PP
<u>I. mixtilis</u>	275	CO

TABLE 5 (continued) :

Species	Collection No.	Locality
<u>Inocybe</u> <u>napipes</u>	271;274	GC;Lairg
<u>I.</u> <u>pudica</u>	226;240	KW;KW
<u>I.</u> <u>sambucina</u>	279	SL
<u>I.</u> <u>subnudipes</u>	293	BF
<u>I.</u> <u>umbrina</u>	265;300	SS;BF
<u>I.</u> spp.	79	MT
<u>I.</u> spp.	193;194	BG;BG
<u>Lactarius</u> <u>torminosus</u>	116	SL
<u>Lepiota</u> <u>cristata</u>	276;327;339	DA;KW;HH
<u>Leptonia</u> <u>incana</u>	134	FB
<u>Nolanea</u> <u>sericea</u>	44;101	SU;DA
<u>N.</u> <u>staurospora</u>	118;147	SL;LL
<u>Panaeolus</u> <u>acuminatus</u>	98	DA
<u>P.</u> <u>foenisecii</u>	42;82;208;212	SU;DA;SU;MR
	214;217	Case 1;A811
<u>P.</u> <u>papilionaceus</u>	102;126	DA;FB
<u>P.</u> <u>retirugis</u>	213	Barrhead
<u>P.</u> <u>rickenii</u>	211	Barrhead
<u>P.</u> <u>speciosus</u>	126;259	DA;SS
<u>P.</u> <u>sphinctrinus</u>	62;87;107	MW;DA;DA
<u>P.</u> <u>subbalteatus</u>	22;352	SU;Wat.herb.
<u>P.</u> spp.	230	A811

TABLE 5 (continued) :

Species	Collection No.	Locality
<u>Psilocybe</u> <u>coprophila</u>	84;113	SL;DA
<u>P.</u> <u>inquilina</u>	146	LL
<u>P.</u> <u>montana</u>	41;145;158	SU;LL;SL
<u>P.</u> <u>semilanceata</u>	68;114;171;205 206;207;228 231;243;302 335;336;344	SU;SL;?;Kil- bernie;GC;? Ross Priory;SU SL;SS;DA;LD;SU
<u>Russula</u> <u>emetica</u>	263	LD
<u>R.</u> <u>emeticella</u>	115	SL
<u>R.</u> <u>mairei</u>	137	BF
<u>Stropharia</u> <u>aeruginosa</u>	348	France
<u>S.</u> <u>albocyanea</u>	157;257	KW;KW
<u>S.</u> <u>albonitens</u>	347	France
<u>S.</u> <u>coronilla</u>	349;350	France
<u>S.</u> <u>cyanea</u>	111;123;185;342	KW;SL;CO;HH
<u>S.</u> <u>ferrii</u>	345;346	Tyrol;France
<u>S.</u> <u>inuncta</u>	159	KW
<u>S.</u> <u>pseudocyanea</u>	110	KW
<u>S.</u> <u>semiglobata</u>	3;60;93;164	MW;MW;DA;BF
<u>Tricholoma</u> <u>argyraceum</u>	245	BF
<u>I.</u> <u>saponaceum</u>	168	Comrie
<u>I.</u> <u>sulfureum</u>	338	KC

4.2.2. Microscopical Data.

All the above samples have been analysed microscopically to help to build up the data bank used in the identification programme. The experimental data have been compared with literature data, where available and any variations or doubtful identifications have been submitted for confirmation to Dr. Watling of the Royal Botanic Garden in Edinburgh (e.g. Inocybe umbrina, collection card No. 265; Figure 31).

Exception has to be made for the group of collections made under the collective name of "Cortinarius cinnamomeus complex". These specimens were all collected within two days and it was clear, at the time, that possibly as many as eleven different species were present. They could not all be fitted into the presently described species belonging to this complex and clearly, there is an interesting taxonomic problem as well as chemical research to be undertaken since Greer (1980) showed the possible presence of orellanine in Cortinarius cinnamomeus.

Slide made

COLLECTION CARD No. 265

LOCALITY *Standing Stone* DATE *21-9-79*
Kundhogan

HABITAT *nr. pine but some dung present too (rabbit/hare?)*

CAP ϕ *50mm.* *late brown marked umbo purplish late*
imutely fibrillose

GILLS *linear to ventricose, dentate pale brown to rusty*
margin rusty, adnate emarginate ? (or line?)

STEM *50x10-13* *no bulb but slightly clavate*
snuff brown to cigar brown up to the apex paler at base


FLESH *white no special smell*


CHEMICAL TEST

REF. No.

Recto

MICROSCOPICAL CHARACTERS

SPORES *nodulose 7.0-8.5 x 5-5.5 nodules not very*
marked 

BASIDIA *4 spores 32-40 x 5-8* 


CAP FLESH

GILL TISSUE *Req.*

CAP SURFACE *man*

CYSTIDIA *marginal cystidia thin walled never crested*
some 50-75µ x 5-10
some shorter bulbous

IDENTITY *facial crested, 60-80 x 12.5*
Inocybe umbrina

Inocybe close to boltonii macroscopically 
but close to lamugrella microscopically (see. large 11.5 µ!)
although quite different

Verso

Figure 31 : Example of a Collection Card and its Data.

4.2.3. Chemical Data.

4.2.3.1. Hallucinogenic Species.

At the early stages of this project, because of the bearing of the results on the drug abuse problem and possible legislative controls on hallucinogenic mushrooms, a group of suspect fungi belonging to the Strophariaceae were to be investigated. This was further expanded to include species of the genus Panaeolus (Coprinaceae) claimed by some authors to contain psilocybin and analogues; claims put in doubts by many others!

French samples of Stropharia were generously donated by Dr. A. Gérault and a number of Australian species of Psilocybe were given for analysis by Dr. Roy Watling.

Screening for the presence of the known hallucinogens psilocybin, psilocin, baeocystin and nor-baeocystin was carried out using standard chemical techniques detailed in the appendix (6.2.4.) with some interesting and some puzzling results, which deserve further investigation, now that a fairly extensive collection is available.

Standard psilocin, psilocybin, N-methyl-tryptamine, N,N-dimethyl-tryptamine and bufotenine were provided by the Home Office Central Research Establishment in Aldermaston. Psilocybe semilanceata was used as the standard for baeocystin, after its presence had been demonstrated in that mushroom by Repke and Leslie (1977).

Voucher specimens have been deposited at the Royal Botanic Garden in Edinburgh, and results have been published (Margot & Watling, 1981). These results are summarised in Table 6.

4.2.3.2. Discussion.

Psilocybe eucalypta, only recently described by Guzmán and Watling (1978), proves to be a new hallucinogenic mushroom growing in Australia. The other two Australian species, suspected of being hallucinogenic because of their blueing properties, Psilocybe australiana and P. subaeruginosa proved negative for known mushroom hallucinogens, and so did the yet undescribed Psilocybe species from New Zealand. These results have to be considered critically since only one analysis was allowed due to the limited amounts of sample available (a fragment of a single carpophore ranging from 10 to 30 mg) and variation between collections are known to

TABLE 6 : Results of the Analysis of some Strophariaceae and Coprinaceae Species by TLC*

with Regard to the Presence of the Hallucinogens Psilocybin (PS), Psilocin (P)

and Baecocystin (B). Unknown (U) Indole Compounds Reacting Towards Acidic

p-Dimethylaminobenzaldehyde (p-DAB) are Indicated.

+++ : over 0.30 % (w/w dry weight) ++ : over 0.10 % + : traces

Species	Collection No.	Date	PS	P	B	U	Notes
<u>Psilocybe australiana</u>	Wat.10962	1974	-	-	-	-	Wat.: Watling Herbarium
<u>P. coprophila</u>	84;113	1978	-	-	-	++	in Edinburgh,
<u>P. cubensis</u>	Wat.10816	1974	+	-	-	-	Royal Botanic
	Wat.10823	1974	+	-	-	-	Garden.
	Wat.10778	1974	+	-	-	-	PM. : P. Margot,
<u>P. cyanescens</u>	Wat.11361	1978	+++	(+)	+	-	collection
<u>P. eucalypta</u>	Wat.10631	1974	++	-	-	++	before 1978
<u>P. inquilina</u>	PM11177;146	1977/8	-	-	-	++	
<u>P. montana</u>	41;145;158	1978	-	-	-	+++	

*TLC : Thin Layer Chromatography

Table 6 (continued) :

Species	Collection No.	Date	PS	P	B	U	Notes
<u>Psilocybe semilanceata</u>	Wat.13127;68	1977/8	+++	+	++	++	
	114;205;228	1978/9					
<u>P.</u> <u>subaeruginosa</u>	Clel.13251	?	-	-	-	-	Clel.: Cleland's
<u>P.</u> spp.	Taylor 477	1969	-	-	-	-	Collection
<u>Stropharia aeruginosa</u>	348	1978	-	-	-	++	<u>legit</u> Gérard
<u>S.</u> <u>albocyanea</u>	157;257	1978/9	-	-	-	++	
<u>S.</u> <u>albonitens</u>	347	1974	-	-	-	++	<u>legit</u> Gérard
<u>S.</u> <u>coronilla</u>	349;350	1978	-	-	-	++	"
<u>S.</u> <u>cyanea</u>	111;123;185	1978	-	-	-	++	
	PM311076	1976	-	-	-	++	
<u>S.</u> <u>ferrii</u>	345;346	1970/1	-	-	-	+	"
<u>S.</u> <u>inuncta</u>	159	1978	-	-	-	++	

Table 6 (continued) :

Species	Collection No.	Date	PS	P	B	U	Notes
<u>Stropharia pseudocyanea</u>	PM91077;110	1977/8	-	-	-	++	
	PM231077		-	-	-	++	
	PM251077		-	-	-	++	
<u>S. semiglobata</u>	PM011177;3	1977/8	-	-	-	++	
	60;93;164	1978	-	-	-	++	
<u>Hypoholoma fasciculare</u>	4;17;32	1978	-	-	-	-	
<u>Panaeolus foenisecii</u>	42;82;208;212	1978/9	-	-	-	+++	at least 2 compounds
<u>P. retirugis</u>	213	1979	-	-	-	+++	at least 3 compounds
<u>P. rickenii</u>	211	1979	-	-	-	++	" " 2 compounds
<u>P. subbalteatus</u>	352	1976	++	-	-	+	
	22	1978	-	-	-	++	

occur. Recently, Perkal et al. (1980), with considerably larger samples, reported the presence of psilocybin in Psilocybe subaeruginosa using high performance liquid chromatography (HPLC). They also showed that the concentration of the drug varied considerably in different specimens. It could not be detected in one sample, using an ultra-violet detector, and ranged up to 0.21 % in others. Using a fluorescence detector, it was possible to measure trace amounts of psilocybin in the former. A factor which could influence these results is the time. The sample analysed at Strathclyde University came from Cleland's collections (spanning from the beginning of the century to the 1950's) i.e. it was at least twenty years old at the time of analysis. Similarly, the Psilocybe species originating from New Zealand was collected in 1969, ten years before analysis. Ageing experiments carried out in the Forensic Science Unit showed that the concentration of a solution of psilocybin in methanol decreased by about 25 % in 8 months, whereas psilocin almost totally disappeared during the same time period. The above discrepancies could therefore be explained by the time lag between collection and analysis.

None of the Stropharia species analysed contained any known mushroom hallucinogens, even though abundant samples were repeatedly analysed. Stropharia cyanea

had previously been found to contain psilocybin (Margot, 1977), but this could not be confirmed. This poses the problem of chemical strains (or species?) within genera, or of chemical variations during the life cycle of a mushroom.

Only extensive studies on single species could bring an answer to this problem and this is outwith the scope of the present thesis. In view of the above results, it is concluded that the Stropharia species analysed are devoid of the hallucinogens psilocybin, psilocin and baeocystin.

Hypholoma fasciculare, confirming previous results (Margot, 1977) does not contain any indole alkylamine or in any detectable levels with standard chemical techniques.

Panaeolus species have been claimed, for a long time and by many, to be hallucinogenic. This has been confirmed by some analysts and disclaimed by others. Panaeolus foenisecii is listed as a hallucinogenic species containing psilocybin in all underground guides. All the results obtained demonstrate the contrary view.

Psilocybin was found in only one sample, Panaeolus subbalteatus (collection No. 352), provided by Dr. Watling

following a case of poisoning which occurred in Edinburgh (Watling, 1977). Another sample of the same species (collection No.22) did not contain psilocybin. This would confirm Ola'h's observations (1969) that chemical strains exist within the genus Panaeolus. He divided Panaeolus species into "psilocybian", "latent psilocybian" and "non psilocybian" groups. It is worth noting that P. foenisecii is reported by Ola'h to be latent psilocybian, whereas P. subbalteatus is psilocybian, thus contradicting the present results.

On the other hand, all Panaeolus species, and especially P. retirugis, analysed were shown to contain compounds reacting similarly to psilocybin and analogues with 5 % acidic p-dimethylaminobenzaldehyde (p-DAB), but having different chromatographic behaviour to the standards used. One of the compounds had an R_f value close to that of psilocin, in all the chromatographic systems investigated. This compound was possibly the same as that found in all the Stropharia.

The widespread publicity about the hallucinogenic properties of Panaeolus species may be well founded, but the effect may be due to yet uncharacterised hallucinogens related to the known psilocybin and its analogues.

4.2.3.3. Cortinarius Species.

After a serious case of poisoning involving Cortinarius speciosissimus in Scotland (Watling, 1980; Short et al., 1980), chemical analysis of C. cinnamomeus (collections No. 223, 229 and 248), C. sanguineus (233), C. semisanguineus (264), as well as samples of C. speciosissimus from the scene of the poisoning were undertaken.

Compounds with similar chemical behaviour to that reported to orellanine (Antkowiak and Gessner, 1979) were found in all the samples, mainly in C. speciosissimus and C. cinnamomeus, and, to a lesser degree, in the other species. This confirms the suspicion that C. cinnamomeus may join the group of deadly poisonous Cortinariii. This contradicts most popular guides which describe this mushroom as "harmless", "best avoided" or "edibility unknown"!!

A clear warning that this species may indeed be deadly will be issued in the Bulletin of the British Mycological Society. Further studies are required because these and related species are widespread and common throughout Scotland.

4.3 Data Bank Collection.

From the world list of reported poisonous and hallucinogenic mushrooms, 223 are selected and included in the data bank of the present programme. Species known to be poisonous are included first, followed by British species suspected to be poisonous as well as foreign species for which a complete set of data was available. A computer read-out of the data retrieval and coded diagnostic data is listed. (Table 7)

4.3.1. Recapitulative Table of the Characters and States
Used for Data Collection and Storage.

1. Symptoms :
- 1) gastroenteritis after 8 hours, no fever, cytolytic hepatitis.
 - 2) minimum 15 hours after ingestion: gastroenteritis, intense thirst, polyuria, then renal failure.
 - 3) after alcohol consumption: flushing face and neck, swelling, tingling of hands; metallic taste, palpitations, hypotension, later nausea and sweating.
 - 4) perspiration, salivation, lacrymation, blurred vision, abdominal cramps, diarrhoea, constriction of pupils, fall in blood pressure, slow pulse.
 - 5) dizziness, drunkenness, incoordination, staggering, muscular cramps, spasms, hyperkinetic activity, deep sleep and visions.
 - 6) mood changing, unmotivated laughter, hilarity; muscle weakness, drowsiness, hallucinations.
 - 7) gastroenteritis; vomiting
 - 8) gastroenteritis after 5 hours; fever; cytolytic and haemolytic action; headaches.

2. Onset of symptoms : 1) within 2 hours

2) 3 to 8 hours

3) 8 to 12 hours

4) typically over 15 hours

3. Geographical Distribution : 1) North temperate

2) South temperate

3) subtropical

4) tropical

5) English

6) Scottish

4. Habitat : 1) ground in coniferous woods

2) ground in hardwoods

3) ground in grass

4) ground in dung

5) on stumps

6) others

5. Time of growth : 1) spring

2) summer

3) autumn

4) winter

6. Spore colour under the microscope : 1) clear - amyloid

- inamyloid

- dextrinoid

- uncertain

6. continued
- 2) pale brown
 - 3) pale pink
 - 4) dark - resistant
 - dissolves
 - uncertain

7. Spore shapes :
- 1) globose
 - 2) ellipsoid
 - 3) angular
 - 4) various

8. Spore surface :
- 1) smooth
 - 2) rough
 - 3) ridged

9. Spore wall thickness :
- 1) thin
 - 2) thick

10. Germ pore :
- 1) present
 - 2) absent

11. Spore length :
- 1) less than 6 μm
 - 2) 6 - 8 μm
 - 3) 8 - 10 μm
 - 4) 10 - 12 μm
 - 5) 12 - 17 μm
 - 6) more than 17 μm

12. Spore breadth : 1) - less than $4\ \mu\text{m}$
- 4 - 6 μm
- 6 - 8 μm
- 8 - 10 μm
- more than 10 μm

13. Colour of the spore print : 1) white
2) pink
3) brown
4) black

14. Toxins : 1) cyclopeptides
2) orellanine
3) muscarine
4) hallucinogens
5) muscimol/ibotenic acid
6) coprine
7) others
8) gyromitrin

15. Duration of symptoms : 1) weeks if not death
2) 2 to 4 hours
3) up to 6 hours
4) 6 to 24 hours
5) few days

16. Pileus trama : 1) homoiomerous
2) heteromerous
17. Gill trama : 1) regular
2) inverse
3) bilateral
4) irregular
5) absent
18. Cuticle structure : 1) cellular
2) filamentous
3) absent
19. Clamp connections : 1) present
2) absent
20. Basidium or ascus: number of spores : 1) 2 spores
2) 4 spores
3) others
21. Basidia/asci sizes : 1) small (up to 35 μ m)
2) medium (35 to 55 μ m)
3) large (over 55 μ m)
22. Marginal cystidia : 1) present : - small : - cylindrical
- medium - lageniform
- large - ampulliform
- utriform
- metuloid
- uncertain
2) absent

23. Facial cystidia : same states as for character 22

24. Chrysocystidia : 1) present

2) absent

7	PSILOCYBE SUBAERUGINASCENS MOHN PSILOCYBIN/INDOL HALL PROPOSED TREATMENT
8	PREVENT ABSORPTION, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSS
9	DIAZEPAM, CHLORPROMAZINE TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 271-272
10	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINGOFF & MITC
11	(1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
12	HALLUCINOGEN, UPSETTING E. & S.E. ASIA
13	41 5 0 2 3 -1 2 1 2 3 0 1 0 1 0 2 2 3 3 2 0 0 1 0 -1 2
14	0 1 0 0 0 -1 2 1 2 0 0 -1 2 1 2 1
15	PSILOCYBE SUBAERUGINOSA CLEL PSILOCYBIN/INDOL HALL PROPOSED TREATMENT:
16	PREVENT ABSORPT, ENHANCE EXCRETI, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSI
17	DIAZEPAM, CHLORPROMAZINE TAXON CLELAND JB (1934) HANDBOOK FLORA & FAUNA SOUTH
18	AUSTRALIA, ADELAIDE CHEM. PERKAL ET AL. (1980) J. CHROMATOG. 196, 180-84 MED
19	LINGOFF & MITCHEL (1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND R. NY
20	HALLUCINOGEN, UPSETTING AUST.
21	35 5 0 1 -1 2 1 2 0 3 0 1 0 1 0 4 2 3 3 2 0 0 1 0 1 1
22	0 0 1 0 -1 2 1 1 1 1
23	SCORTINARIUS ORELLANUS (FR.) FR. ORELLANINE TREATMENT: SYMPTOMATIC
24	REF. TAXON. ORTON PD SCORTINARIUS II THE NATURALIST APRIL 1958 P120
25	CHEM. ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21, 1931-34
26	MEDIC. GERAULT & GIRRE (1977) BUL. SOC. MYCOL. FRANCE, 93, 373-405
27	DEADLY EU & S. EU
28	41 1 3 -1 2 0 4 -1 2 0 1 -1 2 1 2 1 1 1 1 -1 2 2 3 -1 2
29	2 2 1 0 0 1 1 0 1 1 0 1 0 1 1
30	SCORTINARIUS SPECIOSISSIMUS KUHN. & ROMAG. ORELLANINE TREATMENT: SYMPTOMATIC
31	TAXON. ORTON PD (1958) SCORTINARIUS II THE NATURALIST, APRIL, 119
32	CHEM. ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21, 1931-34
33	MEDIC. GERAULT & GIRRE (1977) BUL. SOC. MYCOL. FRANCE 93, 373-405
34	DEADLY EU & N. EU
35	41 1 3 -1 2 0 5 0 -1 2 1 2 1 0 1 1 1 -1 2 2 3 -1 2 3 2
36	1 0 0 0 1 0 1 1 0 -1 2 0 1 0 1 1
37	SAMANITA PANTHERINA (FR.) SECR. UK FREQ 2.4 MUSCIMOL/IBOTENIC ACID PROP.
38	TREATMENT: PREVENT ABSORPTION, INCR. EXCRETION, SYMPTOMATIC (PHYSOSTIGMINE)
39	TAXON. JENKINS DT (1977) BIBLIO. MYCOLOGICA 57, 1-126
40	CHEM. CATALFOMO & EUGSTER (1970) BULL. NARC. 22:4, 33-41
41	MEDIC. WASER PG (1967) ETHNOPHARMAC. SEARCH .. ED. EFRON US PHSP 1645, 41
42	INEBRIANT, DANGEROUS EU; N. AM
43	32 4 0 -1 3 0 4 5 1 -1 2 1 2 0 1 1 0 0 1 3 2 0 4 3 0 1
44	1 1 1 1 1 1 1
45	SAMANITA PHALLOIDES (FR.) SECR. UK FREQ 5.5 CYCLOPEPTIDES TREATMENT: WATER,
46	ELECTROLYTE & BLOOD BALANCE, HYPERTONIC GLUCOSE INF., PENICILLIN, THIOCTIC ACID,
47	EXCHANGE TRANSFUSION TAXON. AMMIRATI ET AL (1977) MYCOLOGIA 69, 1095-1108
48	CHEM. & MED. BERTELLI, FOURNIER ET AL (1977) ED. CURRENT PROBLEMS IN CLIN
49	BIOCHEM 7, H. HUBER, BERN, STUTTGART, VIENNA DEADLY EU; N. AM.; AUST
50	35 0 2 -1 3 0 4 5 1 -1 2 1 2 0 0 0 0 1 2 -1 2 2 3 0 0
51	0 0 1 1 1 1 1 1 1
52	SAMANITA VERNA (BULL EX FRIPERS EX VITT UK FREQ 2.4 CYCLOPEPTIDES TREATMENT
53	WATER, ELECTROLYTES, BLOOD BALANCE, HYPERTONIC GLUCOSE INF, PENICILLIN, THIOCTIC
54	ACID, EXCHANGE TRANSFUSIONS TAXON. MOJEP (1978) KL. KRYPTOG. FLORA IIB/2, FIS-
55	CHEM. STUT. CHEM (MED BERTELLI, FOURNIER ET AL ED. (1977) CURRENT PROBLEMS IN
56	CLIN. BIOCHEM. 7 H. HUBER, BERN, STUTT., VIENNA DEADLY EU; N. AM.
57	38 0 2 -1 3 0 4 5 1 -1 2 0 1 0 0 0 0 1 -1 2 1 2 -1 2 2
58	3 0 0 0 0 1 1 1 1 1 1 1
59	SAMANITA MUSCARIA (L EX FR) HOOKER UK FREQ 12.3 MUSCIMOL/IBOTENIC ACID TREAT--
60	MENT: PREVENT ABS., INCREASE EXCRETION, SYMPTOMATIC (PHYSOSTIGMINE)
61	TAXON. JENKINS DT (1977) BIBLIOTHECA MYCOLOGICA 57, 1-126
62	CHEM. CATALFOMO & EUGSTER (1970) BULL. NARC. 22:4, 33-41
63	MEDIC. WASER PG (1967) ETHNOPHARMAC SEARCH... ED EFRON, US PHSP 1645, 419-3
64	INEBRIANT, DANGEROUS EU; ASIA; N. AM.
65	41 4 0 -1 3 0 4 5 -1 2 1 1 -1 2 1 2 0 1 1 0 0 1 -1 2 2 3
66	-1 2 2 3 0 4 3 0 1 1 1 1 1 1 1 1
67	SAMANITA BISPORIGERA ATK. CYCLOPEPTIDES TREATMENT: WATER, ELECTROLYTE &
68	BLOOD BALANCE, HYPERTONIC GLUCOSE INF., PENICILLIN, THIOCTIC ACID, EXCHANGE
69	TRANSFUSION TAXON. HEIM R (1957) REV MYCOL 22, 208-217
70	CHEM. & MEDIC ED. BERTELLI ET AL. (1977) CURRENT PROBLEMS IN CLINICAL
71	BIOCHEM. 7 H. HUBER, BERN, STUTT., VIENNA DEADLY N. AM.
72	31 0 2 0 1 1 0 0 0 0 0 1 -1 2 1 2 -1 2 2 3 0 0 0 0 1 1
73	1 0 1 1 1 1

Table 7 : List of the Collection of Data Stored in the

Data Bank

3	GAMANITA OCREATA PECK CYCLOPEPTIDES TREATMENT:WATER,ELECTROLYTES,BLOOD																									
4	BALANCE,HYPERTONIC GLUCOSE INF.,PENICILLIN,THIOCTIL ACID,EXCHANGE TRANS-																									
5	FUSION TAXON.AMMIRATI & AL (1977) MYCOLOGIA 69,1095-1108																									
6	CHEM. MORGEN ET AL (1976) LLOYDIA 39:5,368-375 MEDIC. BERTELLI ET AL																									
7	(1977) CURRENT PROBLEMS IN CLIN. BIOCHEM. 7 HUBER,BERN,STUTT.,VIENNA																									
8	DEADLY N. AM.																									
9	31	0	2	2	1	0	0	0	1	0	0	1	-1	2	3	4	-1	2	3	4	6	0	0	0	1	1
10	1	1	1	1	1	1																				
11	SAMANITA REGALIS FR. MUSCIMOL/IBOTENIC ACID TREATMENT:PREVENT ABSORPTION																									
12	INCREASE EXCRETION,SYMPTOMATIC (PHYSOSTIGMINE) TAXON. MOSER M (1978) KLEINE																									
13	KRYPTOG.FLORA,FISCHER,STUT. CHEM. CATALFOMO & EUGSTER (1970)																									
14	BULL. NARC 22:4,33-41 MEDIC. WASER PG (1967) EFRON ET AL ED.																									
15	ETHNOPHARMAC. SEARCH... US PHSP 1645 419-439 INEBRIANT, DANGEROUS N.EU.																									
16	34	4	0	-1	2	0	5	0	2	0	1	1	0	0	1	-1	2	2	3	-1	2	2	3	0	4	3
17	0	1	1	0	1	1	1	1	1																	
18	SENTOLOMA NIPHUIDES ROMAG UK FREQ 0.4 TOX UNKNOWN PROPOSED TREATMENT SYMP-																									
19	TOMATIC TAXON KUHNER & ROMAGNESI (1953) FLORE ANALYTIQUE MASSON CIE PARIS																									
20	CHEM HEROUT V (1968) PLANTA MED. SUPL 90-98																									
21	MED TYLER V JR (1963) PROGRESS IN CHEM TOX 1,339-384																									
22	DANGEROUS TO UPSETTING EU																									
23	41	6	0	-1	3	0	4	5	-1	2	2	5	0	2	2	0	0	1	-1	3	1	2	3	-1	2	1
24	2	1	6	-1	2	3	4	0	0	1	0	1	1	1	1	1										
25	SENTOLOMA RHODOPOLIUM (FR)KUMMER UK FREQ 3.6 TOX UNKNOWN TREATMENT:SYMPTO-																									
26	MATIC TAXON KUHNER & ROMAGNESI (1953) FLORE ANALYTIQUE... MASSON CIE PARIS																									
27	CHEM HEROUT V (1968) PLANTA MED SUPL 90-98																									
28	MED TYLER V JR (1963) PROGRESS IN CHEM TOX 1,339-384																									
29	UPSETTING TO DANGEROUS EU																									
30	31	6	0	-1	3	0	4	5	1	2	2	2	0	0	1	2	2	1	6	-1	2	3	4	0	0	1
31	0	1	0	1	1	1																				
32	SENTOLOMA SINUATUM (BULL EX FR)KUMMER UK FREQ 0.4 TOX UNKNOWN PROPOSED TREAT-																									
33	MENT:SYMPTOMATIC TAXON KUHNER & ROMAGNESI (1953) FLORE ANALYTIQUE... MASSON																									
34	CIE PARIS CHEM HEROUT V (1968) PLANTA MED SUPL 90-98																									
35	MED TYLER V JR (1963) PROGRESS IN CHEM TOX 1,339-384																									
36	UPSETTING TO DANGEROUS EU;N.AM.																									
37	40	6	0	-1	3	0	4	5	1	-1	2	1	2	2	2	0	0	1	2	-1	2	2	3	1	6	-1
38	2	3	4	0	0	1	0	1	-1	2	1	2	1	1	1											
39	SENTOLOMA VERNUM LUNDELL TOX UNKNOWN TREATMENT:SYMPTOMATIC																									
40	TAXON KUHNER & ROMAGNESI (1953) FLORE ANALYTIQUE... MASSON CIE PARIS																									
41	CHEM HEROUT V (1968) PLANTA MED SUPL 90-98																									
42	MED AYER F (1974) SCHWEIZ Z PILZKUNDE 89:2,17-19																									
43	UPSETTING TO DANGEROUS EU																									
44	34	6	0	-1	3	0	4	5	-1	2	2	5	0	2	2	0	0	1	2	2	1	6	-1	2	3	4
45	0	0	1	0	1	0	1	1	1																	
46	SAGARICUS PHAELEPIDOTUS MOELL TOX UNKNOWN TREATMENT:SYMPTOMATIC																									
47	TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER STUTT. NY																									
48	CHEM. NONE																									
49	MED LINGOFF & MITCHEL (1977) TOXIC & HALL. MUSHROOM POIS..VAN NOSTRAND																									
50	REINHOLD NY 197-198 UPSETTING EU																									
51	39	6	0	-1	2	0	4	-1	2	1	2	-1	2	1	2	3	0	1	0	1	0	0	0	3	6	1
52	0	0	1	1	1	0	0	-1	2	0	1	2	1	1												
53	SAGARICUS PLACOMYCES PECK UK FREQ 3.2 TOX UNKNOWN PROPOSED TREATMENT:																									
54	SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER STUTT. NY																									
55	CHEM. NONE																									
56	MED LINGOFF & MITCHEL (1977) TOXIC & HALLU. MUSHROOM POISONING VAN																									
57	NOSTRAND REINHOLD NY 197-198 UPSETTING EU; N. AM.																									
58	40	6	0	-1	3	0	4	5	-1	2	0	1	1	3	0	1	0	1	0	-1	2	0	1	0	3	6
59	1	0	0	1	1	1	0	0	-1	2	1	2	2	1	1											
60	SAGARICUS XANTHODERMUS GEN UK FREQ 2.0 TOX UNKNOWN PROPOSED TREATMENT:																									
61	SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER STUTT. NY																									
62	CHEM. NONE																									
63	MED LINGOFF & MITCHEL (1977) TOXIC & HALLUCINOGENIC MUSHROOM POISONING																									
64	VAN NOSTRAND REINHOLD NY 197-198 UPSETTING EU																									
65	41	6	0	-1	3	0	4	5	-1	3	3	1	2	-1	2	1	2	3	0	1	0	1	0	-1	2	0
66	1	0	3	6	1	0	0	1	1	1	0	0	0	2	1	1										

Table 7 (continued)

6	AMANITA CRENULATA PECK	IBOTENIC ACID/MUSCIMOL	TREATMENT: PREVENT ABSORPTION, INCREASE EXCRETION, SYMPTOMATIC (PHYSOSTIGMINE)	TAXON. JENKINS DT (1977) BIBLIOTHECA MYCOL. 57, 1-126	CHEM CATALFOMO & EUGSTER (1970) BULL. NARC. 22:4, 33-41	MED. JUCK R.W (1965) NEW ENG. J MED. 272, 475-476	WASER PG (1967) ETHNOPHARM. SEARCH... ED EFRON ET AL. US PHSP 1645 419-39	INEBRIANT, DANGEROUS	N. AM.	31	4	0	0	-1	2	0	1	2	0	1	0	0	0	1	-1	2	1	2	3	0	4	3	0	1	1	
7										1	1	1	1	1	1																					
8	AMANITA GEMMATA (FR) BERT.	EUROPE: EDIBLE, US: SUSPECT	IBOTENIC A/MUSCIMOL	S. AM: DEADLY! TREATMENT: SYMPTOMATIC IF SYMPTOMS ONSET > 8 HOURS	WATER, ELECTROLYTES, BLOOD BALANCE, HYPERTONIC GLUC. INF. PENICILLIN, THIOCTIC ACID, EXCH TRANSF.	TAXON JENKINS DT (1977) BIBLIOT. MYCOL 57, 1-126	CHEM CHILTON & OTT (1976) LLOYDIA 39:2, 3, 150-157	MED. SEE AMANITA PHALLOIDES FOR S. AM.	EU: N. AM.	42	4	0	-1	3	0	4	5	-1	2	0	1	-1	3	0	1	2	0	1	1	0	0	1	-1	2	2	
9										3	2	0	4	3	0	1	1	0	1	-1	2	1	2	1	1	1										
10	AMANITA FROSTIANA (PECK) SACC	SUSPECT: IBOTENIC ACID/MUSCIMOL	TREATMENT: PREVENT ABSORPTION, INCREASE EXCRETION, SYMPTOMATIC (PHYSOSTIGMINE)	TAXON JENKINS DT (1977) BIBLIOTHECA MYCOL. 57, 1-126	CHEM CATALFOMO & EUGSTER (1970) BULL. NARC. 22:4, 33-41	MED. COKER WC (1917) J ELISHA MITCH SOC 33:1:2, 1-88	OR WASER PG (1967) EFRON ET AL. ED. ETHNOPHARMAC SEARCH... US PHSP 1645, 419-439	SUSPECT INEBRIANT, DANGEROUS	N. AM.	37	4	0	0	-1	2	0	1	-1	2	1	2	0	1	0	0	0	1	-1	2	1	2	-1	2	2	3	
11										0	4	3	0	1	1	0	1	1	1	1	1															
12	MACROLEPIOTA VENENATA BON	UNKNOWN	TREATMENT: SYMPTOMATIC	TAXON & MED. BON ET AL (1979) DOCUMENTS MYCOL. 9:35, 13-21	CHEM. NONE																															
13										36	6	0	0	-1	2	2	3	-1	2	1	2	0	2	1	0	1	0	3	2	0	6	1	0	0		
14										1	1	-1	2	0	1	0	1	0	1	1																
15	HYPHOLOMA FASCICULARE (HUDS EX FR) KUMMER 49.4	UNKNOWN	TREATMENT: SYMPTOMATIC	TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2	FISCHER STUT CHEM NISHIHARA ET AL (1958) DOSHIDA ENG REV 9:1, 36-38	MED HERBICH ET AL (1966) ARCHIV TOXIKOL 21, 310-320	DANGEROUS	EU: ASIA: N. AM.		41	6	2	-1	3	0	4	5	4	-1	4	0	1	2	3	3	1	1	0	1	0	1	0	3	6	4	
16										0	0	1	0	1	0	0	0	2	0	-1	2	0	1	2	0											
17	PSILOCYBE AZTECORUM HEIM	PSILOCYBIN/INDOLE	HALL. TREATMENT: PREVENT ABSORPTION, ENHANCE EXCRETION, RESTRAIN FROM SELF DESTRUCTIVE BEHAV.	QUIET ? DIAZEPAM, ? CHLORPROMAZINE	TAXON HEIM & WASSON (1958) LES CHAMPIGNONS HALL. DU MEXIQUE. MUSEUM NAT. HIST. NATU, PARIS	CHEM HOFMANN ET AL (1958) IN HEIM & WASSON MED. LINGOFF & MITCHELL (1977) TOXIC AND HALL..	VAN NOSTRAND NY	HALLUCINOGEN, UPSETTING	C. AM.	39	5	0	2	-1	2	0	2	-1	2	1	2	3	0	1	0	1	0	-1	2	3	4	2	3	3	2	
18										0	0	1	0	3	0	0	-1	2	0	1	1	1	1													
19	PSILOCYBE BAEUCYSTIS SING/SM	PSILOCYBIN/INDOLE	HALL. TREATMENT: PREVENT ABSORPTION, ENHANCE EXCRETION, RESTRAINT SELF DESTRUCTIVE BEHAVIOUR, QUIET POSS.	DIAZEPAM, CHLORPROMAZINE	TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 141	CHEM LEUNG & PAUL (1968) J PHARM SCI 57, 1667-1671	MED LINGOFF & MITCHELL (1977) TOXIC AND HALLUC. MUSHROOM POISONING	VAN NOSTRAND R NY	HALLUCINOGEN, UPSETTING	N. AM.	30	5	0	0	4	1	3	0	1	0	1	0	-1	2	3	4	2	3	3	2	0	0	1	0	1	0
20										0	0	2	1	1																						
21	PSILOCYBE CAERULESCENS MURR	PSILOCYBIN/INDOLE	HALLUC. TREATMENT: PREVENT ABSORPTION, ENHANCE EXCRETION, RESTRAINT SELF DESTRUCTIVE BEHAV. POSS.	DIAZEPAM, CHLORPROMAZINE	TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 262-303	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572	MED LINGOFF & MITCHELL (1977) TOXIC & HALLUC. MUSHROOM POISONING, VAN NOSTRAND R, NY	HALLUCINOGEN, UPSETTING	C. AM.; N. AM.	34	5	0	2	-1	3	1	2	3	-1	2	0	1	3	0	1	0	1	0	1	1	3	3	2	0	0	
22										1	0	1	0	0	0	2	1	1																		
23	PSILOCYBE CAERULIPES (PECK) SACC	PSILOCYBIN/INDOLE	HALL TREATMENT: PREVENT ABSORPTION, ENHANCE EXCRETION, RESTRAINT SELF DESTRUCT. BEHAV., QUIET, POSS.	DIAZEPAM, CHLORPROMAZINE	TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 262-30	CHEM HOFMANN ET AL (1959) HELV CHEM ACTA 42, 1557-1572	MED LINGOFF & MITCHELL (1977) TOXIC AND HALLUC. MUSHROOM POISONING	VAN NOSTRAND R, NY	HALLUCINOGEN, UPSETTING	N. AM.	33	5	0	0	4	-1	2	1	2	3	0	1	0	1	0	-1	2	1	2	1	3	3	2	0	0	1
24										0	1	0	0	0	2	1	1																			

Table 7 (continued)

7	6PSILOCYBE CANDIDIPES SING & SM. PSILOCYBIN/INDOLE HALL. TREATMENT: PREVENT
8	ABSORPTION, ENHANCE EXCRETION, RESTRAIN SELF DESTRUCT. BEHAV., QUIET, POSS.
9	DIAZEPAM, CHLORPROMAZINE TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 141
10	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINGOFF & MITC
11	(1977) TOXIC & HALLUC. MUSHROOM POISONING, VAN NOSTRAND R, NY
12	HALLUCINOGEN, UPSETTING C.AM.
13	32 5 0 2 -1 2 2 3 2 3 0 1 0 1 0 1 0 3 3 2 0 0 1 0 1 0
14	0 0 1 0 0 0 1
15	6PSILOCYBE CUBENSIS (EARLE) SING PSILOCYBIN/INDOL HALLU. TREATMENT: PREVENT
16	ABSORPTION, ENHANCE EXCRETION, RESTRAIN SELF DESTRUCT. BEHAV., QUIET, POSS.
17	DIAZEPAM, CHLORPROMAZINE TAXON SINGER (1948) SYDOWIA 2, 37 & 40-41 CHEM
18	HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINGOFF & MITCHELL
19	(1977) TOXIC AND HALLUCI. MUSHROOM POISONING, VAN NOSTRAND R., NY
20	HALLUCINOGEN, UPSETTING SUBTROPICAL; TROPICAL; N.AM; C.AM; S.AM
21	40 5 0 -1 3 0 1 2 3 -1 3 0 1 2 3 0 1 0 1 0 4 -1 2 3 4 3
22	3 2 0 0 1 0 1 0 0 0 1 0 0 2 1
23	6PSILOCYBE CYANESCENS WAKEF PSILOCYBIN/INDOLE HALL. TREATMENT: PREVENT
24	ABSORPTION, ENHANCE EXCRETION, RESTRAIN SELF DESTRUCT BEHAV., QUIET, POSS.
25	DIAZEPAM, CHLORPROMAZINE TAXON WAKEFIELD (1946) TRANS BRIT MYCOL SOC 29,
26	115-143 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LIN-
27	COFF & MITCHEL (1977) TOXIC AND HALLUC. MUSH. POISONING, VAN NOSTRAND R, NY
28	HALLUCINOGEN, UPSETTING EU
29	42 5 0 -1 3 0 4 5 2 2 3 0 1 0 1 0 -1 2 2 3 -1 2 1 2 3 3
30	2 0 0 1 0 1 0 0 0 -1 2 1 2 0 0 1 1
31	6PSILOCYBE EUCALYPTA GUZ & GAT PSILOCYBIN/INDOLE HALLU. PROPOSED TREATMENT
32	PREVENT ABSORPTION, ENHANCE EXCRETION, RESTRAIN SELF DESTRUCT BEHAV, POSSIB.
33	DIAZEPAM, CHLORPROMAZINE TAXON GUZMAN & WATLING (1978) NOTES ROY BOT GD E
34	36, 179-210 CHEM MARGOT & WATLING (1980) TRANS BRIT MYCOL SOC
35	MED LINGOFF & MITCHEL (1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND
36	HALLUCINOGEN, UPSETTING AUST.
37	41 5 0 1 -1 2 2 5 0 3 0 1 0 1 0 -1 2 2 3 2 3 3 2 0 0 1
38	0 1 -1 2 0 1 0 0 -1 2 1 2 0 0 1 1
39	6PSILOCYBE CORUISPORA HEIM PSILOCYBIN/INDOLE HALL. PROPOSED TREATMENT:
40	PREVENT ABSORP., ENHANCE EXCRET., RESTRAIN SELF DESTRUCT BEHAV, QUIET
41	POSS. DIAZEPAM, CHLORPROMAZINE TAXON HEIM & WASSON (1958) CHAMP. HALLUC.
42	MEXIQUE MUS. NAT. HIST. NATU 7 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA
43	42, 1557-1572, MED. LINGOFF & MITCHEL (1977) TOX. & HALLU. MUSH... VAN NOSTRAND
44	HALLUCINOGEN, UPSETTING C.AM.
45	36 5 0 2 -1 2 2 4 1 3 0 1 0 1 0 -1 2 1 2 -1 2 0 1 3 3 2
46	0 0 1 0 1 0 0 0 2 1 1
47	6PSILOCYBE FAGICOLA HEIM/CAIL. PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT:
48	PREVENT ABSORP., ENHANCE EXCRET., RESTRAIN SELF DESTRUCT BEHAV., QUIET, POSSI
49	DIAZEPAM, CHLORPROMAZINE TAXON HEIM & CAILLEUX (1959) REV. MYCOL 24, 437
50	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINGOFF & MITC
51	(1977) TOX AND HALLUC. MUSHROOM POISONING, VAN NOSTRAND R, NY
52	HALLUCINOGEN, UPSETTING C.AM
53	33 5 0 2 1 1 3 0 1 0 1 0 -1 2 0 1 -1 2 0 1 3 3 2 0 0 1
54	0 1 0 0 0 1 1 1
55	6PSILOCYBE FIMETARIA (ORT) WAT PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT:
56	PREVENT ABSORP., ENHANCE EXCRET., RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSI
57	DIAZEPAM, CHLORPROMAZINE TAXON ORTON (1964) NOTES ROY BOT GD EDIN 26, 59-6
58	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINGOFF &
59	MITCHEL (1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND R, NY
60	HALLUCINOGEN, UPSETTING EU
61	34 5 0 -1 3 0 4 5 3 2 3 0 1 0 1 0 -1 2 3 4 2 3 3 2 0 0
62	1 0 1 0 0 0 1 1 1
63	6PSILOCYBE HOOGHSHAGANI HEIM PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT:
64	PREVENT ABSORP., ENHANCE EXCRET., RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
65	DIAZEPAM, CHLORPROMAZINE TAXON HEIM & WASSON (1958) CHAMP. HALL. MEXIQUE
66	MUS. NAT. HIST. NATU 7, 167-169 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42,
67	1557-1572 MED LINGOFF & MITCHEL (1977) TOX. HALL. MUSH. POISONING, VAN NOS.
68	NY HALLUCINOGEN, UPSETTING C.AM.
69	35 5 0 2 -1 2 1 2 1 3 0 1 0 1 1 1 3 3 2 0 0 1 0 1 0
70	0 -1 2 0 1 1 0 0 2 1

Table 7 (continued)

4	6PSILOCYBE KUMAENORUM HEIM PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT:
5	PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB
6	DIAZEPAM, CHLORPROMAZINE TAXON HEIM (1967) MUS NAT HIST NAT 7, 186-188
7	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH
8	(1977) TOX. & HALL MUSH. POISONING, VAN NOSTRAND R, NY
9	HALLUCINOGEN, UPSETTING NW GUINEA
10	33 5 0 3 2 1 3 0 1 0 1 0 -1 2 0 1 -1 2 0 1 3 3 2 0 0 1
11	0 1 0 0 0 1 1 1
12	6PSILOCYBE MEXICANA HEIM PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT:
13	PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
14	DIAZEPAM, CHLORPROMAZINE TAXON HEIM & WASSON (1958) CHAMP. HALL. MEXIQUE,
15	MUS. NAT. HIST. NATU. 7, 129-137 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42,
16	1557-1572 MED. LINCOFF & MITCHEL (1977) TOX & HALL. MUSH. POISONING, VAN NOS.
17	HALLUCINOGEN, UPSETTING C. AM.
18	38 5 0 2 2 -1 2 1 2 3 0 1 0 1 0 -1 2 2 3 2 3 3 2 0 0 1
19	J 1 -1 2 0 1 0 0 1 0 0 1 1
20	6PSILOCYBE MIXAENSIS HEIM PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT:
21	PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT. BEHAV, QUIET, POSSIB
22	DIAZEPAM, CHLORPROMAZINE TAXON HEIM & WASSON (1958) CHAMP. HALL. MEXIQUE
23	MUS. NAT. HIST. NATU. 7, 169-171 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42,
24	1557-1572 MED LINCOFF & MITCHEL (1977) TOX HALLU. MUSH. POISONING, VAN NOST.
25	NY HALLUCINOGEN, UPSETTING C. AM.
26	36 5 0 2 -1 2 1 2 1 3 0 1 0 1 0 -1 2 0 1 -1 2 0 1 3 3 2
27	0 0 1 0 1 0 0 0 1 1 1
28	6PSILOCYBE MONTANA (FR) QUEL PSILOCYBIN/INDOLE HALL PROPOSED TREATMENT:
29	PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB
30	DIAZEPAM, CHLORPROMAZINE TAXON ORTON (1969) NOTES ROY BOT GD EDIN, 29: 1, 75-127
31	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHEL
32	(1977) TOX & HALLU. MUSHROOM POISONING, VAN NOSTRAND R, NY
33	HALLUCINOGEN, UPSETTING EU; N. AM
34	43 5 0 -1 3 1 1 -1 2 2 5 -1 2 1 2 3 0 1 0 1 0 -1 2 0 1
35	-1 2 0 1 3 3 2 0 0 1 0 1 0 0 0 1 1 1
36	6PSILOCYBE PELLILULOSA (SM) SING/SM PSILOCYBIN/INDOLE HALL. PROPOSED
37	TREATMENT: PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET
38	POSSIB. DIAZEPAM, CHLORPROMAZINE TAXON SINGER & SMITH (1958) MYCOLOGIA
39	50, 280-281 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED
40	LINGOFF & MITCHEL (1977) TOX & HALL. MUSHROOM POISONING, VAN NOSTRAND R, NY
41	HALLUCINOGEN, UPSETTING N. AM.
42	42 5 0 0 -1 2 0 5 -1 2 1 2 3 0 1 0 1 0 -1 2 2 3 -1 2 1 2
43	3 3 2 0 0 1 0 1 0 0 -1 2 0 1 1 1 1
44	6PSILOCYBE QUEBECENSIS OLA'H/HEIM PSILOCYBIN/INDOLE HALLU. PROPOSED TREAT
45	PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
46	DIAZEPAM, CHLORPROMAZINE TAXON OLA'H & HEIM (1967) C. R. ACAD SCI D 264, 1601-1603
47	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH
48	(1977) TOX & HALLU. MUSHROOM POISONING, VAN NOSTRAND R, NY
49	HALLUCINOGEN, UPSETTING N. AM.
50	38 5 0 0 -1 2 2 5 -1 2 1 2 3 0 0 0 1 0 -1 2 2 3 2 3 3 2
51	0 0 1 0 1 0 0 0 1 0 0 1 1
52	6PSILOCYBE SEMILANCEATA (FR) KUM UK FREQ 5.9 PSILOCYBIN/INDOL HALL.
53	PROPOSED TREATMENT PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT
54	BEHAV, QUIET POSSIB DIAZEPAM, CHLORPROMAZINE TAXON VON MICHAELIS (1977)
55	Z PILZK 43, 385-310 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572
56	MED LINCOFF & MITCHEL (1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOST-
57	RAND R, NY HALLUCINOGEN, UPSETTING EU; N. AM.
58	34 5 0 -1 3 0 4 5 2 -1 2 1 2 3 0 1 0 1 0 4 2 3 3 2 0 0
59	1 0 1 0 0 0 1 1 1
60	6PSILOCYBE SEMPERVIVA HEIM/CAIL PSILOCYBIN/INDOLE HALLU. PROPOSED TREATMEN
61	PREVENT ABSORP., ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB
62	DIAZEPAM, CHLORPROMAZINE TAXON HEIM (1958) REV MYCOL 23, 352
63	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHEL
64	(1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND R, NY
65	HALLUCINOGEN, UPSETTING C. AM.
66	30 5 0 2 2 1 3 0 1 0 1 0 -1 2 1 2 1 3 3 2 0 0 1 0 1 0
67	0 0 1 1 1

Table 7 (continued)

6	PSILOCYBE SERBICA MOS/HOR	PSILOCYBIN/INDOLE HALL. PROPOSED TREATMENT:
51	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB	
	DIAZEPAM, CHLORPROMAZINE TAXON MOSER & HORAK (1968) Z PILZKUNDE 34, 137-144	
	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHEL	
	(1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND R, NY	
	HALLUCINOGEN, UPSETTING EU	
	47 5 0 0 -1 2 0 1 -1 2 1 2 3 0 1 0 1 0 -1 2 2 3 -1 2 1 2	
	3 3 2 0 0 1 0 -1 2 0 1 0 0 0 2 0 -1 2 0 1 1 1	
6	PSILOCYBE SILVATICA (PK) SING/SM	PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT
6	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB	
	DIAZEPAM, CHLORPROMAZINE TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 277-278	
9	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHE	
	(1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND R, NY	
10	HALLUCINOGEN, UPSETTING EU	
	39 5 0 0 -1 2 0 1 2 3 0 1 0 1 1 -1 2 1 2 -1 2 1 2 3 3 2	
	0 0 1 0 2 0 0 -1 2 0 1 2 1 1	
6	PSILOCYBE STRICTIPES SING/SM	PSILOCYBIN/INDOL HALL PROPOSED TREATMENT:
71	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB	
	DIAZEPAM, CHLORPROMAZINE TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 288-289	
75	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH	
	(1977) TOX AND HALL. MUSHROOM POISONING, VAN NOSTRAND R, NY	
79	HALLUCINOGEN, UPSETTING N.A.M	
	36 5 0 0 0 2 3 0 1 0 1 0 -1 2 2 3 -1 2 1 2 3 3 2 0 0 1	
	0 1 0 0 -1 2 0 1 2 1 1	
22	PSILOCYBE STUNTZII GUZ/OTT	PSILOCYBIN/INDOL HALL. PROPOSED TREATMENT:
22	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB	
	DIAZEPAM, CHLORPROMAZINE TAXON GUZMAN & OTT (1976) MYCOLOGIA 68, 1261-126	
24	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH	
	(1977) TOX & HALLU. MUSHROOM POISONING, VAN NOSTRAND R, NY	
25	HALLUCINOGEN, UPSETTING N.A.M.	
	40 5 0 0 -1 2 1 2 -1 3 0 1 2 3 0 1 0 1 0 -1 2 2 3 -1 2 1	
	2 3 3 2 0 0 1 0 1 0 0 0 1 1 1	
70	PSILOCYBE YUNGENSIS SING/SM	PSILOCYBIN/INDOL HALL PROPOSED TREATMENT:
70	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB	
	DIAZEPAM, CHLORPROMAZINE TAXON SINGER & SMITH (1958) MYCOLOGIA 50, 142	
72	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHE	
	(1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY	
73	HALLUCINOGEN, UPSETTING C.A.M.	
	33 5 0 2 4 1 3 0 1 0 1 0 0 -1 2 0 1 3 3 2 0 0 1 0 1 0	
	0 0 -1 2 1 2 1 1	
78	PSILOCYBE WASSONII HEIM	PSILOCYBIN/INDOLE HALLU. PROPOSED TREATMENT:
78	PREVENT ABSORPT, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB	
	DIAZEPAM, CHLORPROMAZINE TAXON HEIM & WASSON (1958) MUS NAT HIST NATUR, 7	
40	PARIS CHEM HOFMANN ET AL (1958) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF	
	& MITCHEL (1977) TOX & HALLU MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY	
42	HALLUCINOGEN, UPSETTING C.A.M.	
	33 5 0 2 2 -1 2 1 2 3 0 1 0 1 0 -1 2 1 2 1 3 3 2 0 0 1	
	0 1 0 0 0 1 1 1	
16	PSILOCYBE ZAPOTECORUM HEIM	PSILOCYBIN/INDOLE HALLU PROPOSED TREATMENT:
16	PREVENT ABSORPT, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB	
	DIAZEPAM, CHLORPROMAZINE TAXON HEIM & WASSON (1958) MUS NAT HIST NATU, 7, 14	
48	153. PARIS CHEM HOFMANN ET AL (1958) HELV CHIM ACTA 42, 1557-1572 MED LINCO	
	& MITCHEL (1977) TOX & HALLU MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY	
50	HALLUCINOGEN, UPSETTING C.A.M	
	35 5 0 2 -1 2 2 5 1 3 0 1 0 1 0 1 -1 2 0 1 3 3 2 0 0 1	
	0 1 0 0 0 1 0 0 1 1	
54	PSILOCYBE ALBODISCA PECK UK	FREQ 0.4 MUSCARINE PROPOSED TREATMENT: PREVENT
54	ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE	
	TAXON PECK (1898) ANN REP NY ST MUS 51, 290 CHEM CATALFOMO & EUGSTER	
56	(1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC	
	MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY..	
58	UPSETTING TO DANGEROUS S.EU; N.A.M	
	50 3 0 -1 2 0 4 -1 3 0 1 2 -1 2 1 2 1 3 0 0 1 -1 2 1 2 1	
	2 2 3 0 0 1 0 1 0 0 2 -1 2 1 2 0 -1 2 -1 2 -1 2 -1 2 1	

Table 7 (continued)

62 6INOCYBE ASTEROSPORA QUEL UK FREQ 9.1 MUSCARINE PROPOSED TREATMENT:PRE-
 VENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIV
 64 TAXON HEIM (1931) LE GENRE INOCYBE LECHEVALIER, PARIS CHEM CATALFOMO &
 EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS EU;N.AM

50 3 0 -1 3 0 4 5 1 2 1 3 0 1 1 -1 2 2 3 -1 2 2 3 2 2 3
 0 0 1 0 1 1 0 -1 2 1 2 -1 2 2 4 0 -1 2 1 2 -1 2 2 4 1

6 INOCYBE BRUNNEA QUEL UK FREQ 0.4 MUSCARINE PROPOSED TREATMENT:PREVENT
 ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
 TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
 EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS EU.

52 3 0 -1 3 0 4 5 -1 3 0 1 2 -1 2 0 1 1 1 0 0 1 -1 3 2 3
 4 -1 2 1 2 2 2 3 0 0 1 0 1 0 0 2 -1 2 2 4 0 2 -1 2 2
 4 1

6 INOCYBE CINCINNATA (FR) QUEL UK FREQ 2.8 MUSCARINE PROP. TREATMENT:PRE-
 VENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
 TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
 EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS EU;N.AM;N.AF

42 3 0 -1 4 0 2 4 5 -1 2 0 1 2 1 1 0 0 1 2 1 2 2 3 0 0
 1 0 1 0 0 2 -1 2 0 4 0 2 -1 2 0 4 1

6 INOCYBE CORYDALINA QUEL UK FREQ 1.2 MUSCARINE PROPOSED TREATMENT:PREVENT
 ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
 TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
 EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS S.EU;N.AM

40 3 0 -1 2 0 4 1 2 1 1 0 0 1 -1 2 1 2 1 2 2 3 0 0 1 0
 1 0 0 1 -1 2 1 4 0 1 -1 2 1 4 1

6 INOCYBE DULCAMARA (ALB & SCHW EX PERS) KUM UK FREQ 3.2 MUSCARINE PROP.
 TREATMENT:PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS
 SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CA-
 TALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHE
 (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS EU;N.AF

33 3 0 -1 4 0 2 4 5 0 -1 3 0 1 2 1 1 0 0 1 2 1 2 2 3 0
 0 1 0 1 1 1 1 1

6 INOCYBE EUTHELES BK & BR UK FREQ 2.0 MUSCARINE PROPOSED TREATMENT:PRE-
 VENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIV
 TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
 EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHELL
 (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS EU;N.AM

38 3 0 -1 3 0 4 5 -1 2 0 2 -1 2 1 2 1 1 0 0 1 2 1 2 2 3
 0 0 1 0 1 0 0 2 1 0 2 1 1

6 INOCYBE FASTIGIATA (SCH EX FRIQUEL) UK FREQ 17.0 MUSCARINE PROPOSED TREAT
 MENT:PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS.
 SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS
 CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF &
 MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS EU;N.AF;N.AM;S.AM

47 3 0 -1 4 0 2 4 5 -1 3 2 3 5 -1 2 1 2 1 1 0 0 1 -1 2 3
 4 -1 2 1 2 2 2 3 0 0 1 0 1 -1 2 1 2 0 2 0 1 1

6 INOCYBE FLOCCULOSA (BK) SACC UK FREQ 6.3 MUSCARINE PROPOSED TREATMENT:PRE
 VENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTI
 TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
 EUGSTER (1970) HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
 UPSETTING TO DANGEROUS N.EU;N.AM

45 3 0 -1 4 0 2 4 5 1 -1 2 1 2 1 1 0 0 1 -1 2 2 3 1 2 2
 3 0 0 1 0 1 0 0 -1 2 1 2 1 1 -1 2 1 2 1 1

Table 7 (continued)

67	GINOCYBE FIBROSA (SOW)BRES MUSCARINE PROPOSED TREATMENT:PREVENT ABSORP-
	TION,ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE
68	TAXON HEIM (1931)LE GENRE INOCYBE,LECHEVALIER,PARIS CHEM CATALFOMO &
	EUGSTER (1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
	TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS EU;N.AM
	41 3 0 -1 2 0 4 0 -1 2 1 2 1 3 0 0 1 -1 2 1 2 1 2 2 3 0
	0 1 0 1 0 0 2 -1 2 4 0 2 -1 2 4 1
69	GINOCYBE FRIESII HEIM MUSCARINE PROPOSED TREATMENT:PREVENT ABSORP,ENHANCE
	EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE TAXON HEIM (1931)
70	LE GENRE INOCYBE,LECHEVALIER,PARIS CHEM CATALFOMO & EUGSTER (1970)HELV
	CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM
71	POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS EU;N.AF
72	45 3 0 -1 4 0 2 4 5 0 1 1 1 0 0 1 -1 2 2 3 -1 2 1 2 2 2
	3 0 0 1 0 1 0 0 2 -1 2 1 4 0 2 -1 2 1 4 1
73	GINOCYBE GEOPHYLLA (SOW EX FRIKUM UK FREQ 17.0 MUSCARINE PROPOSED TREAT-
	MENT:PREVENT ABSORP,ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,
74	SUPPORTIVE TAXON HEIM (1931)LE GENRE INOCYBE,LECHEVALIER,PARIS CHEM
	CATALFOMO & EUGSTER (1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF &
75	MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS EU;N.AM;N.AF;N.ASIA
76	50 3 0 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 1 1 0 0 1 2 1 2 2 3
	0 0 1 0 1 0 0 -1 2 1 2 -1 2 1 4 0 -1 2 1 2 -1 2 1 4 1
77	GINOCYBE GRISEOLILACINA LGE UK FREQ 2.4 MUSCARINE PROPOSED TREATMENT:PRE-
	VENT ABSORP,ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE
78	TAXON HEIM (1931)LE GENRE INOCYBE,LECHEVALIER,PARIS CHEM CATALFOMO &
	EUGSTER (1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
79	TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS N.EU
80	38 3 0 -1 3 0 4 5 -1 2 1 2 2 1 1 0 0 1 -1 2 2 3 1 2 2 3
	0 0 1 0 1 0 0 2 2 0 2 2 1
81	GINOCYBE HIPTELLA BRES UK FREQ 1.2 MUSCARINE PROPOSED TREATMENT:PREVENT
	ABSORP,ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE
82	TAXON HEIM (1931)LE GENRE INOCYBE,LECHEVALIER,PARIS CHEM CATALFOMO &
	EUGSTER (1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
83	TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS EU;N.AF;N.AM
84	51 3 0 -1 3 0 2 4 -1 3 0 1 2 -1 2 1 2 1 1 0 1 1 -1 2 2 3
	-1 2 1 2 2 2 3 0 0 1 0 1 0 0 2 -1 2 1 4 0 2 -1 2 1 4
85	GINOCYBE HYSTRIX (FRI)KARST UK FREQ 4.3 MUSCARINE PROPOSED TREATMENT:PRE-
	VENT ABSORP,ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE
86	TAXON HEIM (1931)LE GENRE INOCYBE,LECHEVALIER,PARIS CHEM CATALFOMO &
	EUGSTER (1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
87	TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS EU;N.AM
88	49 3 0 -1 3 0 4 5 1 -1 2 1 2 1 1 0 0 1 -1 2 2 3 -1 2 1 2
	2 2 3 0 0 1 0 1 0 0 2 -1 3 1 2 4 0 2 -1 3 1 2 4 1
89	GINOCYBE LACERA (FRI)KUM UK FREQ 4.7 MUSCARINE PROPOSED TREATMENT:PREVENT
	ABSORP,ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE
90	TAXON HEIM (1931)LE GENRE INOCYBE,LECHEVALIER,PARIS CHEM CATALFOMO &
	EUGSTER (1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
91	TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS EU;N.AF;N.AM
92	43 3 0 -1 4 0 2 4 5 -1 2 0 1 -1 3 0 1 2 1 1 0 0 1 -1 2 3
	4 1 2 2 3 0 0 1 0 1 0 0 2 1 0 2 1 1
93	GINOCYBE LANUGINELLA (SCHR)LGE MUSCARINE PROPOSED TREATMENT:PREVENT ABSO
	ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE
94	TAXON PEARSON (1954)THE NATURALIST 117-139 CHEM CATALFOMO & EUGSTER
	(1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)
95	TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND REINHOLD,NY
	UPSETTING TO DANGEROUS EU;N.AF
96	48 3 0 -1 4 0 2 4 5 2 -1 2 1 2 1 1 0 0 1 -1 2 2 3 -1 2 1
	2 2 2 3 0 0 1 0 1 0 0 1 -1 2 2 4 0 1 -1 2 2 4 1

Table 7 (continued)

27	GINOCYBE LANUGINOSA (BUL EX FR)KUM UK FREQ 3.6 MUSCARINE PROPOSED TREATMENT PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU;N.AM	47	3	0	-1	3	0	4	5	0	-1	2	0	1	1	3	0	0	1	2	1	2	2	3	0	0	1	
		0	1	0	0	-1	2	0	1	-1	2	1	4	0	-1	2	0	1	-1	2	1	4	1					
4	GINOCYBE LUCIFUGA (FR)QUEL UK FREQ 0.4 MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU;N.AF	49	3	0	-1	3	0	2	4	0	-1	2	1	2	1	1	0	1	1	-1	2	2	3	-1	2	1	2	
		2	2	3	0	0	1	0	1	0	0	2	-1	3	1	2	4	0	2	-1	3	1	2	4	1			
12	GINOCYBE MACULATA BOUD UK FREQ 4.7 MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACT A 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU;N.AM	33	3	0	-1	3	0	4	5	1	-1	2	1	2	1	1	0	1	1	-1	2	2	3	1	2	2	3	
		3	0	1	0	1	0	0	-1	2	0	1	0	1	1													
20	GINOCYBE MIXTILIS (BRITZ) SAC SS KUHN UK FREQ 2.4 MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU;N.AM	50	3	0	-1	3	0	4	5	-1	2	0	1	-1	2	1	2	1	3	0	0	1	-1	2	1	2	-1	
		2	1	2	2	2	3	0	0	1	0	1	0	0	1	-1	2	2	4	0	1	-1	2	2	4	1		
28	GINOCYBE NAPIPES LGE UK FREQ 8.7 MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE SECRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED. LINCOFF & MITCHEL (1977) TOX AND HALLUC. MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU	38	3	0	-1	3	0	4	5	-1	2	0	1	-1	2	1	2	1	3	0	0	1	2	1	2	2	3	
		0	0	1	0	1	0	0	2	2	0	2	2	1														
38	GINOCYBE OBLECTABILIS (BRITZ) SACC MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON PEARSON (1954) THE NATURALIST OCT-DEC, 117-140 CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU;N.AM	46	3	0	-1	2	0	4	1	-1	2	1	2	1	3	0	0	1	-1	2	2	3	-1	2	1	2	2	
		2	3	0	0	1	0	1	0	0	-1	2	1	2	2	0	-1	2	1	2	2	1						
46	GINOCYBE OBSCUROIDES ORT MUSCARINE PROPOSED TREATMENT: PREVENT ABSORPT, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON ORTON (1960) TRANS BRIT MYCOL SOC 43, 276-277 CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU	40	3	0	-1	2	0	4	1	-1	2	1	2	1	1	0	0	1	2	1	2	2	3	0	0	1	0	
		1	0	0	2	-1	2	1	4	0	2	-1	2	1	4	1												
54	GINOCYBE PALLIDIPES ELL & EV MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU;N.AM	49	3	0	-1	2	0	4	1	-1	2	1	2	1	1	0	0	1	-1	2	2	3	1	2	2	3	0	
		0	1	0	1	0	0	-1	2	1	2	-1	2	0	2	0	-1	2	1	2	-1	2	0	2	1			
62	GINOCYBE OBSCURA (FR EX PERS) GILL UK FREQ 0.4 PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY UPSETTING TO DANGEROUS EU	46	3	0	-1	3	0	4	5	1	-1	2	1	2	1	1	0	0	1	-1	2	2	3	1	2	2	3	
		0	0	1	0	1	0	0	2	-1	3	1	2	4	0	2	-1	3	1	2	4	1						

Table 7 (continued)

3	6INOCYBE PATOULLARDI BRES MUSCARINE PROPOSED TREATMENT: PREVENT
4	ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
5	TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
6	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
7	TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
8	UPSETTING TO DANGEROUS EU; N. AM
9	44 3 0 -1 2 0 4 1 -1 2 1 2 1 1 0 1 1 -1 2 3 4 -1 2 1 2 2
10	2 3 0 0 1 0 1 -1 2 0 1 0 -1 2 1 2 0 1 1
11	6INOCYBE PERLATA (CKE) SACC MUSCARINE PROPOSED TREATMENT: PREVENT
12	ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
13	TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
14	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
15	TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
16	UPSETTING TO DANGEROUS EU
17	38 3 0 -1 2 0 4 -1 2 1 2 -1 2 1 2 1 1 0 0 1 -1 2 2 3 2 2
18	2 3 0 0 1 0 1 0 0 1 0 1 1
19	6INOCYBE POSTERULA (BRITZ) SACC UK FREQ 1.2 PROPOSED TREATMENT: PRE-
20	VENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUP-
21	PORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM
22	CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF &
23	MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
24	UPSETTING TO DANGEROUS EU
25	53 3 0 -1 3 0 4 5 -1 2 3 1 -1 3 0 1 2 1 1 0 0 1 2 1 2 2
26	3 0 0 1 0 1 0 0 -1 2 1 2 -1 3 0 1 2 0 -1 2 1 2 -1 3 0
27	1 2 1
28	6INOCYBE PRAETERVISA QUEL UK FREQ 3.2 PROPOSED TREATMENT: PREVENT
29	ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
30	HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
31	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
32	TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
33	UPSETTING TO DANGEROUS EU; N. AM; N. AF
34	51 3 0 -1 3 0 2 4 -1 3 0 1 2 1 1 3 0 0 1 -1 3 1 2 3 -1 2
35	1 2 2 2 3 0 0 1 0 1 0 0 2 -1 3 1 2 4 0 2 -1 3 1 2 4
36	1
37	6INOCYBE PUDICA KUHN UK FREQ 2.4 PROPOSED TREATMENT: PREVENT ABSORP,
38	ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
39	TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
40	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
41	TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
42	UPSETTING TO DANGEROUS EU
43	49 3 0 -1 3 0 4 5 0 -1 2 1 2 1 1 0 0 1 2 1 2 2 3 0 0 1
44	0 1 0 0 -1 2 1 2 -1 3 1 2 4 0 -1 2 1 2 -1 3 1 2 4 1
45	6INOCYBE QUELETII MRE & K MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP
46	ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
47	TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
48	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
49	TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
50	UPSETTING TO DANGEROUS S. EU; AFR.
51	52 3 0 -1 2 0 2 -1 2 1 -1 2 1 2 1 1 0 3 1 -1 2 2 3 2 2
52	2 3 0 0 1 0 1 0 0 -1 2 1 2 -1 2 1 4 0 -1 2 1 2 -1 2 1
53	4 1
54	6INOCYBE SAMBUICINA (FRIQUEL) MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP,
55	ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM
56	(1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO & EUGSTER (1970)
57	HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX AND HALLUC
58	MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
59	UPSETTING TO DANGEROUS EU
60	47 3 0 -1 3 0 4 5 -1 2 0 2 -1 2 1 2 1 1 0 1 -1 2 2 3 1
61	2 2 3 0 0 1 0 1 0 0 2 -1 2 1 4 0 2 -1 2 1 4 1
62	6INOCYBE UMBRINA BRES UK FREQ 4.3 MUSCARINE PROPOSED TREATMENT: PREVENT
63	ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
64	TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
65	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHELL (1977)
66	TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
67	UPSETTING TO DANGEROUS EU; N. AM
68	55 3 0 -1 3 0 4 5 -1 2 0 1 -1 2 0 1 1 3 0 0 1 -1 2 1 2 1
69	2 2 3 0 0 1 0 1 0 0 -1 2 1 2 -1 3 1 2 4 0 -1 2 1 2 -1
70	3 1 2 4 1

Table 7 (continued)

6	GINOCYBE XANTHOMELAS KUHN & BOURS UK FREQ 0.4 MUSCARINE PROP. TREATMENT:
7	PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE TAXON HEIM (1931) LE GENRE INOCYBE, LECHEVALIER, PARIS CHEM CATALFOMO &
8	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
9	TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
10	UPSETTING TO DANGEROUS EU;N.AM
11	53 3 0 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 1 3 0 0 1 3 -1 2 2 3
12	2 2 3 0 0 1 0 1 0 0 -1 2 1 2 -1 2 1 2 0 -1 2 1 2 -1 2
13	1 2 1
14	6GYMNOPIILUS AERUGINOSUS (PK) SING PSILOCYBIN/INDOLE HALLUC PROP. TREATMENT
15	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
16	DIAZEPAM, CHLORPROMAZINE TAXON HESLER (1969) N AM SPP OF GYMNOPIILUS, HAFNER
17	PUB, NY, LON CHEM HATFIELD ET AL (1978) LLOYDIA 41, 140-144 MED LINCOFF &
18	MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
19	HALLUCINOGEN, UPSETTING N.AM.
20	38 5 0 0 4 -1 3 0 1 2 1 1 1 1 -1 2 1 2 1 2 3 2 0 0 1
21	1 1 0 0 -1 2 0 1 2 0 0 2 1
22	6GYMNOPIILUS LUTEUS (PK) HESLER PSILOCYBIN/INDOLE HALLUC PROP. TREATMENT:
23	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
24	DIAZEPAM, CHLORPROMAZINE TAXON HESLER (1969) N AM SPP GYMNOPIILUS, HAFNER PU
25	NY, LON CHEM HATFIELD ET AL (1978) LLOYDIA 41, 140-144 MED LINCOFF & MIT-
26	CHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
27	HALLUCINOGEN, UPSETTING N.AM.
28	31 5 0 0 4 1 1 1 1 1 1 -1 2 1 2 1 2 3 2 0 0 1 0 0 0
29	0 2 0 0 2 1
30	6GYMNOPIILUS JUNIUNUS (FR) ORT PSILOCYBIN/INDOLE HALLUC PROP. TREATMENT:
31	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
32	DIAZEPAM, CHLORPROMAZINE TAXON HESLER (1969) N AM SPP GYMNOPIILUS, HAFNER PU
33	NY, LON CHEM HATFIELD ET AL (1978) LLOYDIA 41, 140-144 MED LINCOFF & MIT-
34	CHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
35	HALLUCINOGEN, UPSETTING EU;N.AM.
36	44 5 0 -1 3 0 4 5 -1 2 1 4 -1 2 1 2 1 1 1 1 1 2 1 2 3 2
37	0 0 1 0 -1 2 0 1 0 0 -1 2 0 1 2 0 0 2 1
38	6GYMNOPIILUS VALIDIPES (PK) HESLER PSILOCYBIN/INDOLE HALLUC PROP. TREATMENT:
39	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
40	DIAZEPAM, CHLORPROMAZINE TAXON HESLER (1969) N AM SPP GYMNOPIILUS, HAFNER PU
41	NY, LON CHEM HATFIELD ET AL (1978) LLOYDIA 41, 140-144 MED LINCOFF & MITCHEL
42	(1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
43	HALLUCINOGEN, UPSETTING N.AM.
44	37 5 0 0 5 -1 2 1 2 1 1 1 1 0 -1 2 1 2 1 2 3 2 0 0 1 0
45	-1 2 0 1 0 0 0 2 0 0 2 1
46	6GYMNOPIILUS VIRIDANS MURR PSILOCYBIN/INDOLE HALLUC PROP. TREATMENT:
47	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.
48	DIAZEPAM, CHLORPROMAZINE TAXON HESLER (1969) N AM SPP GYMNOPIILUS, HAFNER P
49	NY, LON CHEM HATFIELD ET AL (1978) LLOYDIA 41, 140-144 MED LINCOFF & MITCHEL
50	(1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY
51	HALLUCINOGEN, UPSETTING N.AM.
52	32 5 0 0 5 2 1 1 1 1 1 -1 2 1 2 1 2 3 2 0 0 1 0 0 0
53	0 -1 2 1 2 1 1
54	6GALERINA AUTUMNALIS (PK) SING/SM CYCLOPEPTIDES PROPOSED TREATMENT: WATER,
55	ELECTROLYTE, BLOOD BALANCE, HYPERTONIC GLUCOSE INF, PENICILLIN, THIOCTIC
56	ACID, EXCHANGE TRANSFUS. TAXON SINGER & SMITH (1964) GENUS GALERINA EARLE,
57	HAFNER, NY, LON CHEM TYLER ET AL (1963) LLOYDIA 26, 154-157 MED BERTELLI ET
58	AL (1977) ED. CURRENT PROBLEMS IN CLIN BIOCHEM 7, HUBER, BERN, STUTT, VIENNA
59	DEADLY EU;N.AM.
60	47 0 2 -1 3 0 4 5 1 2 1 1 1 1 -1 2 2 3 -1 2 1 2 2 0 0
61	0 0 1 0 1 0 0 -1 2 1 2 1 0 -1 2 1 2 -1 2 1 2 1
62	6GALERINA MARGINATA (FR) KUHN CYCLOPEPTIDES PROP. TREATMENT: WATER, ELECTROLY
63	BLOOD BALANCE, HYPERTONIC GLUCOSE INF, PENICILLIN, THIOCTIC ACID, EXCHANGE
64	TRANSFUSION TAXON SINGER & SMITH (1964) GENUS GALERINA EARLE, HAFNER PUB, N
65	LON CHEM TYLER ET AL (1963) LLOYDIA 26, 154-157 MED BERTELLI ET AL ED. (1977
66	CURRENT PROBLEMS IN CLIN BIOCHEM 7, HUBER, BERN, STUTT, VIENNA DEADLY EU;N.AM
67	47 0 2 -1 3 0 4 5 0 -1 2 1 2 1 1 1 1 -1 2 2 3 1 2 0 0
68	0 0 1 0 1 0 0 1 -1 2 1 2 0 -1 2 1 2 -1 2 1 2 1

Table 7 (continued)

4	6GALERINA VENENATA SMITH CYCLOPEPTIDES PROPOSED TREATMENT: WATER, ELECTRO-																									
5	LYTE, BLOOD BALANCE, HYPERTONIC GLUCOSE INF., PENICILLIN, THIOCTIC ACID, EXCH																									
6	TRANSFUSION TAXON. SINGER & SMITH (1964) GENUS GALERINA EARLE, HAFNER PUB. N																									
7	LON CHEM TYLER ET AL (1963) LLOYDIA 26, 154-157 MED BERTELLI ET AL. EDS (197																									
8	CURRENT PROBLEMS IN CLIN BIOCHEM 7, HUBER, BERN, STUTT, VIENNA																									
9	DEADLY N.A.M																									
10	46	0	2	0	2	-1	2	2	3	1	1	1	1	1	-1	2	2	3	2	2	0	0	0	1	0	
11	1	0	0	-1	2	1	2	-1	2	1	2	0	-1	2	1	2	-1	2	1	2	1					
12	6COPRINUS ATRAMENTARIUS (BUL EX FR) FR UK FREQ 9.1 COPRINE PROPOSED TREATMENT:																									
13	STOP ALCOHOL CONSUMP! PROPANOLOL IF CARDIAC ARRHYTHM., ISOTONIC FLUIDS FOR HYPO-																									
14	TENSION TAXON ORTON & WATLING (1979) BRIT. FUNGUS FLORA, 2 COPRINACEAE, HMSO EDIN.																									
15	CHEM. HATFIELD, SCHAUMBERG (1975) LLOYDIA 38:6, 489-496 MED LINCOFF & MITCHEL																									
16	(1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY																									
17	UPSETTING, CONDITIONAL COSMOP.																									
18	52	2	0	-1	6	0	1	2	3	4	5	-1	2	2	4	-1	2	1	2	3	1	1	0	0	0	-1
19	2	2	3	1	3	5	1	0	0	0	1	1	0	0	-1	3	0	1	2	-1	2	0	2	0	2	
20	6PANAEOULUS FOENISECII (PERS EX FR) HRE UK FREQ 13.5 INDOLE HALLUC. PROPOSED																									
21	TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET																									
22	POSSIB DIAZEPAM, CHLORPROMAZINE TAXON OLA'H (1969) GENRE PANAEOULUS, REV MYCOL																									
23	MEM HORS SERIE CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572																									
24	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R																									
25	NY HALLUCINOGEN, UPSETTING COSMOPOLITAN																									
26	44	5	0	-1	6	0	1	2	3	4	5	2	-1	2	1	2	3	0	1	1	1	0	4	-1	2	2
27	3	3	3	2	0	0	0	0	1	0	0	0	-1	3	1	2	3	1	1							
28	6COPRINUS INSIGNIS PECK UK FREQ 0.4 SUSPECT COPRINE PROPOSED TREATMENT: STOP																									
29	ALCOHOL CONSUMP! PROPANOLOL IF CARDIAC ARRHYTHM., ISOTONIC FLUIDS FOR HYPO-																									
30	TENSION TAXON ORTON & WATLING (1979) BRIT. FUNGUS FLORA 2 COPRINACEAE, HMSO EDIN.																									
31	CHEM. HATFIELD, SCHAUMBERG (1975) LLOYDIA 38:6, 489-496 MED LINCOFF & MITCHEL																									
32	(1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY..																									
33	UPSETTING, CONDITIONAL N.A.M; EU																									
34	48	2	0	-1	2	0	4	-1	2	2	4	-1	2	1	2	3	1	1	1	0	0	-1	2	3	4	2
35	3	5	1	0	0	0	0	1	0	0	-1	3	0	1	2	2	0	2	-1	2	0	1	1			
36	6COPRINUS MICACEUS (BULL EX FR) FR UK FREQ 17.4 SUSPECT COPRINE PROPOSED																									
37	TREATMENT: STOP ALCOHOL CONSUMP. PROPANOLOL IF CARDIAC ARRHYTHM., ISOTONIC																									
38	FLUIDS FOR HYPOTENSION TAXON ORTON & WATLING (1979) BRIT. FUNGUS FLORA, 2 COPRIN-																									
39	ACEAE, HMSO EDIN. CHEM HATFIELD & SCHAUMBERG (1975) LLOYDIA 38:6, 489-496 MED																									
40	LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD,																									
41	NY UPSETTING, CONDITIONAL COSMOPOLITAN																									
42	53	2	0	-1	6	0	1	2	3	4	5	-1	2	1	3	-1	4	0	1	2	3	3	1	1	0	0
43	0	-1	2	1	2	1	3	5	1	0	0	0	1	0	0	-1	2	1	2	2	0	2	-1	2		
44	6COPRINUS AFRICANUS PEGLER SUSPECT COPRINE PROPOSED TREATMENT: STOP																									
45	ALCOHOL CONSUMP! PROPANOLOL IF CARDIAC ARRHYTHM., ISOTONIC FLUIDS FOR HYPO-																									
46	TENSION TAXON PEGLER (1966) PERSOONIA 4, 82 CHEM HATFIELD & SCHAUMBERG																									
47	(1975) LLOYDIA 38:6, 489-496 MED LINCOFF & MITCHEL (1977) TOX & HALLUC																									
48	MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY..																									
49	UPSETTING, CONDITIONAL AFR.																									
50	41	2	0	3	1	0	3	1	1	0	1	0	-1	2	0	1	-1	2	0	1	3	5	1	0	0	0
51	0	1	0	0	2	-1	2	1	2	0	2	-1	2	1	2	1										
52	6COPRINUS ALUMINATUS (ROMAG) ORT UK FREQ 1.2 SUSPECT COPRINE PROPOSED																									
53	TREATMENT: STOP ALCOHOL CONSUMP! PROPANOLOL IF CARDIAC ARRHYTHM., ISOTONIC																									
54	FLUIDS FOR HYPOTENSION TAXON ORTON (1969) NOTES ROY BOT GD EDIN 29, 86-93																									
55	CHEM HATFIELD & SCHAUMBERG (1975) LLOYDIA 38:6, 489-496 MED LINCOFF &																									
56	MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY.																									
57	UPSETTING, CONDITIONAL EU																									
58	49	2	0	-1	3	0	4	5	-1	3	1	4	5	-1	2	1	2	3	1	1	0	1	0	2	1	3
59	5	1	0	0	0	0	1	0	0	-1	2	1	2	-1	2	0	2	0	2	-1	2	0	2	1		
60	6PSILOCYBE MERDARIA (FR) RICK PSILOCYBIN/INDOLE HALLUC. PROPOSED TREATMEN																									
61	TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB.																									
62	DIAZEPAM, CHLORPROMAZINE TAXON PEGLER (1977) PRELIM AGARIC FLORA E. AFRICA, HMSO																									
63	LON CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF &																									
64	MITCHEL (1977) TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, N																									
65	NY HALLUCINOGEN, UPSETTING COSMOPOLITAN																									
66	43	5	0	-1	5	0	2	3	4	5	3	-1	3	0	1	2	3	0	1	0	1	0	-1	2	3	4
67	-1	2	2	3	3	3	2	0	0	1	0	1	0	0	0	1	1	1								

Table 7 (continued)

4	6PANAEOLOUS ATER (LGEIK & R UK FREQ 1.2 PSILOCYBIN/INDOLE HALLUC. PROPOSED
6	TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUI
8	POSSIB. DIAZEPAM, CHLORPROMAZINE TAXON HORA (1957) THE NATURALIST JUL-SEP 7
10	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MIT-
12	CHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...
14	HALLUCINOGEN, UPSETTING EU
16	44 5 0 -1 3 0 4 5 2 -1 3 0 1 2 3 0 1 0 1 0 -1 2 3 4 2 3
18	3 2 0 0 0 0 -1 3 0 1 2 0 0 0 -1 0 0 1 0
20	6PANAEOLOUS CAMPANULATUS (BULL EX FR) QUEL UK FREQ 3.6 PSILOCYBIN/INDOLE HALLU
22	PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BE-
24	HAV, QUIET POSSIB DIAZEPAM, CHLORPROMAZINE TAXON MOLLER FH (1945) FUNGI OF THE
26	FAEROES, MUNKSGAARD, COPENH. CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572
28	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY
30	HALLUCINOGEN SUSPECT, UPSETTING COSMOPOLITAN
32	46 5 0 -1 6 0 1 2 3 4 5 3 -1 2 1 2 3 0 1 0 1 0 4 -1 2 3
34	4 3 3 2 0 0 0 0 1 0 0 -1 2 0 1 -1 2 1 2 1 1
36	6PANAEOLOUS CASTANEIFOLIUS (MURR) SM PSILOCYBIN/INDOLE HALLUC. PROPOSED
38	TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV,
40	QUIET, POSSIB DIAZEPAM, CHLORPROMAZINE TAXON OLA'H G (1968) REV MYCOL 33, 284-290
42	685 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MIT-
44	CHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...
46	HALLUCINOGEN SUSPECT, UPSETTING EU; N. AM.
48	37 5 0 -1 3 0 4 5 2 -1 2 1 2 3 1 1 1 1 0 4 -1 2 2 3 3 3
50	2 0 0 0 0 1 0 0 0 1 1 1
52	6PANAEOLOUS FIMICOLA (FR) GILL PSILOCYBIN/INDOLE HALLUC PROPOSED TREATMENT:
54	PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB
56	DIAZEPAM, CHLORPROMAZINE TAXON PEGLER (1977) PRELIM AGARIC FLORA E. AFRICA,
58	HMSO, LON CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF
60	& MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY...
62	SUSPECT HALLUCINOGEN, UPSETTING COSMOPOLITAN
64	49 5 0 -1 5 0 2 3 4 5 -1 2 2 3 -1 3 0 1 2 3 0 1 0 1 0 -1
66	2 3 4 -1 2 2 3 3 3 2 0 0 0 0 1 0 0 0 -1 2 1 3 1 1
68	6PANAEOLOUS PAPILIONACEUS (BULL EX FR) QUEL UK FREQ 1.2 INDOLE HALLUC PROPOS
70	TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIE
72	POSSIB DIAZEPAM, CHLORPROMAZINE TAXON PEGLER (1977) PRELIM AGARIC FLORA E.
74	AFRICA, HMSO, LON CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED
76	LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, N
78	NY, SUSPECT HALLUCINOGEN, UPSETTING COSMOPOLITAN
80	47 5 0 -1 6 0 1 2 3 4 5 -1 2 2 3 -1 3 0 1 2 3 0 1 0 1 0
82	4 3 3 3 2 0 0 0 0 1 -1 2 0 1 0 -1 2 0 1 0 1 1
84	6PANAEOLOUS RETIKUGIS (FR) GILL UK FREQ 0.4 INDOLE HALLUC. SUSPECT PROPOSED
86	TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUI
88	POSSIB DIAZEPAM, CHLORPROMAZINE TAXON HORA (1957) THE NATURALIST JUL-SEPT
90	77-88 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF &
92	MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY.
94	SUSPECT HALLUCINOGEN, UPSETTING EU
96	43 5 0 -1 3 0 4 5 -1 2 2 3 -1 2 1 2 3 0 1 0 1 0 4 -1 2 2
98	3 3 3 2 0 0 0 0 -1 2 0 1 0 0 0 0 -1 1 1
100	6PANAEOLOUS SPHINCTRINUS (FR) QUEL UK FREQ 11.5 SUSPECT INDOLE HALLUC.
102	PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BE-
104	HAV, QUIET POSSIB DIAZEPAM, CHLORPROMAZINE TAXON PEGLER (1977) PRELIM AGARIC
106	FLORA E, AFRICA, HMSO, LON CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572
108	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND
110	R, NY. SUSPECT HALLUCINOGEN, UPSETTING COSMOPOLITAN
112	51 5 0 -1 6 0 1 2 3 4 5 -1 2 2 3 -1 3 0 1 2 3 0 1 0 1 0
114	4 -1 2 2 3 3 3 2 0 0 0 0 -1 2 0 1 0 0 0 -1 3 0 1 2 1
116	1
118	6PANAEOLOUS SUBBALTEATUS (BERK & BRISACC UK FREQ 1.2 INDOLE HALLUC? PROP
120	TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET
122	POSSIB DIAZEPAM, CHLORPROMAZINE TAXON HORA (1957) THE NATURALIST JUL-SEP 77-88
124	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH
126	(1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...
128	SUSPECT HALLUCINOGEN, UPSETTING COSMOPOLITAN
130	51 5 0 -1 6 0 1 2 3 4 5 -1 2 2 3 -1 3 0 1 2 3 0 1 0 1 0
132	4 -1 2 2 3 3 3 2 0 0 0 0 -1 2 0 1 0 0 0 -1 3 0 1 2 1
134	1

Table 7 (continued)

4	6ANELLARIA PHALAENARUM (FRIQUEL SUSPECT HALLUC/INDOLES PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB. DIAZEPAM, CHLORPROMAZINE TAXON HORA (1957) THE NATURALIST, JUL-SEP 77-88																									
6	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...																									
8	SUSPECT HALLUCINOGEN, UPSETTING EU;N.AM																									
	42	5	0	-1	3	0	4	5	3	-1	2	1	2	3	0	1	0	1	0	-1	2	4	5	-1	2	3
11	4	3	3	2	0	0	0	0	1	1	0	1	0	0	1	0	0									
12	6ANELLARIA SEMIOVATA (SOW EX FR) PEARLS & DENNIS UK FREQ 5.9 SUSPECT INDOLE HALLUC. PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB DIAZEPAM, CHLORPROMAZINE TAXON HORA ((1957) THE																									
14	NATURALIST JUL-SEP, 77-88 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA, 42, 1557-1572																									
15	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REI																									
16	NY. SUSPECT HALLUCINOGEN, UPSETTING EU;N.AM																									
	50	5	0	-1	3	0	4	5	3	-1	3	0	1	2	3	0	1	0	1	0	-1	2	4	5	-1	2
17	3	4	3	3	2	0	0	0	0	1	1	0	0	0	0	-1	2	1	2	-1	3	0	1	2	0	
20	6COPELANDIA CAMBODGINIENSIS OLA'H/HEIM INDOLE HALLUC PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB																									
22	DIAZEPAM, CHLORPROMAZINE TAXON OLA'H G (1968) REV MYCOL 33, 284-290																									
24	CHEM HOFMANN ET AL (1959) HE V CHIM ACTA 42, 1557-1572 MED LINCOFF & MITC (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY..																									
26	HALLUCINOGEN, UPSETTING S.E.ASIA																									
28	49	5	0	-1	2	2	3	3	-1	3	0	1	2	3	0	1	0	1	0	-3	-1	2	2	3	3	3
29	2	0	0	0	0	1	0	0	0	-1	2	0	1	0	-1	2	1	2	-1	3	0	1	3	1		
30	6COPELANDIA CYANESCENS (BERK & BR) SING INDOLE HALLUC PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB																									
32	DIAZEPAM, CHLORPROMAZINE TAXON SINGER R (1968) LILLOA 30, 124-126																									
34	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...																									
36	HALLUCINOGEN, UPSETTING PANTROPICAL																									
38	50	5	0	-1	2	2	3	3	-1	3	0	1	2	3	0	1	0	1	0	4	-1	3	2	3	4	3
39	3	2	0	0	0	0	1	0	0	-1	2	0	1	0	-1	2	1	2	-1	1	1	2	4	1		
42	6COPELANDIA TROPICALIS OLA'H INDOLE HALLUC. PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB. DIAZEPAM																									
44	CHLORPROMAZINE TAXON OLA'H (1968) REV MYCOL 33, 284-290 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...																									
46	HALLUCINOGEN, UPSETTING AFR;S.ASIA																									
48	48	5	0	3	3	-1	3	0	1	2	3	0	1	0	1	0	3	-1	2	2	3	3	3	2	0	0
49	0	0	-1	2	0	1	0	0	0	-1	2	0	1	0	-1	2	1	2	-1	2	2	4	1			
50	6PANAEOULUS AFRICANUS OLA'H INDOLE HALLUC. PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB. DIAZEPAM																									
52	CHLORPROMAZINE TAXON OLA'H (1968) REV MYCOL 33, 284-290																									
54	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...																									
56	HALLUCINOGEN, UPSETTING AFR.																									
58	39	5	0	3	3	-1	2	0	1	3	0	1	0	1	0	-1	2	3	4	-1	2	3	4	3	3	2
59	0	0	0	0	-1	2	0	1	0	0	0	0	1	0												
60	6PANAEOULUS MICROSPORUS OLA'H (CAILLEUX INDOLE HALLUC. PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB																									
62	DIAZEPAM, CHLORPROMAZINE TAXON OLA'H G (1968) REV MYCOL 33, 284-290																									
64	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCH (1977) TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...																									
66	SUSPECT HALLUCINOGEN, UPSETTING AF																									
68	45	5	0	3	3	-1	2	0	1	3	0	1	0	1	0	-1	2	1	2	1	3	3	2	0	0	0
69	0	1	0	0	-1	2	0	1	-1	3	0	1	2	0	0	-1	2	0	1	1						
70	6PSILOCYBE BANDERILLIENSIS GUZ INDOLE HALLUC PROPOSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB. DIAZEPAM																									
72	CHLORPROMAZINE TAXON GUZMAN (1978) NOVA HEDWIGIA 29, 625-664 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND REINHOLD, NY...																									
74	HALLUCINOGEN, UPSETTING MEX																									
76	38	5	0	2	1	1	3	0	1	0	1	0	-1	2	1	2	-1	2	0	1	3	3	2	0	0	1
77	0	1	0	0	0	1	0	0	-1	2	1	2	1													

Table 7 (continued)

4	6PSILOCYBE HEIMII GUZ INDOLE HALLUC PROPOSED TREATMENT:PREVENT ABSORP, ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB DIAZEPAM,CHLOR- PROMAZINE TAXON GUZMAN (1978) NOVA HEDWIGIA 29,625-664 CHEM HOFMANN ET 6 AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING,VAN NUSTRAND REINHOLD,NY...
8	HALLUCINOGEN, UPSETTING MEX 37 5 0 2 1 1 3 0 1 0 1 0 1 0 3 3 2 0 0 1 0 1 0 0 0 -1 10 4 0 1 2 3 0 0 -1 2 1 2 1
12	6PSILOCYBE GALINDII GUZ INDOLE HALLUC PROPOSED TREATMENT:PREVENT ABSORP, ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB DIAZEPAM, CHLORPROMAZINE TAXON GUZMAN (1978) NOVA HEDWIGIA,29,625-664 CHEM HOFMANN 14 ET AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF, & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING,VAN NUSTRAND REINHOLD,NY...
16	HALLUCINOGEN, UPSETTING MEX 36 5 0 -1 2 2 3 -1 2 0 2 1 3 0 1 0 1 0 -1 2 2 3 2 3 3 2 18 0 0 1 0 1 0 0 0 1 0 0 1 1
20	6PSILOCYBE HERRERAEE GUZ INDOLE HALLUC PROPOSED TREATMENT:PREVENT ABSORP, ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB DIAZEPAM,CHLOR- PROMAZINE, TAXON GUZMAN (1978) NOVA HEDWIGIA 29,625-664 CHEM HOFMANN ET 22 AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING,VAN NUSTRAND REINHOLD,NY...
24	HALLUCINOGEN, UPSETTING MEX 35 5 0 2 -1 2 1 4 1 3 0 1 0 1 0 0 0 3 3 2 0 0 1 0 1 0 26 0 0 1 0 0 -1 2 1 2 1
28	6PSILOCYBE RZEDOWSKII GUZ INDOLE HALLUC PROPOSED TREATMENT:PREVENT ABSORP ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB DIAZEPAM,CHLOR- PROMAZINE TAXON GUZMAN (1978) NOVA HEDWIGIA 29,625-664 CHEM HOFMANN ET A 30 (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HAL MUSHROOM POISONING,VAN NUSTRAND REINHOLD,NY...
34	HALLUCINOGEN, UPSETTING MEX 32 5 0 2 1 1 3 0 1 0 1 0 1 0 3 3 2 0 0 1 0 1 0 0 0 2 36 0 0 -1 2 1 2 1
38	6PSILOCYBE VERAECRUCIS GUZ & ORTIZ INDOLE HALLUC PROPOSED TREATMENT:PREVE ABSORP,ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB DIAZEPAM CHLORPROMAZINE TAXON GUZMAN (1978) NOVA HEDWIGIA 29,625-664 CHEM HOFMANN 40 ET AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING,VAN NUSTRAND REINHOLD,NY...
44	HALLUCINOGEN, UPSETTING MEX 38 5 0 3 1 1 3 0 1 0 1 0 1 -1 2 0 1 3 3 2 0 0 1 0 1 0 46 0 0 -1 2 1 2 0 0 -1 2 1 2 1
48	SCORTINARIUS GENTILIS (FR)FR UK FREQ 0.4 SUSPECT ORELLANINE PROPOSED TREAT- MENT SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER STUT.NY CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21,1931-34 MED FLAMMER 46 R (1980) DIFFERENTIALDIAGNOSE DER PILZVERGIFTUNGEN,FISCHER,STUTT,NY DEADLY EU
50	37 1 3 -1 3 0 4 5 0 -1 2 1 2 1 1 1 1 1 -1 2 1 2 -1 2 1 2 52 2 1 0 0 0 1 0 1 1 1 1 1
54	SCORTINARIUS LIMONIUS (FR EX FR)FR ORELLANINE SUSPECT PROPOSED TREATMENT SYMPTOMATIC TAXON MOSER M (1979) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER, 52 STUT.NY CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21,1931-34 MED FLAMMER (1980) DIFFERENTIALDIAGNOSE UER PILZVERGIFTUNGEN,FISCHER,STUTT. NY SUSPECT DEADLY, EU
56	37 1 3 -1 3 0 4 5 0 -1 2 1 2 1 1 1 1 1 -1 2 1 2 -1 2 1 2 58 2 1 0 0 0 1 0 1 1 1 1 1
60	SCORTINARIUS GYANITES FR UK FREQ 0.4 SUSPECT ORELLANINE PROPOSED TREATMEN SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLOFA IIB/2, STUTT,NY CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21,1931-34 MED FLAMMER R 50 DIFFERENTIALDIAGNOSE DER PILZVERGIFTUNGEN,FISCHER,STUTT.NY. SUSPECT DEADLY EU
62	33 1 3 -1 3 0 4 5 1 -1 2 1 2 1 1 1 1 1 2 1 2 1 0 0 0 1 64 0 1 0 0 0 0 1 1
64	SCORTINARIUS ORELLANOIDES MRY ORELLANINE SUSPECT PROPOSED TREATMENT:SYM- TOMATIC TAXON ORTON PD (1976)KEM BULL 31:3,707-721 CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L. 21,1931-34 MED. GERAULT & GIRRE (1977) BULL SOC MYCOL FRANCE 93,373-405 SUSPECT DEADLY EU
66	37 1 3 -1 3 0 4 5 1 -1 2 1 2 1 1 1 1 1 -1 2 2 3 -1 2 2 3 68 2 1 0 0 0 1 0 1 1 1 1 1

Table 7 (continued)

4	SCORTINARIUS SPLENDENS HRV SUSPECT ORELLANINE PROPOSED TREATMENT: SYMPTOMA
	TAXON MOSEK M (1978) KLEINE KRYPTOGAMENFLORA IIB/2, FISCHER, STUT. NY.
6	CHEM ANTKOWIAK & GESSNER (1979) TETRAEDRON L. 21, 1931-34 MED FLAMMER R
	(1980) DIFFERENTIALDIAGNOSE DER PILZVERGIFTUNGEN, FISCHER, STUT. NY.
8	SUSPECT DEADLY EU
	34 1 3 -1 3 0 4 5 1 -1 2 1 2 1 1 1 1 -1 2 2 3 1 2 1 0
10	0 0 1 0 1 1 1 1 1
12	6CONOCYBE CYANOPUS (ATK) KUHN UK FREQ 0.4 PSILOCYBIN/INDOLE HALLUCINOGEN
	PROPOSED TREATMENT: PREVENT ABSORPTION, ENHANCE EXCRET, RESTRAIN SELF DESTR
	BEHAV, QUIET, POSSIB DIAZEPAM, CHLORPROMAZINE TAXON BENEDICT ET AL (1967)
14	LLOYDIA 30, 150-157 CHEM BENEDICT ET AL (1962) LLOYDIA 25, 156-159 MED LIN
	& MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY
16	HALLUCINOGEN, UPSETTING N.AM, EU
	33 5 0 -1 3 0 4 5 2 -1 2 1 2 1 1 0 1 0 1 1 2 3 2 0 0 0
18	0 1 0 0 1 2 1 1
20	6CONOCYBE FILARIS (FR) KUHN UK FREQ 1.6 CYCLOPEPTIDES PROPOSED TREATMENT:
	WATER, ELECTROLYTE (BLOOD BALANCE, HYPERTONIC GLUCOSE INF., PENICILLIN,
	THIOCTIC ACID, EXCHANGE TRANSFUSION TAXON KUHN R (1935) ENCYCLOP. MYCOL,
22	PARIS 7 CHEM BRADY ET AL (1975) LLOYDIA 38, 172-179 MED BERTELLI, FOURNIER
	ET AL (1977) CURRENT PROBLEMS IN CLIN BIOCHEM 7, HUBER, BERN, STUT. VIENNA
24	DEADLY EU; N.AM.
	51 0 2 -1 3 0 4 5 -1 2 1 2 -1 2 1 2 1 1 0 1 0 -1 2 2 3 -1
26	2 1 2 2 0 0 0 0 0 0 -1 2 0 1 0 8 -1 2 8 -1 -1 2 1 2 1
	1
28	6CONOCYBE SILIGINEOIDES HEIM PSILOCYBIN/INDOLE HALLUC PROPOSED TREATMENT:
	PREVENT ABSORPTION, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POS
30	DIAZEPAM, CHLORPROMAZINE TAXON HEIM (1956) COMPTES RENDUS ACAD. SC. 242, 1390
	CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42, 1557-1572 MED LINCOFF & MITCHE
32	(1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY
	HALLUCINOGEN, UPSETTING MEX
34	35 5 0 2 4 1 1 1 0 -1 0 4 -1 2 2 3 2 3 2 0 0 0 0 0 0 0
	-1 2 0 1 -1 2 2 3 1 1
36	6CONOCYBE SMITHII WATL PSILOCYBIN/INDOLE HALLUC PROPOSED TREATMENT: PREVEN
	ABSORPTION, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB,
38	DIAZEPAM, CHLORPROMAZINE TAXON & CHEM BENEDICT ET AL (1967) LLOYDIA 30,
	150-157 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING,
40	VAN NOSTRAND R, NY. HALLUCINOGEN, UPSETTING N.AM.
	41 5 0 0 -1 2 2 5 -1 2 0 1 1 1 0 1 0 -1 2 1 2 1 2 3 2 0
42	0 0 0 1 0 0 -1 2 0 1 -1 2 1 2 1 1
44	SHYGROPHORUS LONICUS (SCOP EX FR) FR UK FREQ 10.3 UNKNOWN PROPOSED
	TREATMENT: SYMPTOMATIC TAXON, MOELLER FH (1945) FUNGI OF THE FAEROES,
	MUNKSGAARD, COPENH. CHEM. NONE
46	MED. FLAMMER R (1980) DIFFERENTIALDIAGNOSE DER PILZVERGIFTUNGEN, FISCHER,
	STUT. NY. UPSETTING EU; N.AM
48	49 6 0 -1 4 0 2 4 5 2 -1 3 1 2 3 0 1 1 0 0 1 -1 3 1 2 3
	-1 2 1 2 0 6 3 0 0 1 0 -1 2 0 1 1 1 0 1 -1 2 0 1 1
50	SHYGROPHORUS NIGRESCENS (QUEL) QUEL UK FREQ 6.7 UNKNOWN PROPOSED
	TREATMENT: SYMPTOMATIC TAXON, PEGLER (1977) PRELIM AGARIC FLORA OF E.
52	AFRICA, HMSU. LON CHEM NONE
	MED FLAMMER R (1980) DIFFERENTIALDIAGNOSE DER PILZVERGIFTUNGEN, FISCHER
54	STUT. NY. UPSETTING COSMOPOLITAN
	43 6 0 -1 6 0 1 2 3 4 5 2 -1 3 1 2 0 1 1 0 0 1 -1 2 2 3
56	1 0 6 3 0 0 1 0 1 1 1 0 1 -1 2 0 1 1
58	6CLITOCYBE ANGUSTISSIMA (LASCH) KUM UK FREQ 0.4 MUSCARINE PROPOSED TREAT-
	MENT: PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS,
	SUPPORTIVE TAXON KUHN R & ROMAGNESI (1977) COMPLEMENT FLORE ANALYTIQUE,
60	MASSON, PARIS CHEM, CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851
	MED LINCOFF & MITCHEL (1977) TOX & HALLUC. MUSHROOM POISONING, VAN NOSTRAN
62	REIN., NY UPSETTING TO DANGEROUS EU
	34 3 0 -1 2 0 4 -1 2 1 2 -1 2 1 2 0 1 1 0 0 1 0 0 0 2 3
64	0 0 1 0 1 0 1 1 1
66	6CLITOCYBE CANDICANS (PERS EX FR) KUM UK FREQ 0.4 MUSCARINE PROPOSED TREAT-
	MENT PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUP
	PORTIVE TAXON HARMAJA H (1969) KARSTENIA 10, 5-168 CHEM CATALFOMO &
68	EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
	TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R., NY UPSETTING TO DANGEROUS
70	EU
	38 3 0 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 0 1 1 0 0 1 -1 2 0 1
72	0 0 2 3 0 0 1 0 1 0 1 1 1

Table 7 (continued)

6CLITOCYBE CERUSSATA (FR)KUM UK FREQ 1.2 MUSCARINE PROPOSED TREATMENT:
 9 PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS TAXON
 BIGELOW H E (1965) LLOYDIA 28:2, 139-180 CHEM CATALFOMO & EUGSTER (1970)
 10 HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX & HALLUC
 MUSHROOM POISONING, VAN NOSTRAND R., NY UPSETTING TO DANGEROUS EU ;N.AM.

12
 36 3 3 -1 3 0 4 5 -1 3 0 1 2 -1 2 1 2 0 1 1 0 0 1 0 0 0
 14 2 3 0 0 1 0 1 0 1 1 1

6CLITOCYBE DEALBATA (SW EX FR)KUM MUSCARINE PROPOSED TREATMENT: PREVENT
 16 ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
 TAXON HARMAJA H (1969) KARSTENIA 10, 5-168 CHEM CATALFOMO & EUGSTER
 18 (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977) TOX &
 HALLUCINOGENIC MUSHROOM POISONING, VAN NOSTRAND R., NY UPSETTING TO
 20 DANGEROUS EU; N.AM

22 32 3 0 -1 3 0 4 5 2 -1 2 1 2 0 1 1 0 0 1 0 0 0 2 3 0 0
 1 0 1 0 1 1 1

6CLITOCYBE DIATRETA (FR EX FR)KUM MUSCARINE PROPOSED TREATMENT: PREVENT
 24 ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
 TAXON RICKEN A (1914) DIE BLATTERPILZE CHEM CATALFOMO &
 26 EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R., NY UPSETTING TO
 28 DANGEROUS EU

30 32 3 0 -1 3 0 4 5 0 -1 2 1 2 0 1 1 0 0 1 0 0 0 2 3 0 0
 1 0 1 0 1 1 1

6CLITOCYBE ERICETORUM BULL EX QUÉL MUSCARINE PROPOSED TREATMENT:
 32 PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPOR
 TIVE TAXON HARMAJA H (1969) KARSTENIA 10, 5-168 CHEM CATALFOMO &
 34 EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R., NY UPSETTING TO
 36 DANGEROUS EU

38 32 3 0 -1 3 0 4 5 2 -1 2 1 2 0 1 1 0 0 1 0 0 0 2 3 0 0
 1 0 1 0 1 1 1

6CLITOCYBE FESTIVA FAVRE MUSCARINE PROPOSED TREATMENT: PREVENT ABSORP
 40 ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
 TAXON FAVRE (1955) CHAMPIGNONS PARC NAT. SUISSE, LIESTAL CHEM CATALFOMO &
 42 EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R., NY UPSETTING TO
 44 DANGEROUS, EU

46 34 3 0 0 1 -1 2 1 2 0 1 1 0 0 1 -1 2 0 1 -1 2 0 1 0 2 3
 0 0 1 0 1 0 1 1 1

6CLITOCYBE PHYLLOPHILA (FR)KUM UK FREQ 0.8 MUSCARINE PROPOSED TREATMENT:
 48 PREVENT ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS,
 SUPPORTIVE TAXON BIGELOW H E (1965) LLOYDIA 28:2, 139-180 CHEM CATALFOMO &
 50 EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF & MITCHEL (1977)
 TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R., NY UPSETTING TO
 52 DANGEROUS EU; N.AM.

54 32 3 0 -1 3 0 4 5 0 -1 2 1 2 0 1 1 0 0 1 0 0 0 2 3 0 0
 1 0 1 0 1 1 1

6CLITOCYBE PITHYOPHILA (SECRIGILL) MUSCARINE PROPOSED TREATMENT: PREVENT
 56 ABSORP, ENHANCE EXCRET, ATROPINE UNTIL CESSATION SECRETIONS, SUPPORTIVE
 TAXON MOSER (1978) KLEINE KRYPTOG. FLORA IIB, 2 FISCHER, STUTT. CHEM.
 58 CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851 MED LINCOFF &
 MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R., NY
 60 UPSETTING TO DANGEROUS EU

62 32 3 0 -1 3 0 4 5 0 -1 2 1 2 0 1 1 0 0 1 1 1 0 2 3 0 0
 1 0 1 0 1 1 1

6CLITOCYBE RIVULOSA (PEKS EX FR)KUM UK FREQ 2.4 MUSCARINE PROPOSED TREAT-
 64 MENT: PREVENT ABSORP, ENHANCE SECRET, ATROPINE UNTIL CESSATION SECRETIONS
 SUPPORTIVE TAXON KUHNER & ROMAGNESI (1977) COMPLEMENT FLORE ANALYTIQUE,
 MASSON, PARIS CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-851
 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN
 2 NOSTRAND, R., NY UPSETTING TO DANGEROUS EU

74 32 3 0 -1 3 0 4 5 1 -1 2 1 2 0 1 1 0 0 1 0 0 0 2 3 0 0
 1 0 1 0 1 1 1

Table 7 (continued)

8	6CLITOCYBE SUAVEULENS (SCHUM EX FR)KUM UK FREQ 1.2 MUSCARINE PROPOSED TREATMENT:PREVENT ABSORP,ENHANCE EXCRET,ATROPINE UNTIL CESSATION SECRETIONS,SUPPORTIVE TAXON HARMAJA H (1969)KARSTENIA 10,5-168	
9	CHEM CATALFOMO & EUGSTER (1970)HELV CHIM ACTA 53:4,848-851 MED LINCOFF & MITCHEL (1977)TOX AND HALLUC MUSHROOM POISONING,VAN NOSTRAND R. NY	
10	UPSETTING TO DANGEROUS EU	35 3 0 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 0 1 1 0 0 1 1 0 0 2
11		3 0 0 1 0 1 0 1 1 1
12	3TRICHOLOMOPSIS PLATYPHYLLA (PERS EX FR)SING UK FREQ 13.1 UNKNOWN TREATMENT SYMPTOMATIC TAXON SMITH A H (1960)BRITTONIA 12,41-76 CHEM NONE	
13	MED GOOS & SHOOP (1980)MYCOLOGIA 72,433-435 UPSETTING EU;N.AM	
14		42 6 0 -1 5 0 1 2 4 5 4 -1 2 1 2 0 1 0 0 0 1 1 2 0 6 3
15		0 0 1 0 1 -1 2 0 1 0 -1 2 1 2 2 1 1
16	5TRICHOLOMA ALBOBRUNNEUM (PERS EX FR)KUM UK FREQ 3.6 UNKNOWN,SUSPECT TREATMENT:SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMEN FLORA,IIB	
17	2,FISCHER,STUTT CHEM NONE	
18	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY UPSETTING EU;N.AM	
19		32 6 0 -1 3 0 4 5 0 -1 2 1 2 0 1 1 0 0 1 0 0 6 3 0 0
20		1 1 1 0 1 1 1
21	4TRICHOLOMA BUFONIUM (PERS EX FR)QUEL UNKNOWN TREATMENT:SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB/2,FISCHER,STUTT CHEM NONE	
22	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY UPSETTING EU;N.AM.	
23		38 6 0 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 0 1 1 0 0 1 -1 2 2 3
24		1 0 6 3 0 0 1 0 1 1 1 1
25	4TRICHOLOMA HELVIODOR PIL & SVRC UNKNOWN TREATMENT:SYMPTOMATIC TAXON PILAT & SVRCEK (1946)STUD BOT CSL 7,2-8 CHEM NONE	
26	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY.. UPSETTING EU	
27		35 6 0 -1 3 0 4 5 0 -1 2 1 2 0 1 0 0 0 1 0 -1 2 0 1 0 6
28		3 0 0 1 0 1 0 1 1 1
29	4TRICHOLOMA PARDINUM QUEL UNKNOWN TREATMENT:SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOG.FLORA IIB/2,FISCHER,STUTT. CHEM NONE	
30	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY UPSETTING TO DANGEROUS EU;N.AM	
31		34 6 0 0 -1 2 0 1 -1 2 1 2 0 1 1 0 0 1 2 -1 2 1 2 0 6 3
32		0 0 1 0 1 1 1 1 1
33	4TRICHOLOMA PESSUNDATUM (FR)QUEL UK FREQ 0.8 UNKNOWN,SUSPECT TREATMENT: SYMPTOMATIC TAXON BUN M (1967)BUL SOC MYCOL FR 83,324-335 CHEM NONE	
34	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM INTOXICATION,VAN NOSTRAND R,NY... UPSETTING EU	
35		32 6 0 -1 3 0 4 5 0 -1 2 1 2 0 1 1 0 0 1 0 0 0 6 3 0 0
36		1 1 1 0 1 1 1
37	5TRICHOLOMA SEJUNCTUM (SOW EX FR)QUEL UK FREQ 2.4 UNKNOWN SUSPECT TREATMENT:SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOG.FLORA IIB/2,FISCHER,STUTT. CHEM,NONE	
38	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY UPSETTING EU	
39		35 6 0 -1 3 0 4 5 0 -1 2 1 2 0 1 1 0 0 1 0 1 0 6 3 0 0
40		1 1 1 -1 2 0 1 1 1 1
41	5TRICHOLOMA SULPHUREUM (BUL EX FR)KUM UK FREQ 6.7 UNKNOWN TREATMENT: SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOG.FLORA IIB/2 FISCHER,STUTT CHEM NONE	
42	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R,NY UPSETTING EU	
43		38 6 0 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 0 1 1 0 0 1 -1 2 2 3
44		1 0 6 3 0 0 1 0 1 1 1 1
45	5TRICHOLOMA VIRGATUM (FR EX FR)KUM UK FREQ 2.0 UNKNOWN TREATMENT: SYMPTOMATIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB,2 FISCHER,STUTT CHEM NONE	
46	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY UPSETTING EU;N.AM	
47		40 6 0 -1 3 0 4 5 0 -1 2 1 2 0 1 1 0 0 1 1 1 0 6 3 0 0
48		1 1 1 -1 2 0 1 0 0 -1 2 1 2 1 1

Table 7 (continued)

4	SCOPELANDIA CHLOROCYSTIS WEEKS & SING PSILOCYBIN/INDOLE HALLUC. PROP- POSED TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB. DIAZEPAM, CHLORPROMAZINE TAXON/CHEM WEEKS ET AL (1979) J. NAT. PRODUCTS 42, 469-474 MED LINCOFF & MITCHEL (1977) TOX AND HALLUC MUSHROOM POISONING, VAN NOSTRAND R. NY HALLUCINOGEN, UPSETTING N. AM.	50	5	6	-1	2	0	2	2	-1	2	1	2	3	0	1	0	1	0	3	-1	2	2	3	3	3	2		
10		0	0	0	1	0	0	0	-1	2	0	1	-1	2	2	4	0	-1	2	0	1	-1	2	2	4	1			
12	4CHLOROPHYLLUM HOLYBDITES (MEYER EX FR) HASSEE UNKNOWN PROPOSED TREATMENT: SYMPTOMATIC TAXON PEGLER D (1977) PRELIM AG RIC FLORA E. AFRICA, HMSO, LOND CHEM. EILERS & NELSON (1974) TOXICON 12, 557-563 MED BLAYNEY ET AL (1980) WESTERN J. MED 132, 74-77 UPSETTING TO DANGEROUS N. AM; AFR; S. AM; C. AM.	44	6	0	-1	2	2	3	2	-1	4	0	1	2	3	0	2	0	0	1	0	-1	2	2	3	2	0		
16		6	3	0	0	1	1	1	0	0	-1	2	0	1	-1	2	0	2	1	1									
18	5CORTINARIUS AUREIFOLIUS PECK SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC TAXON AMMIRATI & GILLIAM (1975) NOVA HEDW BEIH. 51, 39-52 CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? N. AM	33	1	3	-1	2	0	4	0	-1	2	1	2	1	1	1	1	1	1	3	1	2	1	0	0	0	1	0	
24		1	1	1	1	1																							
26	4CORTINARIUS BRUNNEOFULVUS FR. S BRES SUSPECT ORELLANINE TREATMENT: SYMP- TOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER, STUTT. CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU	36	1	3	-1	2	0	4	1	-1	2	1	2	1	1	1	1	1	1	-1	2	2	3	-1	2	2	3	2	
30		1	0	0	0	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
32	4CORTINARIUS BOLARIS (PERS EX FR) FR UK FREQ 3.2 SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA, IIB/2 FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU	31	1	3	-1	3	0	4	5	1	-1	2	1	2	1	0	1	1	1	1	1	1	1	1	2	1	0	0	1
36		0	1	1	1	1	1																						
38	4CORTINARIUS CINNABARINUS FR SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU N. AM	35	1	3	-1	2	0	4	1	-1	2	1	2	1	1	1	1	1	1	1	2	1	2	1	0	0	1	0	
42		1	-1	2	0	1	0	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
44	4CORTINARIUS BULLIARDII (FR) FR SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2, FISCHER, STUTT. CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU	36	1	3	-1	2	0	4	1	-1	2	1	2	1	1	1	1	1	1	1	-1	2	2	3	1	2	1	0	0
48		0	1	0	1	-1	2	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
50	4CORTINARIUS CALLISTEUS (FR) FR UK FREQ 0.4 SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU	39	1	3	-1	3	0	4	5	0	-1	2	1	2	1	0	1	1	1	1	2	2	2	1	0	0	0	1	
54		0	1	-1	2	0	1	0	-1	2	0	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
56	4CORTINARIUS CINNAMOMEUS (L EX FR) UK FREQ 2.0 SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU N. AM	37	1	3	-1	3	0	4	5	-1	2	0	1	-1	2	1	2	1	1	1	1	0	1	1	1	1	2	1	0
60		0	0	1	0	1	-1	2	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
62	4CORTINARIUS CUTONEUS FR UK FREQ 0.4 SUSPECT ORELLANINE TREATMENT: SYMPTO- MATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU	41	1	3	-1	2	0	4	1	-1	2	1	2	1	0	1	1	1	1	1	-1	2	1	2	2	2	1	0	0
64		0	1	0	1	-1	2	0	1	0	-1	2	0	1	0	1	1	1	1	1	1	1	1	1	1	1	1		

Table 7 (continued)

4	CORTINARIUS MALICORIS FR UK FREQ 1.6 SUSPECT ORELLANINE TREATMENT:
2	SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER,
	STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934
4	MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	34 1 3 -1 3 0 4 5 0 -1 2 1 2 1 1 1 1 1 -1 2 0 1 0 2 1 0
6	0 0 1 0 1 1 1 1 1
4	CORTINARIUS PHOENICEUS (BUL) MRE SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC
8	TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2, FISCHER, STUTT,
	CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT &
10	GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	37 1 3 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 1 1 1 1 1 1 0 2 1 0
12	0 0 1 0 1 -1 2 0 1 1 1 1
4	CORTINARIUS PUNICEUS (RT) SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC
14	TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER, STUTT,
	CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-1934 MED GERAULT
15	& GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	36 1 3 -1 2 0 4 -1 2 0 1 -1 2 1 2 1 1 1 1 1 -1 2 1 2 1 2
18	1 0 0 0 1 0 1 0 1 1 1
4	CORTINARIUS RUBICUNDULUS (REA) PEARS UK FREQ 1.2 SUSPECT ORELLANINE
20	TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2
	FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34
22	MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	44 1 3 -1 3 0 4 5 1 -1 2 1 2 1 1 1 1 1 1 -1 2 0 1 2 1 0
24	0 0 1 0 1 0 0 -1 2 0 1 0 0 -1 2 0 1 0 1
4	CORTINARIUS SANGUINEUS (WULF EX FR) FR UK FREQ 0.4 SUSPECT ORELLANINE
26	TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2
	FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34
28	MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	34 1 3 -1 3 0 4 5 0 -1 2 1 2 1 1 1 1 1 -1 2 1 2 1 2 1 0
30	0 0 1 0 1 0 1 1 1
4	CORTINARIUS SEMISANGUINEUS (FR) GILL UK FREQ 4.0 SUSPECT ORELLANINE
32	TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2
	FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34
34	MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU IN AM
	31 1 3 -1 3 0 4 5 0 -1 2 1 2 1 1 1 1 1 1 1 2 1 0 0 0 1
36	0 1 0 1 1 1
4	CORTINARIUS TOPHACEUS (FR) FR SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC
38	TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2, FISCHER STUTT,
	CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34 MED GERAULT &
40	GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	39 1 3 -1 2 0 4 1 -1 2 1 2 1 1 1 1 1 -1 2 1 2 -1 2 1 2 2
42	1 0 0 0 1 0 1 -1 2 0 1 1 1 1
4	CORTINARIUS TOPHACEOIDES (MOS) SUSPECT ORELLANINE TREATMENT: SYMPTOMATIC
44	TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER, STUTT
	CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34 MED
46	GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	30 1 3 0 0 -1 2 1 2 1 0 1 1 1 1 2 2 1 0 0 0 1 0 1 -1 2
48	0 1 1 1 1
4	CORTINARIUS TRAGANUS (FR EX FR) FR UK FREQ 1.2 SUSPECT ORELLANINE TREAT-
50	MENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2
	FISCHER, STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34
52	MED GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	33 1 3 -1 3 0 4 5 0 -1 2 1 2 1 1 1 1 1 2 1 2 1 0 0 0 1
54	0 1 1 0 0 0 1 1
4	CORTINARIUS ULIGINOSUS (BERK) UK FREQ 1.6 SUSPECT ORELLANINE TREATMENT:
56	SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER
	STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34 MED
58	GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	31 1 3 -1 3 0 4 5 1 -1 2 1 2 1 1 1 1 1 2 1 2 1 0 0 0 1
60	0 1 1 1 1 1
4	CORTINARIUS SANIOSUS (FR) FR UK FREQ 0.4 SUSPECT ORELLANINE TREATMENT:
62	SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB, 2 FISCHER
	STUTT CHEM ANTKOWIAK & GESSNER (1979) TETRAHEDRON L 21, 1931-34 MED
64	GERAULT & GIRRE (1977) BUL SOC MYCOL FR 93, 373-405 DEADLY? EU
	37 1 3 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 1 1 1 1 1 2 1 2 1 0
	0 0 1 0 1 -1 2 0 1 1 1 1

Table 7 (continued)

1	6MORCHELLA ESCULENTA ST AM SUSPECT GYROMITRIN TREATMENT: PREVENT ABSORP ENHANCE EXCRET, SUPPORTIVE, PYRIDOXINE HCL, IV GLUCOSE, IF FREE HEMOGLOBIN
6	UP: UIUKESIS TAXON MOSER M (1963) KLEINE KRYPTOGAMENFLORA IIA, FISCHER, STUT
	CHEM PYYSALO H (1976) Z LEBENSM UNTERS FORTSCH 160, 325-9
9	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOST- RAND R, NY.. EU DANGEROUS RAW; EDIBLE COOKED SUSPECT?
10	32 7 1 -1 3 0 4 5 -1 2 2 5 0 0 0 1 0 0 1 5 4 0 7 0 0 4 2 0 2 1 1 1 1
12	STYLOPILUS FELLEUS (FR) KARST UK FREQ 4.7 UNKNOWN TREATMENT: SYMPTOMATIC TAXON WATLING R (1970) BRITISH FUNGUS FLORA 1, HMSO, EDINBURGH
14	CHEM NONE MED LINCOFF & MICHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU; N.AM
16	44 6 0 -1 3 0 4 5 1 -1 2 1 2 2 1 0 0 1 -1 2 3 4 1 1 6 3 0 1 1 1 1 1 0 -1 2 0 1 1 0 -1 2 0 1 1 1
18	5PAXILLUS INVOLUTUS (BATSCH EX FR) FR UK FREQ 30.9 UNKNOWN TREATMENT: SYMPTOMATIC TAXON WATLING R (1970) BRITISH FUNGUS FLORA 1, HMSO EDINBURGH
20	CHEM NONE MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. HEMOLYTIC? EDIBLE TO DANGEROUS? EU; N.AM
24	47 6 8 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 1 1 0 0 1 2 1 2 6 4 3 1 1 0 1 -1 2 0 1 0 -1 2 1 2 1 0 -1 2 1 2 1 1
26	SHYGROPHOROPSIS AURANTIACA (FR) HRE UK FREQ 13.9 SUSPECT MUSCARINE? EDIBLE TREATMENT: SYMPTOMATIC TAXON MOSER M (1978) KLEINE KRYPTOGAMENFLORA IIB/2 FISCHER, STUT. CHEM CATALFOMO & EUGSTER (1970) HELV CHIM ACTA 53:4, 848-51
28	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU; N.AM; AF
30	42 3 0 -1 4 0 1 4 5 -1 2 0 1 -1 2 1 2 0 2 1 0 0 1 -1 2 0 1 -1 2 0 1 0 2 3 0 1 1 0 1 0 1 1 1
32	4LACTARIUS TORMINOSUS (SCHAEFF EX FR) GRAY UK FREQ 7.5 UNKNOWN TREATMENT: SYMPTOMATIC TAXON PEARSON (1950) NATURALIST JULY-SEP, 81-100 CHEM NONE
34	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU; N.AM
36	35 6 0 -1 3 0 4 5 1 -1 2 1 2 0 0 1 2 0 1 2 2 0 6 3 1 3 1 1 1 -1 2 0 1 1 1 1
38	4LACTARIUS CHRYSORRHEUS FR UK FREQ 4.3 UNKNOWN TREATMENT: SYMPTOMATIC TAXON PEARSON (1950) NATURALIST, JULY-SEP, 81-100 CHEM NONE
40	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU; N.AM
42	35 6 0 -1 3 0 4 5 1 -1 2 1 2 0 0 0 2 0 1 1 2 0 6 3 1 3 1 1 1 -1 2 0 1 1 1 1
44	4LACTARIUS GLAUDESCENS CROSS UK FREQ 0.4 UNKNOWN TREATMENT: SYMPTOMATIC TAXON PEARSON (1950) NATURALIST, JULY-SEP, 81-100 CHEM NONE
46	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU
48	38 6 0 -1 3 0 4 5 1 -1 2 1 2 0 0 0 2 0 1 1 -1 2 1 2 0 6 3 1 3 1 1 1 -1 2 0 1 1 1 1
50	4LACTARIUS HELVUS FR UK FREQ 2.4 UNKNOWN TREATMENT: SYMPTOMATIC TAXON PEARSON (1950) NATURALIST, JULY-SEP 81-100 CHEM NONE
52	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU; N.AM
54	38 6 0 -1 3 0 4 5 0 -1 2 1 2 0 0 1 2 0 1 2 -1 2 1 2 0 6 3 1 3 1 1 1 -1 2 0 1 1 1 1
56	4LACTARIUS REPRESENTANAEOUS BRITZ UK FREQ 0.4 UNKNOWN TREATMENT: SYMPTOMATIC TAXON PEARSON (1950) NATURALIST, JULY-SEP, 81-100 CHEM NONE
58	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU; N.AM
60	35 6 0 -1 3 0 4 5 0 -1 2 1 2 0 0 0 2 0 1 -1 2 2 3 3 0 6 3 1 3 1 1 1 1 1 1 1
62	4LACTARIUS RUFUS (SCOPI) FR UK FREQ 22.6 UNKNOWN TREATMENT: SYMPTOMATIC TAXON PEARSON (1950) NATURALIST, JULY-SEP, 81-100 CHEM NONE
64	MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY.. UPSETTING EU; N.AM
2	35 6 0 -1 3 0 4 5 0 -1 2 1 2 0 0 1 2 0 1 2 2 0 6 3 1 3 1 1 1 -1 2 0 1 1 1 1

Table 7 (continued)

4	4LACTARIUS SCROBICULATUS (SCOPI)FR UNKNOWN TREATMENT:SYMPTOMATIC
5	TAXON PEARSON (1950)NATURALIST,JULY-SEP,81-100 CHEM NONE
6	MED LINGOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOST-
7	RAND R,NY... UPSETTING EU;N.AM
8	35 6 0 -1 3 0 4 5 0 -1 2 1 2 0 0 1 2 0 1 2 2 0 6 3 1 3
9	1 1 1 -1 2 0 1 1 1 1
10	4LACTARIUS UVIDUS FR UK FREQ 3.2 UNKNOWN TREATMENT:SYMPTOMATIC
11	TAXON PEARSON (1950)NATURALIST,JULY-SEP,81-100 CHEM NONE
12	MED LINGOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN
13	NOSTRAND R,NY... UPSETTING EU,N.AM
14	32 6 0 -1 3 0 4 5 1 -1 2 1 2 0 0 1 2 0 1 3 3 0 6 3 1 3
15	1 1 1 1 1 1 1
16	SRUSSULA EMETICA FR UK FREQ 14.3 UNKNOWN TREATMENT:SYMPTOMATIC
17	TAXON KAYNER RW (1974)BRITISH SPP RUSSULA,B.MYCOL SOC,CAMBRIDGE
18	CHEM NONE
19	MED LINGOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN
20	NOSTRAND R,NY... UPSETTING EU;N.AM
21	40 6 0 -1 5 0 2 3 4 5 0 -1 2 1 2 0 0 1 2 0 1 -1 2 2 3 -1
22	2 2 3 0 6 3 1 3 1 1 1 1 1 1
23	SRUSSULA FOETENS FR UK FREQ 19.0 SUSPECT UNKNOWN TREATMENT:SYMPTOMATIC
24	TAXON RAYNER RW (1974)BRITISH SPP RUSSULA,BR.MYCOL SOC,CAMBRIDGE
25	CHEM NONE
26	MED LINGOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN
27	NOSTRAND,R,NY... UPSETTING EU;N.AM
28	38 6 0 -1 3 0 4 5 -1 2 0 1 -1 2 1 2 0 0 0 1 0 1 2 -1 2 2
29	3 0 6 3 1 3 1 1 1 1 1 1 1
30	SRUSSULA FRAGILIS (PERS EX FR)FR UK FREQ 15.0 SUSPECT UNKNOWN
31	TREATMENT:SYMPTOMATIC TAXON RAYNER RW (1974)BRITISH SPP RUSSULA,BR.MYCOL
32	SOC,CAMBRIDGE CHEM NONE
33	MED LINGOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN
34	NOSTRAND R,NY... UPSETTING EU
35	37 6 0 -1 5 0 2 3 4 5 -1 2 0 1 -1 2 1 2 0 0 1 2 0 1 2 2
36	0 6 3 1 3 1 1 1 1 1 1 1
37	SRUSSULA HAIREI SING UK FREQ 19.8 SUSPECT UNKNOWN TREATMENT:SYMPTOMATIC
38	TAXON RAYNER RW (1974)BRITISH SPP RUSSULA,B MYCOL SOC,CAMBRIDGE
39	CHEM NONE
40	MED LINGOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN
41	NOSTRAND R,NY... UPSETTING EU
42	35 6 0 -1 3 0 4 5 1 -1 2 1 2 0 0 0 2 0 1 1 2 0 6 3 1 3
43	1 1 1 -1 2 0 1 1 1 1
44	SRUSSULA SARDONIA FR UK FREQ 5.9 SUSPECT UNKNOWN TREATMENT:SYMPTOMATIC
45	TAXON RAYNER RW (1974)BRITISH SPP RUSSULA,B MYCOL SOC,CAMBRIDGE
46	CHEM NONE
47	MED LINGOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN
48	NOSTRAND R,NY... UPSETTING EU
49	38 6 0 -1 3 0 4 5 0 -1 2 1 2 0 0 0 2 0 1 -1 2 1 2 2 0 6
50	3 1 3 1 1 1 -1 2 0 1 1 1 1
51	SLEPIOTA BRUNNEOINCARNATA CHOD & MARTIN CYCLOPEPTIDES PROPOSED TREATMENT:
52	WATER,ELECTROLYTES,BLOOD BALANCE,HYPERTONIC GLUCOSE INF,PENICILLIN,
53	THIOCTIC ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)KLEINE KRYPTO-
54	GAMENFLORA IIB/2 FISCHER,STUTT CHEM & MED BERTELLI ,FOURNIER ET AL
55	(1977)CURRENT PROBLEMS IN CLIN BIOCHEM 7,HUBER STUTT... DEADLY EU
56	41 0 2 -1 3 0 2 4 -1 2 1 2 -1 1 1 2 0 2 1 0 0 1 -1 2 1 1
57	2 1 0 0 0 0 0 0 0 1 0 0 0 0 1 1
58	SLEPIOTA CASTANEA QUEL UK FREQ 1.6 CYCLOPEPTIDES PROPOSED TREATMENT:WATER
59	ELECTROLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INF,PENICILLIN,THIOCTIC
60	ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA
61	IIB,2 FISCHER,STUTT CHEM & MED BERTELLI ET AL (1977)CURRENT PROBLEMS
62	IN CLIN BIOCHEM 7,HUBER,STUTT.. DEADLY EU
63	43 0 2 -1 3 0 2 4 -1 2 0 1 -1 2 1 2 0 2 1 0 0 1 -1 2 2 3
64	-1 2 0 1 0 0 0 0 0 1 0 1 1 0 0 0 1 1
65	SLEPIOTA GLYPEOLARIOIDES REA UK FREQ 0.4 CYCLOPEPTIDES PROPOSED TREATMENT
66	WATER,ELECTROLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INF,PENICILLIN
67	THIOCTIC ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)KLEIN KRYPTO-
68	AMENFLORA,IIB/2,FISCHER,STUTT. CHEM & MED BERTELLI ET AL (1977)CURRENT
69	PROBLEMS IN CLIN BIOCHEM 7,HUBER STUTT.. DEADLY EU
70	34 0 2 -1 3 0 2 4 0 -1 2 1 2 0 2 1 0 0 1 1 1 0 0 0 0 0
71	0 0 1 0 0 0 0 1 1

Table 7 (continued)

5	SLEPIOTA CRISTATA (BOLT EX FR)KUM UK FREQ 10.7 SUSPECT CYCLOPEPTIDES																									
6	PROPOSED TREATMENT:WATER,ELECTROLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE																									
7	INF,PENICILLIN,THIOCTIC ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)																									
8	KLEINE KRYPTOGAMENFLORA IIB/2,FISCHER,STUTT CHEM & MED BERTELLI ET AL																									
9	(1977)CURRENT PROBLEMS IN CLIN BIOCHEM 7,HUBER,STUTT DEADLY? EU;N.AM																									
10	36	0	2	-1	5	0	2	3	4	5	2	-1	2	1	2	0	2	1	0	0	1	1	0	0	0	0
11		J	J	J	0	0	0	0	0	1	1															
12	SLEPIOTA FELINA (PERS EX FR)KARST UK FREQ 2.0 CYCLOPEPTIDES PROPOSED																									
13	TREATMENT:WATER,ELECTROLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INFUSION																									
14	PENICILLIN,THIOCTIC ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)KLEINE																									
15	KRYPTOGAMENFLORA IIB/2,FISCHER,STUTT..CHEM & MED BERTELLI ET AL (1977)																									
16	CURRENT PROBLEMS IN CLIN BIOCHEM 7,HUBER,STUTT.. DEADLY EU																									
17	34	0	2	-1	3	0	2	4	0	-1	2	1	2	0	2	1	0	0	1	1	0	0	0	0	0	0
18		0	0	1	0	0	0	0	1	1																
19	SLEPIOTA FUSCOVINALEA MOLL & LGE UK FREQ 0.4 CYCLOPEPTIDES PROPOSED																									
20	TREATMENT:WATER,ELECTROLYTE,BLOOD BALANCE,HYPERTONIC GLUCOSE INFUSION																									
21	PENICILLIN,THIOCTIC ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)																									
22	KLEINE KRYPTOGAMENFLORA IIB/2,FISCHER,STUTT CHEM & MED BERTELLI ET AL																									
23	(1977)CURRENT PROBLEMS IN CLIN BIOCHEM 7,HUBER,STUTT.. DEADLY EU																									
24	37	0	2	-1	3	0	2	4	-1	2	1	2	-1	2	1	2	0	2	1	0	0	1	0	0	0	0
25		0	0	0	1	1	J	0	0	1	1															
26	SLEPIOTA GRISEOVIRENS MRE CYCLOPEPTIDES PROPOSED TREATMENT:WATER,																									
27	ELECTROLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INF,PENICILLIN,THIOCTIC																									
28	ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA																									
29	IIB/2,FISCHER,STUTT CHEM & MED BERTELLI ET AL (1977)CURRENT PROBLEMS																									
30	IN CLIN BIOCHEM 7,HUBER,STUTT... DEADLY EU																									
31	37	0	2	-1	3	0	2	4	-1	2	0	1	-1	2	1	2	0	2	1	0	0	1	1	0	0	0
32		J	0	0	0	0	1	0	0	0	1	1														
33	SLEPIOTA HETIERI BOUD UK FREQ 0.8 CYCLOPEPTIDES PROPOSED TREATMENT:WATER																									
34	ELECTROLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INF,PENICILLIN,THIOCTIC																									
35	ACID,EXCHANGE TRANSFUSION TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA																									
36	IIB/2,STUTT FISCHER CHEM & MED BERTELLI ET AL (1977)CURRENT PROBLEMS IN																									
37	CLIN BIOCHEM 7,HUBER STUTT... DEADLY EU																									
38	40	0	2	-1	3	0	2	4	-1	2	0	1	-1	2	1	2	0	1	1	0	0	1	0	0	0	0
39		0	0	0	0	1	0	0	0	-1	2	1	3	1	1											
40	SLEPIOTA OCHRACEOFULVA ORT CYCLOPEPTIDES PROPOSED TREATMENT:WATER,ELEC-																									
41	TROYLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INF.,PENICILLIN,THIOCTIC ACID																									
42	EXCHANGE TRANSFUSION TAXON MOSEP M (1978)KLEINE KRYPTOGAMENFLORA IIB/2																									
43	FISCHER,STUTT. CHEM & MED BERTELLI ET AL (1977)CURRENT PROBLEMS IN																									
44	CLIN BIOCHEM 7,HUBER,STUTT... DEADLY EU																									
45	43	0	2	-1	3	0	2	4	-1	2	0	1	-1	2	1	2	0	2	1	0	0	1	-1	2	1	2
46		-1	2	0	1	0	0	0	0	0	0	1	0	0	0	0	1	1								
47	SLEPIOTA PSEUDOFELINA LGE CYCLOPEPTIDES PROPOSED TREATMENT:WATER,ELEC-																									
48	TROYLYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INF,PENICILLIN,THIOCTIC ACID																									
49	EXCHANGE TRANSFUSION MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB/2																									
50	FISCHER,STUTT CHEM & MED BERTELLI ET AL (1977)CURRENT PROBLEMS IN CLIN																									
51	BIOCHEM 7,HUBER,STUTT.. DEADLY, EU																									
52	38	0	2	-1	4	0	2	4	5	-1	2	0	1	-1	2	1	2	0	2	1	0	0	1	2	0	0
53		0	0	0	0	0	1	0	0	0	1	1														
54	SLEPIOTA SUBINCARNATA LGE CYCLOPEPTIDES PROPOSED TREATMENT:WATER,ELEVTR-																									
55	LYTE & BLOOD BALANCE,HYPERTONIC GLUCOSE INF,PENICILLIN,THIOCTIC ACID,																									
56	EXCHANGE TRANSFUSION TAXON PEGLER (1977)PRELIM AGARIC FLORA E. AFRICA																									
57	HMSO,LON CHEM & MED BERTELLI ET AL (1977)CURRENT CONTENT IN CLIN BIOCHEM																									
58	7,HUBER,STUTT... DEADLY EU;AF																									
59	44	0	2	-1	4	0	2	3	4	-1	2	0	1	-1	2	1	2	0	2	1	0	0	1	-1	2	0
60		1	-1	2	0	1	0	0	0	0	0	0	1	0	0	0	0	1	1							
61	GLEUCOCOPRINUS BADHAMII JK & BR UK FREQ 0.4 (GLASSHOUSE) CYCLOPEPTIDE																									
62	SUSPECT PROPOSED TREATMENT:WATER,ELECTROLYTE & BLOOD BALANCE,HYPERTONIC																									
63	GLUCOSE INF,PENICILLIN,THIOCTIC ACID,EXCHANGE TRANSFUSION TAXON MOSER																									
64	M (1978)KLEINE KRYPTOGAMENFLORA IIB/2,FISCHER,STUTT CHEM & MED BERTELLI																									
65	ET AL (1977)CURRENT PROBLEMS IN CLIN BIOCHEM 7,HUBER,STUTT...DEADLY																									
66	COSMOPOLITAN																									
67	49	0	2	-1	6	0	1	2	3	4	5	1	-1	2	1	2	0	1	1	0	1	0	-1	2	0	1
68		-1	2	0	1	J	0	0	0	0	1	1	0	0	-1	2	0	1	-1	2	0	2	1	1		

Table 7 (continued)

1	SNOLANEA SERICEA (BUL EX MER)ORT UK FREU 6.7 UNKNOWN TREATMENT:SYMPTOMA-
2	TIC TAXON MOSER M (1978)KLEINE KRYPTOGAMENFLORA IIB/2,FISCHER,STUTT.
3	CHEM NONE
4	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOST-
5	RAND,NY... UPSETTING EU
6	3 6 0 -1 3 0 4 5 2 -1 2 1 2 2 2 0 0 1 -1 2 1 2 2 1 6 3
7	0 0 1 0 1 0 1 1 1
8	6COPELANDIA MEXICANA GUZ PSILOCYBIN/INDOLE HALLUC TREATMENT:PREVENT ABSOR
9	ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET, POSSIB DIAZEPAM,
10	CHLORPROMAZINE TAXON GUZMAN G (1978)BOL SOC MEX MIC 12,27-31 CHEM
11	HOFMANN ET AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL
12	(1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY..
13	HALLUCINOGEN,UPSETTING G.AM
14	46 5 0 2 -1 2 2 3 2 3 0 1 0 1 0 -1 2 1 2 2 3 3 2 0 0 0
15	0 1 -1 2 0 1 0 0 -1 3 1 2 4 0 1 -1 3 1 2 4 1
16	6PSILOCYBE UXPANAPENSIS GUZ PSILOCYBIN/INDOLE HALLUC TREATMENT:PREVENT
17	ABSORP,ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIBLY DIAZE-
18	PAM,CHLORPROMAZINE TAXON GUZMAN G(1979)BEIH SYDOWIA 8,168-181 CHEM
19	HOFMANN ET AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL
20	(1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY..
21	HALLUCINOGEN,UPSETTING MEX
22	42 5 0 3 1 1 3 0 1 0 1 0 -1 2 0 1 -1 2 0 1 3 3 2 0 0 1
23	0 1 0 0 0 -1 3 1 2 3 0 0 -1 2 1 2 0
24	6PSILOCYBE NAEMATOLIFORMIS GUZ PSILOCYBIN/INDOLE HALLUC SUSPECT TREATMENT
25	PREVENT ABSORP,ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB
26	DIAZEPAM,CHLORPROMAZINE TAXON GUZMAN G (1979)BEIH SYDOWIA 8,168-181
27	CHEM HOFMANN ET AL (1959)HELV CHIM ACTA 52,1557-1572 MED LINCOFF &
28	MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY..
29	HALLUCINOGEN UPSETTING MEX
30	40 5 0 3 1 1 3 0 1 0 1 0 0 -1 2 0 1 3 3 2 0 0 1 0 1 0
31	0 0 -1 3 1 2 4 0 0 -1 3 1 2 4 0
32	6PSILOCYBE SINGERI GUZ SUSPECT PSILOCYBIN/INDOLE HALLUC TREATMENT:PREV-
33	ENT ABSORP,ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB
34	DIAZEPAM,CHLORPROMAZINE TAXON GUZMAN G (1979)BEIH SYDOWIA 8,168-181
35	CHEM HOFMANN ET AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF &
36	MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY...
37	HALLUCINOGEN,UPSETTING, MEX
38	36 5 0 3 1 1 3 0 1 0 1 0 0 -1 2 0 1 3 3 2 0 0 1 0 1 0
39	0 0 1 0 0 -1 3 1 2 4 1
40	6PSILOCYBE WELDENII GUZ SUSPECT PSILOCYBIN/INDOLE HALLUC TREATMENT:PREVEN
41	ABSORP,ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB DIAZEPAM
42	CHLORPROMAZINE TAXON GUZMAN G (1979)BEIH SYDOWIA 8,168-181 CHEM HOFMANN
43	ET AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977)
44	TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY.. HALLUCINOGEN,
45	UPSETTING MEX
46	41 5 0 3 1 1 3 0 1 0 1 0 -1 2 0 1 -1 2 0 1 3 3 2 0 0 1
47	0 1 0 0 0 -1 2 1 2 0 0 -1 2 1 2 0
48	6PSILOCYBE NEOCALEDONICA GUZ & HOR SUSPECT INDOLE HALLUC TREATMENT:PREVEN
49	ABSORP,ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSSIB DIAZE-
50	PAM,CHLORPROMAZINE TAXON GUZMAN & HORAK (1978)BEIH SYDOWIA 31,44-54
51	CHEM HOFMANN ET AL (1959)HELV CHIM ACTA 42,1557-1572 MED LINCOFF &
52	MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOSTRAND R,NY..
53	HALLUCINOGEN,UPSETTING? NEW CALEDONIA
54	38 5 0 3 -1 2 1 4 3 3 0 1 0 1 0 0 1 3 3 2 0 0 1 0 1 0
55	0 0 -1 2 1 2 0 0 -1 2 1 2 0
56	6PSILOCYBE BRUNNEOCYSTIDIATA GUZ & HOR SUSPECT INDOLE HALLUC TREATMENT:
57	PREVENT ABSORP,ENHANCE EXCRET,RESTRAIN SELF DESTRUCT BEHAV,QUIET,POSS-
58	IB DIAZEPAM,CHLORPROMAZINE TAXON GUZMAN & HORAK (1978)BEIH SYDOWIA 31,
59	44-54 CHEM HOFMANN ET AL (1959)HELV CHIM ACTA 42,1552-1572
60	MED LINCOFF & MITCHEL (1977)TOX & HALLUC MUSHROOM POISONING,VAN NOST-
61	RAND,R,NY.. HALLUCINOGEN,UPSETTING? NW GUINEA
62	45 5 0 3 -1 2 1 4 -1 3 0 2 3 3 0 1 0 1 0 1 -1 2 0 1 3 3
63	2 0 0 1 0 1 0 0 -1 5 0 1 2 3 4 0 0 2 1

Table 7 (continued)

6PSILOCYBE NOTHOAGENSIS GUZ & HOR SUSPECT INDOLE HALLUC TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB DIAZEPAM, CHLORPROMAZINE TAXON GUZMAN & HORAK (1978) BEIH SYDOWIA 31,44-54 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTR. R, NY... HALLUCINOGEN, UPSETTING? NW GUINEA
 30 5 0 3 -1 2 1 4 2 3 0 1 0 1 0 0 8 3 3 2 0 0 1 0 1 0
 0 0 2 1 1

6PSILOCYBE PAPUANA GUZ & HOR SUSPECT INDOLE HALLUC TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB DIAZEPAM, CHLORPROMAZINE TAXON GUZMAN & HORAK (1978) BEIH SYDOWIA 31,44-54 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND, R, NY... HALLUCINOGEN, UPSETTING? NW GUINEA
 39 5 0 3 1 -1 2 2 3 3 0 1 0 1 0 -1 2 0 1 -1 2 0 1 3 3 2
 0 0 1 0 1 0 0 0 -1 2 1 2 1 1

6PSILOCYBE INCONSPICUA GUZ & HOR INDOLE HALLUC SUSPECT TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB DIAZEPAM, CHLORPROMAZINE TAXON GUZMAN & HORAK (1978) BEIH SYDOWIA 31,44-54 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND, R, NY... UPSETTING, HALLUCINOGEN? NEW GUINEA
 33 5 0 3 1 3 3 0 1 0 1 0 -1 2 0 1 0 3 3 2 0 0 1 0 1 0
 0 0 -1 2 1 2 1 1

6PSILOCYBE NOVAZELANDIAE GUZ & HOR SUSPECT HALLUC. INDOLE TREATMENT: PREVENT ABSORP, ENHANCE EXCRET, RESTRAIN SELF DESTRUCT BEHAV, QUIET, POSSIB DIAZEPAM, CHLORPROMAZINE TAXON GUZMAN & HORAK (1978) BEIH SYDOWIA 31,44-54 CHEM HOFMANN ET AL (1959) HELV CHIM ACTA 42,1557-1572 MED LINCOFF & MITCHEL (1977) TOX & HALLUC MUSHROOM POISONING, VAN NOSTRAND R, NY... UPSETTING, HALLUCINOGEN? NW ZEALAND
 34 5 0 1 1 2 3 0 1 0 1 0 3 -1 2 0 1 3 3 2 0 0 1 0 1 0
 0 0 -1 3 1 2 4 1 1

Table 7 (end).

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CHAPTER V. : CONCLUSION.

The first and almost unique question which always arises in case of toxicological emergency is "What did cause the intoxication?". A question which, despite two centuries of mycotoxicology and specialised training in some countries, still cannot be satisfactorily answered in a great majority of cases of mushroom poisoning.

The task, in itself, seems straightforward and strikingly simple: poisonous species have to be recorded and permanent or stable characteristic features selected and used in a manner which allows a rapid identification. It needs a multidisciplinary approach as well as an understanding of the practical requirements for an efficient management of mushroom poisoning.

The almost unlimited storage facilities of computers indicated that they are the ideal tool for storing and retrieving data. The completely interactive facilities that they offer are the most sophisticated type of identification methods yet available because they can be used while the identification of a specimen is in progress. Any classical identification or diagnostic key can be programmed into modern computers which can suit the most stringent requirements, within the limits of logic, and

provide a quick and reliable answer. Full advantage of these facilities have been taken to suit a number of obligatory (e.g. multi-access facility, because of the fragmentary nature of the samples usually submitted for identification) as well as desirable requirements (e.g. the use of English in a form of dialogue with the machine), to make the key efficient, useful, and, at the same time, easy to operate by even untrained personnel.

The data matrix is made up of 223 species of mushrooms selected from a world list of 432 suspected or poisonous species, because 1) they are known to be poisonous, 2) they are British species suspected to be poisonous or 3) they are non British species suspected to be poisonous and a complete set of identification features were available from literature sources.

The characters used for identification have been selected because they are, preferably, easy to observe and, at the same time, provide potentially useful information in the identification process. A total of twenty four characters have been selected, showing a choice of 127 possible states or options. Only a limited number of characters may be used for most

practical purposes and still provide a useful result.

Each taxon is defined by a combination of the 127 possible states described in this thesis and the data is further complemented by nine features which are retrieved as essential information to users, when an identification is obtained. Therefore, each taxon is described by a set of 136 combined data and the total information stored in the data bank is of 30'328 data which can be easily altered or extended at any time.

These data have been checked, complemented or expanded by microscopical and chemical analysis while the data bank was built. Significant results were obtained with the analysis of a number of species of the Strophariaceae and Coprinaceae families. The presence of controlled hallucinogenic drugs (psilocybin and derivatives) was confirmed in Scottish samples of Psilocybe semilanceata and P. cyanescens. It was also confirmed in Australian specimens of P. cubensis and identified for the first time in P. eucalypta.

Samples of Stropharia , Hypoholoma and Panaeolus did not contain any of the known mushroom hallucinogens, except for one sample of Panaeolus subbalteatus submitted for analysis after an intoxication occurred in Edinburgh.

Other results were obtained in this laboratory by Greer (1980) on species of Cortinarius analysed for the presence of orellanine. These results urgently warrant further research. The deadly, still uncharacterised, toxin orellanine was tentatively identified in Scottish specimens of Cortinarius speciosissimus, and in lesser concentration in the more common C. cinnamomeus, C. sanguineus and C. semisanguineus.

At the time of writing this thesis, the key is as complete as is necessary and practical. It should provide an important tool for toxicologists dealing with emergency cases and forensic scientists employed in the identification of drugs with abuse potential. This is still a long way from providing all the answers, and a lot more work is necessary if solutions to the questions of the pharmacokinetic, the analysis in body fluids, etc of the different toxins, are required. Throughout the thesis, suggestions are made where further research is needed and selected questions are raised and should be the catalyst to further fruitful research.

The programme has some shortcomings and users are advised against its indiscriminate use. It is primarily designed to identify a limited number of mushroom species

(up to about 300) which are poisonous or suspect, out of the 10'000 or so described in the mycological literature. For each poisonous species, there may be as many as 10 non poisonous closely related species showing the same characters and options selected here. Therefore, only if any evidence of toxicity is present can the identification be accurate. A mushroom sample collected in the field identified solely on the basis of its microscopical and ecological characters may be poisonous. The key indicates only which poisonous species it can be!

All poisonous mushrooms are not yet known, and definite cases of poisoning may occur where no identification is obtained. When such a case arises, samples should immediately be forwarded to specialists and the new information thus made available used to augment the data stored.

Finally, the key is not intended to replace the specialist, but it is an attempt to provide selected specialist's information to the practising scientist. It has been tested on a number of occasions with a data bank of about 100 species. Selected notoriously difficult species (in the genus Inocybe) were readily and correctly identified in the presence of an expert

mycologist, Dr. R. Watling from the Royal Botanic Garden in Edinburgh.

This programmed key is presented as a cornerstone to the management and diagnosis of mushroom poisoning based on interdisciplinary sources ranging from anamnesic, clinical, ecological, microscopical to chemical information.



Cortinarius cinnamomeus collected

by the author

Original watercolour

by R. Furlong.

CHAPTER VI : APPENDIX.

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VI. APPENDIX.

In the investigation of microscopical characters (2.2.2.1.1.) a number of reagents are used because they give useful information, are easy to prepare and are readily available to most scientists. The composition of such reagents for investigating microscopical features for inclusion into the data bank are described.

Analysis for the different mushroom toxins is outlined.

6.1. Reagents.

6.1.1. Melzer's Reagent.

Mycologists have found in Melzer's reagent a convenient standard solution for the study of the Agaricales. Used in a standard fashion, it always produces identical results, no matter the age or the method of drying of the sample.

Composition:	KI	1.5 g	
	I ₂	0.5 g	
	H ₂ O	20.0 g	
	Cl ₃ CCH(OH) ₂	22.0 g	(chloral hydrate)

The microscopical preparation must first be wetted in conc. ammonia for a few seconds, the ammonia is then removed before applying an excess of Melzer's solution. An amyloid reaction is a grayish blue to black colouration of the element observed (here: clear spores under the microscope 2.2.2.1.1.9.) and a dextrinoid reaction is expressed by a brown to purple brown colouration. (Singer, 1975).

6.1.1. Various Reagents.

All the microscopical preparations are made in a 3 % (v/v) aqueous solution of ammonia, unless a specific reagent is mentioned.

Acetocarmine: boil 45 % (v/v) acetic acid with excess carmine for $\frac{1}{2}$ hour, filter and dilute with 45 % (v/v) ethanol. Add two drops of ferric hydroxide to every 50 mls of solution (germ pore 2.2.2.1.1.14.).

Ammonia: 75 % (v/v) aqueous solution (chrysocystidia 2.2.2.1.1.18.4.)

Cresyl blue : dilute aqueous solution (approx. 3-5 %; germ pore, 2.2.2.1.1.14. and clamp connection, 2.2.2.1.1.15.).

Sulphuric acid : 50 % (v/v) or concentrated (colour resistance of dark spores, 2.2.2.1.1.9.)

Reference for further useful reagents: Henderson et al., 1969.

6.2. Analyses of the Toxins.

6.2.1. Cyclopeptides.

In fatal cases of Amanita phalloides intoxication, the amatoxins (Figure 18, pp.107) are the sole cause of death. The phallotoxins do not contribute to human poisoning. The most commonly used method for the detection of amatoxins is their reaction with cinnamic aldehyde (1 ml in 100 ml of methanol) and hydrochloric acid fumes. A rapid and sensitive high performance thin layer chromatographic method has been developed by Stijve and Seeger (1979) which enabled them to determine α -, β - and γ -amanitin in crude methanolic extracts of mushrooms. The detection limits are 50 ng for all the amanitins.

This is favoured over adsorption chromatography on Sephadex LH 20 followed by thin layer chromatography or amino-acid analysis as reported by Faulstich et al. (1974),

inhibition of RNA polymerase as advocated by Cochet-Meilhac and Chambon (1974) or radioimmunoassay (Faulstich, 1979).

A spot test for amatoxins has been developed by Wieland (Faulstich, 1979). Crushed mushroom juice containing amatoxins developed with 8N hydrochloric acid on newspaper gives a prussian blue colour. This assay may be used to detect amanitins in all deadly poisonous mushrooms.

6.2.2. Orellanine.

Orellanine is efficiently extracted into methanol after a first treatment with petroleum ether, "defatting" the mushroom. Good resolution of a number of yet unidentified fluorescent compounds is obtained, using thin layer chromatography on silica gel plates and a variety of eluents. A cyclo-hexane : ethyl acetate (3 : 1 v/v) solvent was found to give the best resolution by Greer (1980). No reagent give a sensitive reaction to orellanine and consequently ultra-violet fluorescence is recommended. Standard extracts of Cortinarius orellanus or C. speciosissimus may be used as reference until the structure of orellanine is characterised.

6.2.3. Muscarine.

Isolation of muscarine involves a large number of operations and all chromatographic estimations have to be carried out on partially enriched material isolated as a mixture of Reineckate salts of a group of quaternary ammonium compounds.

Good qualitative and quantitative results were obtained by Cunningham (1975) using derivatisation before gas chromatographic analysis. Defatted ethanolic extracts of mushrooms are purified on an alumina column eluted with methanol. Muscarine is demethylated with sodium benzene thiolate to its volatile nor-base, nor-muscarine. Nor-muscarine is separated from the reaction mixture with chloroform. Concentrated extracts are injected onto a 1 % Carbowax 20M gas chromatographic column at 130°C and detection is obtained by flame ionisation. Dimethylaminoethyl benzoate is recommended as an internal standard.

6.2.4. Hallucinogens : Psilocybin and Analogues.

Defatted methanolic extracts of mushrooms give a fairly pure mixture of the indole hallucinogens psilocybin and baeocystin. A lengthy soxhlet extraction is proposed

in the literature, but it was found that 3 methanolic extractions gave just as good a result. No psilocybin could be detected in the third extract.

Several techniques have been used to analyse for psilocybin, psilocin and baeocystin in extracts of mushrooms.

Thin layer chromatography is a powerful analytical tool, best separation being obtained on silica gel G employing a solvent of butanol : acetic acid : water (2 : 1 : 1 v/v). Psilocin (R_f 0.52; 10 ng) and psilocybin (R_f 0.33; 15 ng) can be detected using acidic para-dimethylaminobenzaldehyde (p-DAB) as the chromogenic reagent.

UV spectrophotometry: psilocin has an $A_1^1 = 297$ in methanol at $\lambda = 267$ nm and psilocybin an $A_1^1 = 206$ in methanol at $\lambda = 268$ nm

IR spectrophotometry: peaks are listed in order of decreasing intensity. Psilocin: 817; 1340; 1470; 1258 and 1041 and 3250; 1233 cm^{-1} . Psilocybin : 920; 1100; 1040; 1347; 1060; 748 and 960 and 1178 cm^{-1} .

High performance liquid chromatography: Perkal et al. (1980) recommend a system based on an ion exchange column of Whatman Partisil SCX 10, using a mobile phase

of methanol : water (20 : 80 v/v) containing 0.2 % of ammonium phosphate and 0.1 % KCl. Detection is by UV and fluorescence.

Psilocin and psilocybin have been analysed on a 1.5 % SE-30 (chromosorb W) gas chromatographic column by Repke et al. (1977).

6.2.5. Isoxazole Derivatives.

Ibotenic acid and muscimol are extracted from mushrooms with 50 to 75 % (v/v) ethanol and separated by thin layer chromatography using butanol : acetic acid : water (12 : 3 : 5 v/v) as solvent and ninhydrin as the chromophoric reagent.

Electrophoresis has also been proposed by Chilton and Ott (1976).

6.2.6. Coprine.

Coprine will extract into water but no rapid screening method has been described in the literature. The method of isolation described (Hatfield & Schaumberg, 1975) requires that the water extract is fractionated using anion exchange, silica-gel dry-column and Sephadex G-10 gel filtration chromatography. The compound reacts with ninhydrin.

6.2.7. Gyromitrin.

Gyromitrin and its analogues are volatile toxins which are readily distilled and analysed by gas liquid chromatography. Pyysalo (1975) proposes the use of FFAP capillary columns and mass spectrometric detection.

The Ehrlich reagent, dimethylaminobenzaldehyde, has been used for detection of gyromitrin on chromatograms. (List & Luft, 1968).

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