



**PHARMACOLOGICAL CHARACTERIZATION OF  
IMPROVED LIGANDS AND PROSTANOID  
RECEPTORS IN ISOLATED TISSUE PREPARATIONS**

A thesis presented by

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## RESEARCH OUTPUTS

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# TABLE OF CONTENTS

	Pages
ACKNOWLEDGEMENTS	I
RESEARCH OUTPUTS	II
TABLE OF CONTENTS	III
ABBREVIATIONS	IX
ABSTRACT	XII
CHAPTER ONE: GENERAL INTRODUCTION	<b>1</b>
1.1 Prostaglandin biosynthesis	3
1.2 Molecular biology	6
1.2.1 Isoforms	6
1.2.2 Signal transduction	7
1.3 Prostanoid receptors classification and distribution	9
1.3.1 DP <sub>1</sub> receptor	10
1.3.2 EP receptor	10
1.3.2.1 EP <sub>1</sub> receptor	11
1.3.2.2 EP <sub>2</sub> receptor	11
1.3.2.3 EP <sub>3</sub> receptor	12
1.3.2.4 EP <sub>4</sub> receptor	12
1.3.3 FP receptor	13
1.3.4 IP receptor	13
1.3.5 TP receptor	14
1.4 Prostanoid receptor ligands	14
1.4.1 Selective agonists	15
1.4.2 Selective antagonists	21
1.5 Physiological role of prostanoids in the human body	25
1.5.1 Heart	25
1.5.2 Thrombosis and haemostasis	26

1.5.3	Hypertension	28
1.5.4	Pulmonary vasculature	29
1.5.5	Kidney function	30
1.5.6	Ductus arteriosus	31
1.5.7	Intraocular pressure	32
1.6	Diabetes	33
1.6.1	Diabetes overview	33
1.6.2	Diabetes and endothelium	33
1.6.3	Diabetes and prostaglandins	34
1.7	Pharmacological principles	35
1.7.1	Synergism and its measurement	35
1.7.2	Types of antagonism and $pA_2$ measurement	36
1.8	Project aims	44
<b>CHAPTER TWO: GENERAL METHODS AND MATERIALS</b>		<b>46</b>
2.1	Tissue isolation	47
2.1.1	Guinea-pig trachea	47
2.1.2	Rat mesenteric artery	47
2.1.3	Rat urinary bladder	48
2.1.4	Rat femoral artery	48
2.2	Wire myograph	51
2.2.1	Krebs solution	51
2.2.2	Organ bath	51
2.2.2.1	Guinea-pig trachea	51
2.2.2.2	Rat mesenteric artery	52
2.2.2.3	Rat urinary bladder	53
2.2.2.4	Rat femoral artery	54
2.3	Streptozotocin-induced diabetes	54
2.4	Antagonist protocol and $pA_2$ estimation	55

2.5	Data analysis	57
2.5.1	Statistical analysis	57
2.5.2	Two-site competition model	58
2.6	Reagents and chemicals	59
 <b>CHAPTER THREE: GUINEA-PIG TRACHEA</b>		<b>63</b>
3.1	Introduction	64
3.2	Methods	66
3.2.1	Setting up of preparations	66
3.2.2	Statistical analysis	67
3.3	Results	69
3.3.1	Effects of histamine	69
3.3.2	Effects of prostanoid agonists	69
3.3.3	Effects of PGE <sub>2</sub> and butaprost on precontracted guinea-pig tracheal ring under low tone	77
3.3.4	Comparison of EP <sub>2</sub> agonists on precontracted guinea-pig tracheal ring under low tone and high tone	79
3.3.5	Two-site competition model	87
3.4	Discussion	89
3.4.1	The tracheal response to prostanoid agents	89
3.4.2	Relaxant response of the trachea to EP <sub>2</sub> agonists	90
3.4.3	Dual receptor activity of AH-13205	91
3.5	Conclusions	92
 <b>CHAPTER FOUR: RAT MESENTERIC ARTERY</b>		<b>93</b>
4.1	Introduction	94
4.2	Methods	98
4.2.1	Setting up of preparations	98
4.2.2	Antagonist protocol and pA <sub>2</sub> estimation	99

4.2.3	Statistical analysis	102
4.3	Results	103
4.3.1	Initial investigations	103
4.3.2	Effects of U-46619 and BMS-180291	106
4.3.3	Effects of prostaglandin E <sub>2</sub>	109
4.3.4	Interaction of L-798106, L-826266 and SC-51322 with sulprostone	110
4.3.5	Interaction of 17-phenyl PGE <sub>2</sub> with SC-51322 and L-826266	115
4.3.6	Effects of ONO-D1-004 and ONO-AE-248	120
4.3.7	Relaxant studies	121
4.3.8	Two-site competition model	122
4.4	Discussion	124
4.4.1	Defining optimum conditions for studying prostanoid systems	125
4.4.2	The artery response to non-prostanoid agents	125
4.4.3	Defining the prostanoid receptors involved in the contractile response	126
4.4.4	The two-site competition model versus one-site competition model in relation to EP receptor-mediated contraction	131
4.4.5	Utility of the novel EP agonists	131
4.4.6	Relaxant EP and IP receptors	132
4.5	Conclusions	133
 <b>CHAPTER FIVE:    RAT URINARY BLADDER</b>		<b>134</b>
5.1	Introduction	135
5.2	Methods	141
5.2.1	Setting up of preparations	141
5.2.2	Antagonist protocol and pA <sub>2</sub> estimation	142
5.2.3	Statistical analysis	144
5.3	Results	145
5.3.1	Initial investigations of rat urinary bladder strips	145
5.3.2	Effects of SC-51322 on contractile effects of prostanoid agonists	147



5.3.3	Effects of L-798106 on contractile effects of prostanoid agonists	151
5.3.4	Effects of a combination, of SC-51322 and L-798106 on contractile effects of prostanoid agonists	155
5.3.5	Antagonist potency of L-798106 under inhibition-curve protocol	159
5.3.6	Effects of GW-848687 on contractile effects of prostanoid agonists	161
5.3.7	Antagonist potency of GW-848687 under inhibition-curve protocol	163
5.3.8	Two-site competition model	166
5.4	Discussion	171
5.4.1	General overview	171
5.4.2	Response to prostanoid agonists	172
5.4.3	The nature of the EP receptor(s) mediating contraction of the rat urinary bladder preparation	174
5.4.4	Two-site competition model versus one-site competition model	177
5.4.5	Insurmountable antagonism and the hemi-equilibrium concept	178
5.4.6	EP <sub>4</sub> receptor	179
5.5	Conclusions	180
<b>CHAPTER SIX: RAT FEMORAL ARTERY AND DIABETES</b>		<b>181</b>
6.1	Introduction	182
6.2	Methods	191
6.2.1	Streptozotocin induced diabetes	191
6.2.2	Setting up of preparations	193
6.2.3	Antagonist protocol and pA <sub>2</sub> estimation	194
6.2.4	Statistical analysis	194
6.3	Results	195
6.3.1	General condition of animals	195
6.3.2	Effects of endothelium removal on acetylcholine	196
6.3.3	Effects of U-46619 and its block by BMS-180291	197

6.3.4	Effects of endothelium on potassium chloride and phenylephrine contraction	200
6.3.5	Effects of phenylephrine on streptozotocin and control arteries	202
6.3.6	Effects of prostanoid antagonists on phenylephrine-induced contraction	203
6.3.7	Effects of COX-inhibitors on phenylephrine responses	207
6.4	Discussion	210
6.4.1	General response to streptozotocin	210
6.4.2	Endothelium removal and effects on contractile response	210
6.4.3	Induction of endogenous prostanoids by diabetes	212
6.4.4	Roles of prostacyclin	216
6.5	Conclusions	217
<b>CHAPTER SEVEN: GENERAL DISCUSSION AND CONCLUSION</b>		<b>218</b>
7.1	Functional studies	219
7.2	Streptozotocin-induced diabetes study	222
<b>REFERENCES</b>		<b>224</b>
<b>APPENDIX</b>		<b>256</b>

## ABBREVIATIONS

5-HT	- 5-hydroxytryptamine
AA	- Arachidonic acid
AMI	- Acute myocardial infarction
ANOVA	- Analysis of variance
Ba <sup>2+</sup>	- Barium
BOO	- Bladder outlet obstruction
Ca <sup>2+</sup>	- Calcium
[Ca <sup>2+</sup> ] <sub>i</sub>	- Intracellular calcium
cAMP	- Cyclic adenosine monophosphate
CI	- Confidence interval
ClogP	- Predicted <i>n</i> -octanol / water partition coefficient
COX	- Cyclo-oxygenase
CRC	- Concentration response curve
CRTh2	- Chemoattractant receptor-homologous molecule of Type 2 helper T cells
DAG	- Diacylglycerol
df	- Degrees of freedom
DM	- Diabetes mellitus
DP receptor	- Prostaglandin D receptor
DR	- Dose ratio
EC	- Endothelial cells
EC <sub>50</sub>	- The molar concentration of an agonist that produces 50% of the maximal possible effect of that agonist
EDHF	- Endothelium derived hyperpolarising factor
EDRF	- Endothelium derived relaxant factor
EH	- Essential hypertension
E <sub>max</sub>	- The maximal response by the contractile agent / agonist attained
EMR	- Equi-effective molar ratio
EP receptor	- Prostaglandin E receptor
FLIPR	- Fluorometric imaging plate reader

FA	- Free acid
FP receptor	- Prostaglandin F <sub>2α</sub> receptor
GK	- Goto-Kakizaki
GPT	- Guinea-pig trachea
HPO	- 15-hydroperoxidase
HPT	- Hypertension
i.p.	- Intraperitoneal
I/R	- Ischaemic-reperfusion
IC <sub>50</sub>	- The molar concentration of an antagonist that reduces the response to an agonist by 50%
IOP	- Intraocular pressure
IP receptor	- Prostacyclin receptor
IP3	- Inositol triphosphate
IsoPs	- Isoprostanes
K <sub>b</sub>	- Equilibrium dissociation constant
KCl	- Potassium chloride
K <sub>i</sub>	- Antagonist dissociation constant
KO	- Knockout
NSAID	- Nonsteroidal anti-inflammatory drug
pA <sub>2</sub>	- The negative logarithm to base 10 of the molar concentration of an antagonist that makes it necessary to double the concentration of the agonist to elicit the original submaximal response
PAH	- Pulmonary artery hypertension
pEC <sub>50</sub>	- The negative logarithm to base 10 of the EC <sub>50</sub> of an agonist
PG	- Prostaglandin
PGD <sub>2</sub>	- Prostaglandin D <sub>2</sub>
PGE <sub>2</sub>	- Prostaglandin E <sub>2</sub>
PGF <sub>2α</sub>	- Prostaglandin F <sub>2α</sub>
PGHS	- Prostaglandin H synthase
PGI <sub>2</sub>	- Prostacyclin
PGIS	- Prostacyclin synthase

Phe	- Phenylephrine
PIP	- Phosphatidyl inositol phosphate
PKA	- Protein kinase A
PKC	- Protein kinase C
PLA <sub>2</sub>	- Phospholipase A <sub>2</sub>
PRP	- Platelet-rich plasma
$r^2$	- Regression correlation coefficient
RFA	- Rat femoral artery
RMA	- Rat mesenteric artery
RUB	- Rat urinary bladder
SMC	- Smooth muscle cells
SS	- Sum-of-squares
STZ	- Streptozotocin
Th2	- Type 2 helper T cells
TP receptor	- Thromboxane receptor
TXA <sub>2</sub>	- Thromboxane A <sub>2</sub>
TXB <sub>2</sub>	- Thromboxane B <sub>2</sub>
VSM	- Vascular smooth muscles
WT	- Wild-type
ZF	- Zuckers fatty

# **ABSTRACT**

The demise of the COX-2 inhibitors in treating rheumatoid arthritis has renewed interest in the development of prostanoid receptor antagonists. While data from radio-ligand binding assays remain useful, functional studies on isolated smooth muscle preparations are also valuable. The present study focussed on the characterization of EP<sub>1</sub> and EP<sub>3</sub> receptors and their novel antagonists in the rat mesenteric artery and the rat urinary bladder. Inhibition-curve protocols (based on the Cheng-Prusoff equation) were used in the first instance and data were analysed by the two-site competition equation in combination with the modified F test to assess the presence of two contractile systems.

In agreement with literature, the prostaglandin receptors in guinea-pig trachea mediating contractile responses appeared to be EP<sub>1</sub> and TP, whereas the relaxant response was attributed to the EP<sub>2</sub> receptor. Contractile responses to the TP agonist, U-46619 contractile were prevented by a selective TP antagonist. Equally, the selective EP<sub>1</sub> antagonist, SC-51322 completely antagonised the contractile response to 17-phenyl PGE<sub>2</sub>. The EP<sub>2</sub> agonist AH-13205 showed no evidence of partial agonism, and the apparent two site activities are more likely to be explained by the presence of two isomers having different activities.

In the rat mesenteric artery, only contractile prostanoid receptors were demonstrated with equal contributions from EP<sub>1</sub> and EP<sub>3</sub> receptors. The two-site competition model produced a better fit than the one-site model to the inhibition curve of SC-51322 against 17-phenyl PGE<sub>2</sub>.

The contractile responses in the rat urinary bladder appeared to be mediated, in part, by EP<sub>1</sub> receptors but other prostanoid receptor contribute and these remain to be elucidated. The possibility of non-competitive behaviour or a hemi-equilibrium state of antagonist-agonist interaction needs to be considered.

The contractile responses to phenylephrine and KCl in the endothelium-denuded rat femoral artery were not modified by streptozotocin-induced diabetes. The phenylephrine response was reduced by the EP<sub>3</sub> antagonist, L-798106 but not by the TP antagonist. COX-inhibitors did not affect the contractile response. These results suggest the involvement of an endogenous facilitator of contractile responses that acts on EP<sub>3</sub> receptors but which is not generated via the COX pathway.

In conclusion, conventional organ bath methodology coupled with the inhibition-curve protocol is useful to assess pharmacological antagonism and to characterize prostanoid receptors in isolated tissue preparations.

# **CHAPTER ONE**

## **GENERAL INTRODUCTION**



Prostanoids comprise prostaglandins (PGs) and thromboxanes (TXs). The first prostanoid receptor antagonist was described in the late 1960s. SC-19220 (Sanner, 1969) inhibited the contractile actions of prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) on intestinal smooth muscle preparations by blocking what we now know to be the EP<sub>1</sub> receptor (see later). Since then, the discoveries of other natural prostanoids, prostaglandin D<sub>2</sub> (PGD<sub>2</sub>), thromboxane A<sub>2</sub> (TXA<sub>2</sub>), prostacyclin (PGI<sub>2</sub>) and prostaglandin F<sub>2α</sub> (PGF<sub>2α</sub>) and their specific receptors have been followed by corresponding developments of receptor antagonists. However, the pace of development quickened in the mid-1990's owing to the isolation and molecular characterization of these prostanoid receptors. These receptors are termed P receptors, with a preceding letter indicating the natural prostanoid to which each receptor is most sensitive - i.e. DP, EP, FP, IP and TP, respectively (Kennedy *et al.*, 1982). Subsequently, single recombinant prostanoid receptors were introduced into carrier cells to produce efficient screening assays, paralleling developments for other receptors systems (e.g. 5-HT<sub>1A</sub>, Albert *et al.*, 1990). Accompanying this trend was the use of combinatorial chemistry to produce many closely related analogues with much greater efficiency than classical synthetic methods. The outcome has been the appearance in the literature of novel antagonists for DP<sub>1</sub> and DP<sub>2</sub> receptors and EP receptors. EP receptors are subdivided into four groups, EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub> and EP<sub>4</sub>, originally based on their relative sensitivities to a range of selective agonists and antagonists (see later).

While there is a temptation to proceed to *in vivo* testing of specific agents based on data from radio-ligand binding assays, information from functional studies on isolated tissues can be of considerable value. These first-line preparations ought to be robust and reproducible in their responses to prostanoid standard agonists. Preparations containing a single prostanoid receptor are rare; one such preparation is the rabbit ear artery, which only has an EP<sub>2</sub> receptor (Lydford and McKechnie, 1994). Multiple prostanoid receptors can be a disadvantage in characterizing an antagonist, particularly if opposing agonist actions are possible. Thus activation of EP<sub>1</sub> receptors in HEK-293 cells by 17-phenyl PGE<sub>2</sub> functionally antagonized the TP receptor-induced [Ca<sup>2+</sup>]<sub>i</sub> mobilization (Walsh and Kinsella, 2000). On the other hand, multiple receptors do allow one to test the specificity of the antagonist. In addition, rapid responses to the standard agonist are an

advantage in constructing consecutive concentration-response curves. Many tissues in conventional organ baths (e.g. rabbit and guinea-pig aortic rings) exhibit slow responses to potent agonists and it was hoped that the use of thinner tissues in a Mulvany (wire) myograph would rectify this problem (Mulvany and Halpern, 1977).

In the following sections, I shall describe the biosynthesis of the natural prostanoids, the nature of the prostanoid receptors on which they act, and the properties of the currently available agonists and antagonists for these receptors. The receptor / second messenger nomenclature used in the current study conforms to internationally accepted practice (Neubig *et al.*, 2003; Alexander *et al.*, 2008).

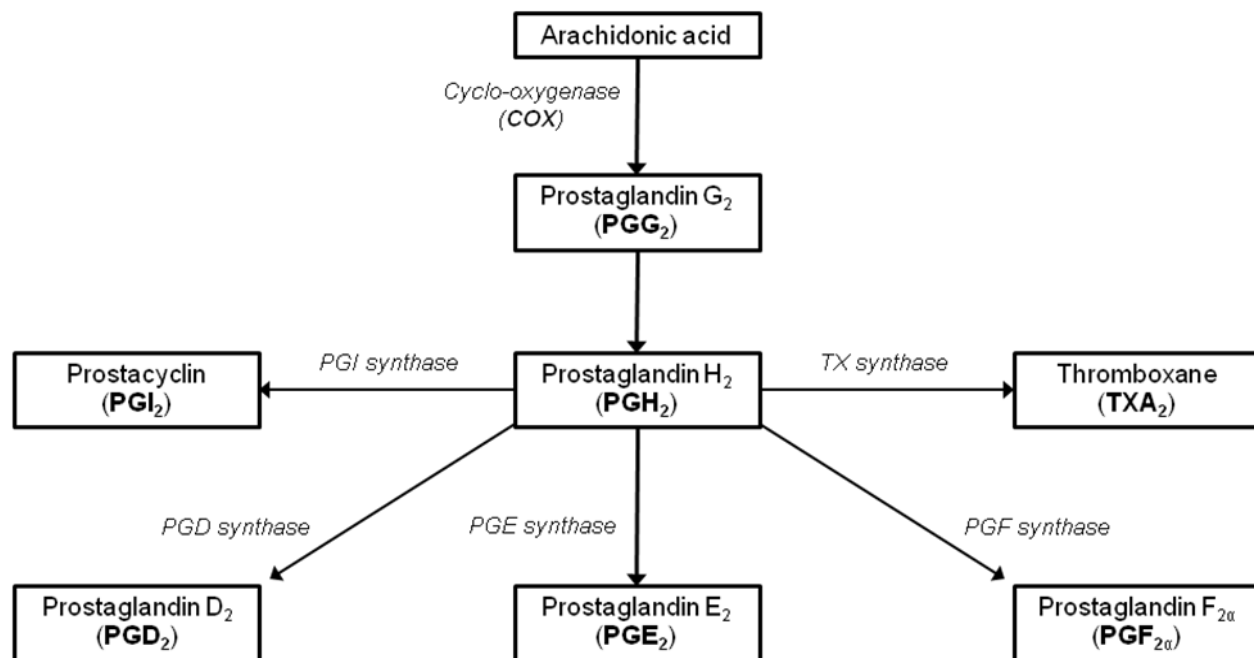
## 1.1 Prostaglandin biosynthesis

Prostanoids are a group of lipid hormone mediators that are derived from C-20 fatty acids, particularly arachidonic acid (AA). They consist of the prostaglandins and thromboxane. AA has two quite different functions in the body. As a component of cell membrane phospholipids, its *cis*-double bonds disorder the hydrophobic core of the membrane. This will influence the fluidity, permeability and the behaviour of embedded proteins. However, of greater interest is its second function, where in response to various physiological and pathological stimuli, this fatty acid is liberated from the internal membranes by the activity of phospholipase A<sub>2</sub> (PLA<sub>2</sub>). AA that is liberated will be converted to prostaglandin G<sub>2</sub> and then to prostaglandin H<sub>2</sub> (PGH<sub>2</sub>) by the sequential actions of cyclo-oxygenase (COX) and 15-hydroperoxidase (HPO).

PGH<sub>2</sub> serves as a substrate for different prostanoid synthase enzymes, which are responsible for the production of the five principle bioactive prostanoids generated *in vivo*, PGE<sub>2</sub>, PGF<sub>2α</sub>, PGD<sub>2</sub>, PGI<sub>2</sub>, and TX<sub>A</sub>. The prostanoids produced by a given cell largely depend on the expression

profile of the individual prostanoid synthase enzyme. This is summarized in Figure 1.1 (redrawn from Hata and Breyer, 2004).

There are currently three known COX isoforms: COX-1, COX-2 and COX-3. The expression of the three COX isoenzymes is differently regulated. COX-1 gene exhibits the features of a housekeeping gene, constitutively expressed in most tissues. COX-2 expression is more tightly regulated and under most normal physiological conditions, it is not expressed. However, under a variety of pathophysiological conditions its expression can be rapidly induced (Scholz, 2003). Hence, it is widely believed that prostanoids (PGs) derived from COX-1 play a role in normal body function, whereas those from COX-2 are associated with a pathological process in the body. COX-3 was discovered later on and has drawn a considerable attention (Hirai *et al.*, 2001). It is a splice variant of the COX-1 gene isolated from canine brain that retains intron 1. Unlike COX-1 and COX-2, this isoenzyme is highly responsive to acetaminophen. However, COX-3 is not expressed in humans and its relevance to human pathophysiological processes remains questionable.



**Figure 1.1** Biosynthesis of prostanoids. Redrawn from Hata and Breyer, 2004.

## 1.2 Molecular biology

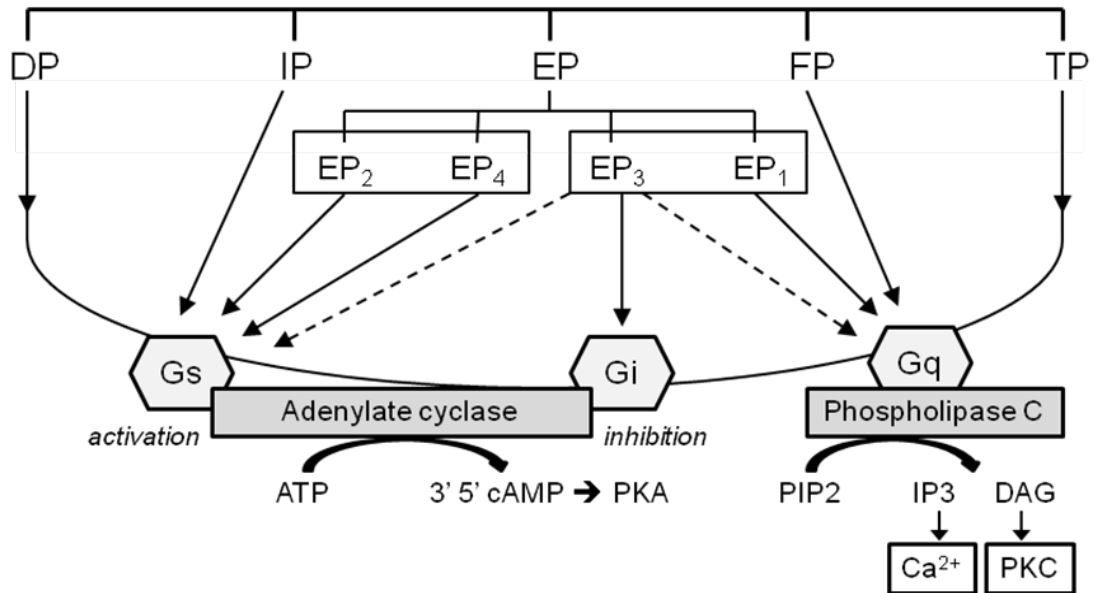
An individual gene encodes each prostanoid receptor. The receptors responsive to COX products are evolutionally distinct from the leukotriene receptors of the 5-lipoxygenase pathway (Toh *et al.*, 1995). The first cluster includes EP<sub>2</sub>, EP<sub>4</sub>, DP and IP receptors, which induce smooth muscle relaxation via activation of adenylate cyclase. EP<sub>1</sub>, FP and TP receptors, which cause smooth muscle contraction, form another cluster. On its own, the EP<sub>3</sub> receptor, which usually stimulates smooth muscle contraction, defines the third cluster.

### 1.2.1 Isoforms

A striking feature of the EP<sub>3</sub> receptor that sets it apart from the other EP receptors is the large number of splice variants generated by alternate splicing of the C-terminal tail. In humans, at least eight EP<sub>3</sub> splice variants have been identified, and multiple splice variants exist for other species. In humans, the TP receptor exists as two alternatively spliced variants, TP<sub>α</sub> (placental / platelet) and TP<sub>β</sub> (endothelial) (Breyer *et al.*, 2001). Despite the existence of these variants, the mRNAs for both splice variants have been detected in most tissues that express TP receptor including platelets, placenta, vascular smooth muscle, brain, small intestine, and thymus (Raychowdhury *et al.*, 1994; Miggin *et al.*, 2002). Two differentially spliced variants of the FP receptor have been reported (FP<sub>A</sub> and FP<sub>B</sub>), which differ from each other in C-terminal tail length. Both are coupled to G<sub>q</sub> protein but have a different sensitivity to desensitization (Hata and Breyer, 2004). The C-terminal of tail of the FP<sub>A</sub> can be phosphorylated by PKC, leading to agonist-induced desensitization and internalization. Generally, prostanoid receptor isoforms exhibit similar ligand responsiveness (Wright *et al.*, 2001).

## 1.2.2 Signal transduction

The signal transduction pathways of prostanoid receptors have been studied by examining agonist-induced changes in the levels of second messengers (cAMP, free  $\text{Ca}^{2+}$ , and inositol phosphates), and by identifying G protein coupling by various methods. The first cluster of receptors, which is normally associated with smooth muscle relaxation (EP<sub>2</sub>, EP<sub>4</sub>, DP and IP), couple via G<sub>s</sub> to mediate increase in intracellular cAMP levels (Breyer *et al.*, 2001). The increases in intracellular cAMP were demonstrated after stimulation of the recombinant human EP<sub>2</sub> (Regan *et al.*, 1994), EP<sub>4</sub> (Bastien *et al.*, 1994), DP (Boie *et al.*, 1995) and in IP (Boie *et al.*, 1994) receptors. Similarly, PGD<sub>2</sub> and PGE<sub>1</sub> / PGI<sub>2</sub>-responsive receptors have been shown to stimulate the production of cAMP in platelets (Whittle *et al.*, 1978). EP<sub>1</sub>, FP and TP receptors couple via G<sub>q</sub> of phospholipase C, with subsequent liberation of inositol phosphate (Breyer *et al.*, 2001). The end-result is to elevate  $\text{Ca}^{2+}$ , leading to muscular contraction. This pathway has been demonstrated in the platelet; the stimulation of TP receptor involved the G<sub>q</sub> activation as the primary effector pathway (Arita *et al.*, 1989). The EP<sub>3</sub> receptor, which constitutes the final cluster, has isoforms that inhibit adenylate cyclase through G<sub>i</sub> protein; individual isoforms may also couple to G<sub>q</sub> and even G<sub>s</sub> (Breyer *et al.*, 2001). Figure 1.2 summarizes the signaling modulator involved in each receptor.



**Figure 1.2** Prostanoid receptors, signal transduction and their final effects. Adapted from Wise and Jones, 2000. Broken lines indicate alternative coupling modes of isoforms of the EP<sub>3</sub> receptor.

### 1.3 Prostanoid receptors classification and distribution

Prostanoids are ubiquitously produced. It is believed that prostanoids work locally in autocrine or paracrine manner. By acting through prostanoid receptor subtypes, these chemical compounds elicit various pharmacological and pathopharmacological reactions in different tissue types. The prostanoid receptor family consists of eight distinct rhodopsin-like receptor proteins, which are classified according to the natural prostanoid ligand that each binds with greatest affinity. The first receptor purified and cloned was the human platelet TXA<sub>2</sub> receptor in 1991 by Hirata *et al.* Since then, homology screening based on amino acid sequence of this receptor was performed in various species. These receptors have been expressed, and their ligand binding properties and signal transductions have been examined. The tissue and cell distribution of the receptors was studied by Northern blot and by *in situ* hybridization analyses of their mRNA expression. Correlation of such knowledge with findings accumulated by pharmacological studies using COX inhibitors and using various prostanoid analogues having agonistic and antagonistic activities helps to define the actions of these receptors. They also help to reveal novel actions of these receptors.

Earlier classification of the prostanoid receptor was based on the study of isolated tissues preparation. Collectively, the receptors have been termed prostanoid DP, EP, FP, IP and TP receptors based on the agonist potencies in functional systems (Kennedy *et al.*, 1982). Further studies subdivided the EP receptor into four subtypes based on their agonist / antagonist specificity and signal transduction, each denoted by a subscript numeral (i.e. EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub> and EP<sub>4</sub>) (Coleman *et al.*, 1994b). PGD<sub>2</sub> as been identified as a potent agonist at the CRTh2 (chemoattractant receptor-homologous molecule of Type 2 helper T cells) or DP<sub>2</sub> receptor (Hirai *et al.*, 2001). Thus, including DP<sub>2</sub> receptor, the total number of prostanoid receptor subtypes is nine. The original DP receptor is now known as the DP<sub>1</sub> receptor. However, the DP<sub>2</sub> receptor does not contain the characteristic molecular signatures of prostanoid receptors but rather of chemokine receptors (Nagata *et al.*, 1999). This receptor will be not discussed further as it is outside the scope of the current study.



### 1.3.1 DP<sub>1</sub> Receptor

The DP<sub>1</sub> receptor shows significant sequence identity with IP and EP<sub>2</sub> receptors (Boie *et al.*, 1997). Among the prostanoid receptors, the DP<sub>1</sub> receptor is the least abundant. In humans, on Northern blot analysis, its expression was detected in retina and small intestine. However, human platelets contain functional DP<sub>1</sub> receptor (Giles *et al.*, 1989). DP<sub>1</sub> has been implicated in regulation of immune and skin allergic responses (Angeli *et al.*, 2004). This is correlated with earlier study where PGD<sub>2</sub> induced an asthmatic response in WT-mice but not in DP-KO-mice (Matsuoka *et al.*, 2000).

### 1.3.2 EP Receptor

The EP receptor family is subdivided into four subtypes: EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub>, and EP<sub>4</sub>. The EP<sub>3</sub> receptor was the first to be cloned (Sugimoto *et al.*, 1994), followed by the EP<sub>1</sub> and EP<sub>4</sub> receptors (Funk *et al.*, 1993; Honda *et al.*, 1993). The EP<sub>2</sub> receptor was later cloned and pharmacologically characterized (Regan *et al.*, 1994). The main endogenous prostanoid that displays the highest potency at receptors of the EP type is PGE<sub>2</sub>. It is a major COX product in a number of physiological settings and has by far the largest variety of biological actions. Of the four EP receptors, the EP<sub>3</sub> and EP<sub>4</sub> receptors bind PGE<sub>2</sub> with highest affinity ( $K_d < 1$  nM), whereas the EP<sub>1</sub> and EP<sub>2</sub> receptors bind with lower affinity ( $K_d > 10$  nM, Abramovitz *et al.*, 1994).

PGE<sub>2</sub> usually relaxes isolated vascular preparations and this action is mediated by activation of the EP<sub>2</sub> and / or EP<sub>4</sub> prostanoid receptor subtypes (Narumiya *et al.*, 1999). However, PGE<sub>2</sub> can also activate other prostanoid receptors depending on the vascular preparation examined. In particular, PGE<sub>2</sub> has been shown to activate the EP<sub>3</sub> receptor (Qian *et al.*, 1994; Jadhav *et al.*, 2004) and EP<sub>1</sub> receptor (Jadhav *et al.*, 2004) leading to contraction of vascular smooth muscle.

In some vessels, for example the rat femoral artery, PGE<sub>2</sub> is a weak contractile agent, but exhibits pronounced synergism with strong vasoconstrictors such as phenylephrine ( $\alpha_1$ -adrenoceptor agonist) and U-46619 (TP receptor agonist) (Hung *et al.*, 2006).

Correspondingly, PGE<sub>2</sub> elicits vasodilatation and / or vasoconstriction *in vivo* in wild-type animals. A study in EP<sub>2</sub>-KO mice showed that intravenous infusions of PGE<sub>2</sub> and sulprostone (EP<sub>1/3</sub> agonist) raise blood pressure (Kennedy *et al.*, 1999). When fed on a high-salt diet, the EP<sub>2</sub>-KO animals developed significant hypertension with concomitant increase in urinary excretion of PGE<sub>2</sub>. These results indicate that PGE<sub>2</sub> is produced in the body in response to a high-salt diet and works to negatively regulate the blood pressure via the relaxant EP<sub>2</sub> receptor; dysfunction of this pathway may be involved in producing the salt-sensitive hypertension.

### **1.3.2.1 EP<sub>1</sub> receptor**

The EP<sub>1</sub> receptor was originally characterized as coupling to stimulation of intracellular Ca<sup>2+</sup>, and little still known about other signal transduction pathways (Hata and Breyer, 2004). The EP<sub>1</sub> receptor is restricted to several organs, such as lungs, stomach and in the kidneys (Breyer and Breyer, 2001).

### **1.3.2.2 EP<sub>2</sub> receptor**

The EP<sub>2</sub> receptor plays a role in uterine implantation and salt-sensitive hypertension, but most importantly is in relaxing vascular and bronchiolar smooth muscle. Human EP<sub>2</sub> receptors are mainly expressed in uterus, lungs and small intestine (Boie *et al.*, 1997, Bastien *et al.*, 1994,

Regan *et al.*, 1994; Breyer *et al.*, 2001). However, Jensen *et al* (2001) showed that the EP<sub>2</sub> receptor could be found in the rat kidneys.

### **1.3.2.3 EP<sub>3</sub> receptor**

The EP<sub>3</sub> receptor is known to have a variety of actions due to its wide tissue distribution. It is highly expressed in the kidneys, uterus, adrenal gland and stomach tissues (Breyer *et al.*, 2001). In brief, it mediates contraction of smooth muscle, including vascular and uterine smooth muscle, enhances platelet aggregation, inhibits lipolysis and gastric acid secretion, and elicits cytoprotection of the gut. The EP<sub>3</sub> receptors have been implicated in febrile responses to pyrogens (Ushikubi *et al.*, 1998).

### **1.3.2.4 EP<sub>4</sub> receptor**

The EP<sub>4</sub> receptor is widely distributed throughout the body and its mRNA has been found to be expressed in almost all mouse tissues examined. Important vasodilator effects of EP<sub>4</sub> receptor activation have been described in venous and arterial beds (Coleman *et al.*, 1994a; Coleman *et al.*, 1994b). A particular role for the EP<sub>4</sub> receptor in regulating the closure of the pulmonary ductus arteriosus has also been suggested by the studies of EP<sub>4</sub>-KO mice (Nguyen *et al.*, 1997; Segi *et al.*, 1998).

### **1.3.3 FP receptor**

The FP receptor was originally cloned from human kidneys, uterus, and placental cDNA libraries (Abramovitz *et al.*, 1994). Activation of FP receptor in the cardiovascular system increases systemic blood pressure, partly due to vasoconstriction (Ducharme *et al.*, 1968). PGF<sub>2α</sub>, an endogenous FP agonist plays a critical role in mammalian reproduction by inducing luteolysis (Poyser, 1995). It has also been implicated in influencing renal function, regulation of intraocular pressure and in proliferative states such as endometrial carcinoma and cardiac hypertrophy. This reflects the fact that FP receptor expression has been demonstrated in the corpus lustrum, the kidneys, ocular tissues and ventricular myocytes (Hata and Breyer, 2004).

### **1.3.4 IP receptor**

The human IP receptor was first cloned from lung and megakaryote cDNA libraries and has been shown to be expressed in many tissues including the kidneys, liver, lungs, platelets, heart and aorta (Wise and Jones, 2000; Breyer *et al.*, 2001). Activation of the IP receptor has been implicated in vascular homeostasis, because of vasodilatory and antithrombotic effects (see next section). Apart from its vascular effects, the IP receptor has been shown to mediate nociceptive pain during acute inflammation (Murata *et al.*, 1997; Bley *et al.*, 2006). In contrast to the pro-inflammatory effects of this receptor activation in nonallergic acute inflammation, recent studies have suggested that IP receptor signaling suppresses Th2-mediated allergic inflammatory responses. By suppressing the Th2 cell production, lung inflammation is greatly reduced (Takahashi *et al.*, 2002).

### 1.3.5 TP receptor

The TP receptor is important in modulating cardiovascular function. Stimulation by its endogenous agonists, PGH<sub>2</sub> and TXA<sub>2</sub> leads to shape change and aggregation of blood platelets. Deficiency in TP receptors leads to coagulation defects (Thomas *et al.*, 1998). The TP receptor is also responsible for smooth muscle contraction in various tissues (Eglen and Whiting, 1988). TP receptor signaling has been proposed to play a role in modulating T cell activation. Despite the existence of two variants, the mRNAs for both splice variants have been detected in most tissue that express TP receptor including platelets, placenta, vascular smooth muscle, brain, small intestine, and thymus (Raychowdhury *et al.*, 1994; Miggin *et al.*, 2002).

## 1.4 Prostanoid receptor ligands

Most of the endogenous prostanoids are prone to be metabolised into inactive compounds. In addition, PGI<sub>2</sub> and TXA<sub>2</sub> are chemically unstable under physiological conditions and their synthetic analogues have usually been used for receptor studies. The structure of synthetic agonists is usually close to the natural agonists, although non-prostanoid agonists exist for EP<sub>3</sub> and IP receptors (Jones *et al.*, 2009). Several routes to prostanoid antagonists have been followed. Some antagonists have emerged by serendipity (SC-19220; Sanner, 1969), while others have been derived from the chemical modification of a partial agonist (EP-045; Armstrong *et al.*, 1985). High throughput screening using cloned receptors is now the dominant route. For functional assays, the additional introduction of chimeric G-protein allows all prostanoid receptors to be investigated through mobilization of [Ca<sup>2+</sup>]<sub>i</sub> using the fluorometric imaging plate reader (FLIPR) technology (Matias *et al.*, 2004; Woodward *et al.*, 2007). In this technique, all 96 wells are stimulated and optically measured simultaneously. Bimaprost has been demonstrated to selectively bind to HEK-293 cells expressing the cloned human ciliary body FP receptor (Sharif *et al.*, 2003).

### 1.4.1 Selective agonists

Table 1.1 summarizes the prostanoid receptor subtypes and their selective agonists (Jones *et al.*, 2009). Although of considerable pharmacological interest, partial agonists (right-hand column of Table 1.1) are not usually preferred as standard agonists. Table 1.2 listed the  $K_i$  value of the agonists used in the current study. Figure 1.3 and 1.4 listed the chemical structure of prostanoid agonists used in the current study.

17-Phenyl- $\omega$ -trilor PGE<sub>2</sub> (17-phenyl PGE<sub>2</sub>) is a moderately selective EP<sub>1</sub> agonist, and in combination with sulprostone (EP<sub>3</sub> > EP<sub>1</sub>) it can be used to discriminate between EP<sub>1</sub> and EP<sub>3</sub> receptors. A 6-oxo PGE<sub>1</sub> analogue, ONO-DI-004 also has been reported to be EP<sub>1</sub> selective (Cao *et al.*, 2002; Norel *et al.*, 2004). The prostacyclin analogue, iloprost behaved as a potent partial agonist on the EP<sub>1</sub> receptor (Dong *et al.*, 1986).

Two commonly used EP<sub>2</sub> agonists are butaprost, which is moderately potent, and AH-13205, which has a low potency. However, ONO-AEI-259 is increasingly being used in preference to butaprost, as it is more potent; its selectivity for EP<sub>2</sub> systems is well demonstrated by the studies of Jones and Chan (2005) and Hung *et al.* (2006). CAY-10399, a close relative of butaprost, has a high selectivity on mouse EP<sub>2</sub> receptor with binding  $K_i$  value of 2.2 nM; it exhibits much less IP agonism than butaprost (Tani *et al.*, 2001). A non-prostanoid EP<sub>2</sub> agonist, CP-533536 has recently been reported (Cameron *et al.*, 2009).

Sulprostone is a standard agonist used to identify EP<sub>3</sub> receptors as it has the highest affinities for this receptor; it has been demonstrated to be better in controlling post-partum haemorrhage compared with placebo (Poeschmann *et al.*, 1991). However, cardiovascular complications were reported in two patients (Stock *et al.*, 1995), which are likely due to its direct pulmonary

vasoconstrictor action (Qian *et al.*, 1994). Previously, SC-46275 was shown to be the most selective ligand for this receptor (Jones, 2004; Wilson *et al.*, 2004).

A selective agonist has been developed for EP<sub>4</sub> receptor. ONO-AE-329 binds selectively to mouse EP<sub>4</sub> receptor with inhibitory constant ( $K_i$ ) of 10 nM and to human receptor with  $K_i$  of 24 nM (Tsuboi *et al.*, 2002). The non-prostanoid ONO-AP-324 has also been reported to behave as a partial agonist in some preparations (Jones *et al.*, 1998).

PGI<sub>2</sub> has limited use to study IP receptor function due to its unstable nature. Therefore, synthetic agonists such as iloprost, cicaprost, AFP-07 and carbacyclin are commonly used. Cicaprost is the most selective of these agonists (Jones, 2004). Iloprost potently activates the IP receptor but activates the EP<sub>1</sub> receptor as well (Dong *et al.*, 1986). Taprostene behaves as a partial agonist at the IP receptor (Jones and Chan, 2001).

Endogenous TXA<sub>2</sub> is unstable in aqueous solution, being rapidly hydrolyzed to TXB<sub>2</sub>, and is therefore not used in receptor binding and signal transduction assays. For that reason, analogues such as U-46619 have been synthesized and most commonly used to probe TP receptor function. Other TP agonists include I-BOP and STA<sub>2</sub>. Partial agonists are common, for example CTA<sub>2</sub>, PTA<sub>2</sub> and U-44069 (see Jones, 2004).

Selective FP agonists such as fluprostenol and latanoprost-FA have been developed (see Jones, 2004). Fluprostenol could only bind to the FP receptor, indicating the high selectivity of this ligand, with  $K_i$  value of 2.7 nM in human receptor.

BW-245C has been widely used as a selective DP<sub>1</sub> agonist.

**Table 1.1** Prostanoid receptor agonists for defining antagonist profiles (Jones *et al.*, 2009)

Prostanoid receptor	Full agonists		Partial agonists
	High selectivity	Moderate selectivity	
EP <sub>1</sub>	ONO-D1-004	17-Phenyl PGE <sub>2</sub>	Iloprost
EP <sub>2</sub>	ONO-AE1-259, CAY-10399	Butaprost-FA, CP-533536, 19(R)-hydroxy PGE <sub>2</sub>	
EP <sub>3</sub>	ONO-AE-248, SC-46275	Sulprostone, MB-28767	ONO-AP-324
EP <sub>4</sub>	ONO-AE1-329, tetrazolo PGE <sub>1</sub>	(PGE <sub>2</sub> )	
IP	Cicaprost	AFP-07, iloprost	Octimibate Taprostone
TP	STA <sub>2</sub> , U-46619		CTA <sub>2</sub> , PTA <sub>2</sub> , U-44069
FP	Fluprostenol, lataprost-FA	Cloprostenol	AL-8810
DP <sub>1</sub>	BW-245C	BW A868C	BW-192C86

\* FA – free acid

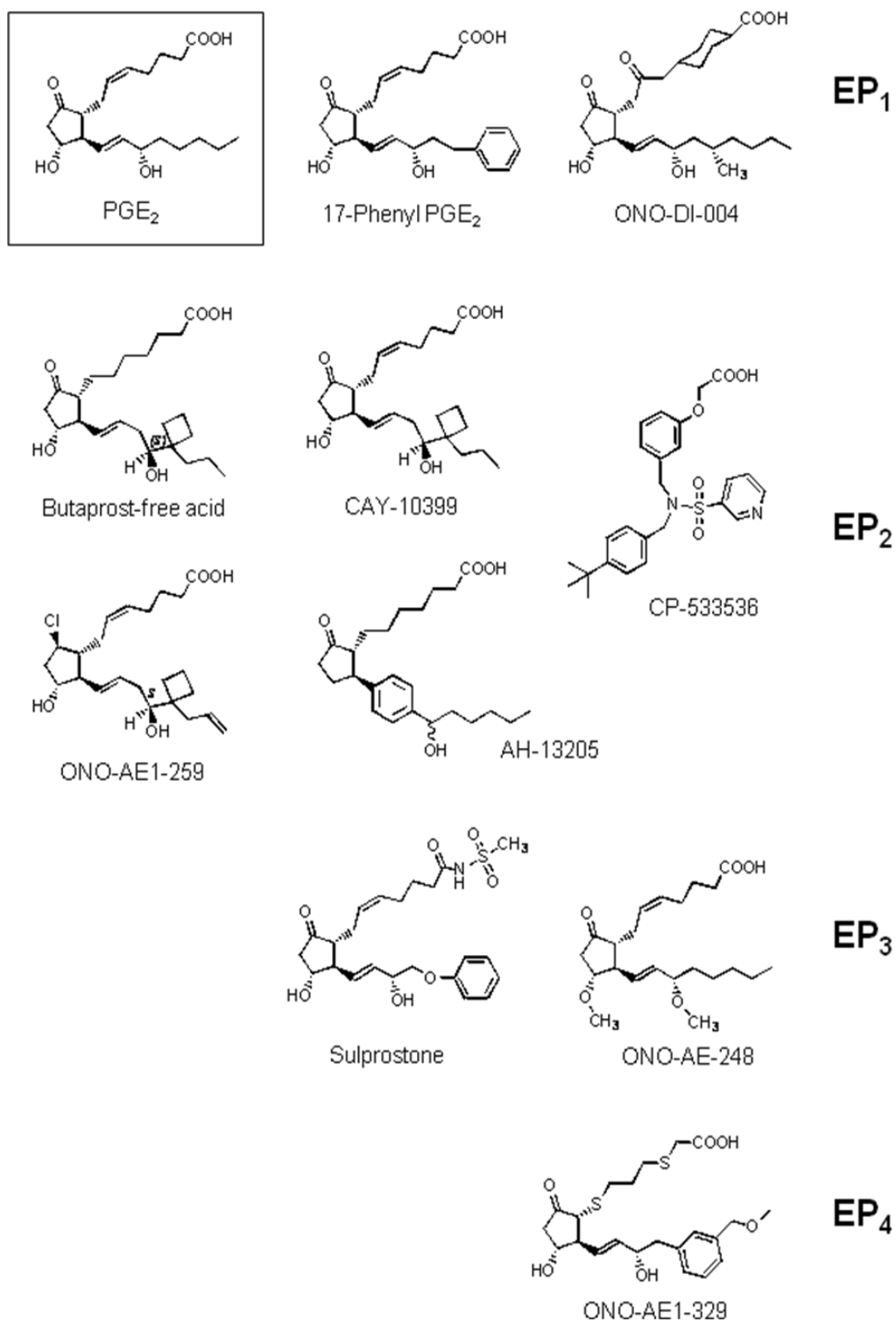


**Table 1.2**  $K_i$  values (nM) for agonists used in the current study

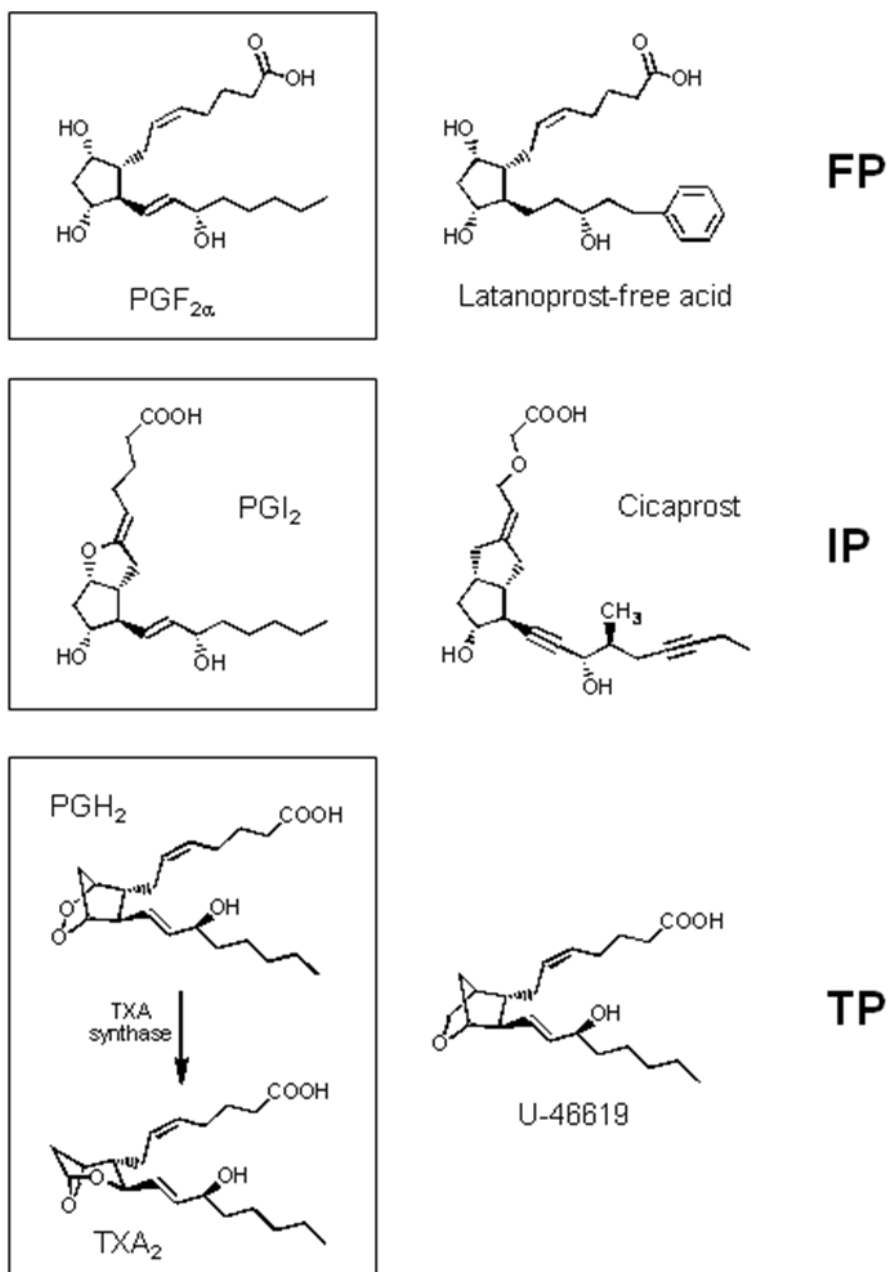
Ligand	EP <sub>1</sub>		EP <sub>2</sub>		EP <sub>3</sub>		EP <sub>4</sub>	
	Human	Mouse	Human	Mouse	Human	Mouse	Human	Mouse
U-46619	28,000	>10,000	12,000	>10,000	13,000	>10,000	2,000	>10,000
PGE <sub>2</sub>	9.1	20	4.9	12	0.33	0.85	0.79	1.9
17-Phenyl PGE <sub>2</sub>		14		>10,000		3.7		1,000
Sulprostone	110	21	>100,000	>10,000	0.35	0.60	7,700	>10,000
ONO-D1-004		150		>10,000		>10,000		>10,000
ONO-AE1-259		>10,000		3		>10,000		6000
Butaprost-FA	28,000		91		1,600		19,000	
AH-13205		>10,000		240		82		>10,000
CP-533535				50				>3,200
CAY-10399		>10,000		92		>10,000		>10,000
ONO-AE-248		>10,000		3,700		7.5		4,200
PGD <sub>2</sub>	5,800	10,000	3,000	>10,000	420	280	1,500	>10,000
Cicaprost	>1,300	>10,000	>1,300	1,300	260	170	44	>10,000
Iloprost	11	21	1,900	1,600	56	22	280	2,300
PGF <sub>2α</sub>	550	1,300	960	>10,000	38	75	290	>10,000
Latanoprost-FA	1,800		40,000		6,500		>100,000	

Ligand	DP		FP		IP		TP	
	Human	Mouse	Human	Mouse	Human	Mouse	Human	Mouse
U-46619	4,000	>10,000	240	1,000	57,000	>10,000	35	67
PGE <sub>2</sub>	310	>10,000	120	100	>10,000	>10,000	29,000	>10,000
17-Phenyl PGE <sub>2</sub>		>10,000		60		>10,000		>10,000
Sulprostone	>100,000	>10,000	200	580	>100,000	>10,000	>100,000	>10,000
ONO-D1-004								
ONO-AE1-259								
Butaprost-FA	12,000		>100,000		55,000		20,000	
AH-13205		>10,000		>10,000		>10,000		>10,000
CP-533535								
CAY-10399								
ONO-AE-248								
PGD <sub>2</sub>	1.7	21	6.7	47	>100,000	>10,000	6,600	>10,000
Cicaprost	>1,300	>10,000	>1,300	>10,000	17	10	>1,300	>10,000
Iloprost	1,000	>10,000	620	>10,000	11	11	6,500	>10,000
PGF <sub>2α</sub>	860	>10,000	3.2	3.4	>100,000	>10,000	8,700	>10,000
Latanoprost-FA	55,000		2.8		>100,000		16,000	

\* Abramovitz *et al.*, (1994); Kiriyama *et al.*, (1997); Suzawa *et al.*, (2000); Tani *et al.*, (2001); Amano *et al.*, (2003)



**Figure 1.3** Structures of agonists showing selectivity for EP receptor subtypes. The natural agonist PGE<sub>2</sub> is shown in the box. CP-533536 has a non-prostanoid structure.



**Figure 1.4** Structures of agonists showing selectivity for FP, IP and TP receptors. The natural agonists are shown in the boxes; both PGH<sub>2</sub> and TXA<sub>2</sub> activate the TP receptor.

## 1.4.2 Selective antagonists

Table 1.3 lists selective prostanoid antagonists that were available for the current studies. Figure 1.5 shows the chemical structure of the selective prostanoid antagonists used in the current study.

Several EP<sub>1</sub> antagonists have been developed. SC-51322 was chosen as the selective EP<sub>1</sub> antagonist in the current study. It antagonised PGE<sub>2</sub> with pA<sub>2</sub> value of 8.1 in guinea-pig ileum assay (Hallinan *et al.*, 1994) and pA<sub>2</sub> of 8.45 in guinea-pig trachea (Hung *et al.*, 2006). A recent EP<sub>1</sub> antagonist developed by the GlaxoSmithKline research group, GW-848687 has been shown to be a competitive antagonist at EP<sub>1</sub> receptor with pA<sub>2</sub> of 9.1 in a recombinant receptor assay (Giblin *et al.*, 2007).

Selective EP<sub>2</sub> antagonists are essentially not available for the current antagonist study.

L-798106 showed high selectivity for EP<sub>3</sub> receptors based on ligand binding assays involving recombinant prostanoid receptors (Juteau *et al.*, 2001). In functional studies, it blocked the action of sulprostone with pA<sub>2</sub> values of 7.82 in guinea-pig trachea (Clarke *et al.*, 2004) and 7.43 – 8.03 in rat femoral artery (Hung *et al.*, 2006) (Table 1.2). L-798106 (known as CM9) has been shown to inhibit PGE<sub>2</sub>-induced Ca<sup>2+</sup> entry in a rat recombinant EP<sub>3</sub> receptor assay, with pK<sub>i</sub> value of 7.12 (Jugus *et al.*, 2009). L-826266 is a chloro-analogue of L-798106. In guinea-pig aorta, L-826266 blocked EP<sub>3</sub> agonist-induced contraction with pA<sub>2</sub> value of 7.58 (Jones *et al.*, 2008). Both antagonists had a slow onset of block. This may relate to their high lipophilicity with predicted *n*-octanol / water partition coefficients (ClogP) of 6.9 and 7.4 respectively (ChemAxon freeware). ONO-AE3-240 is reported to be a highly selective EP<sub>3</sub> antagonist (mouse EP<sub>3</sub> K<sub>i</sub> = 0.23 nM; Amano *et al.*, 2003), but its structure has not been disclosed.

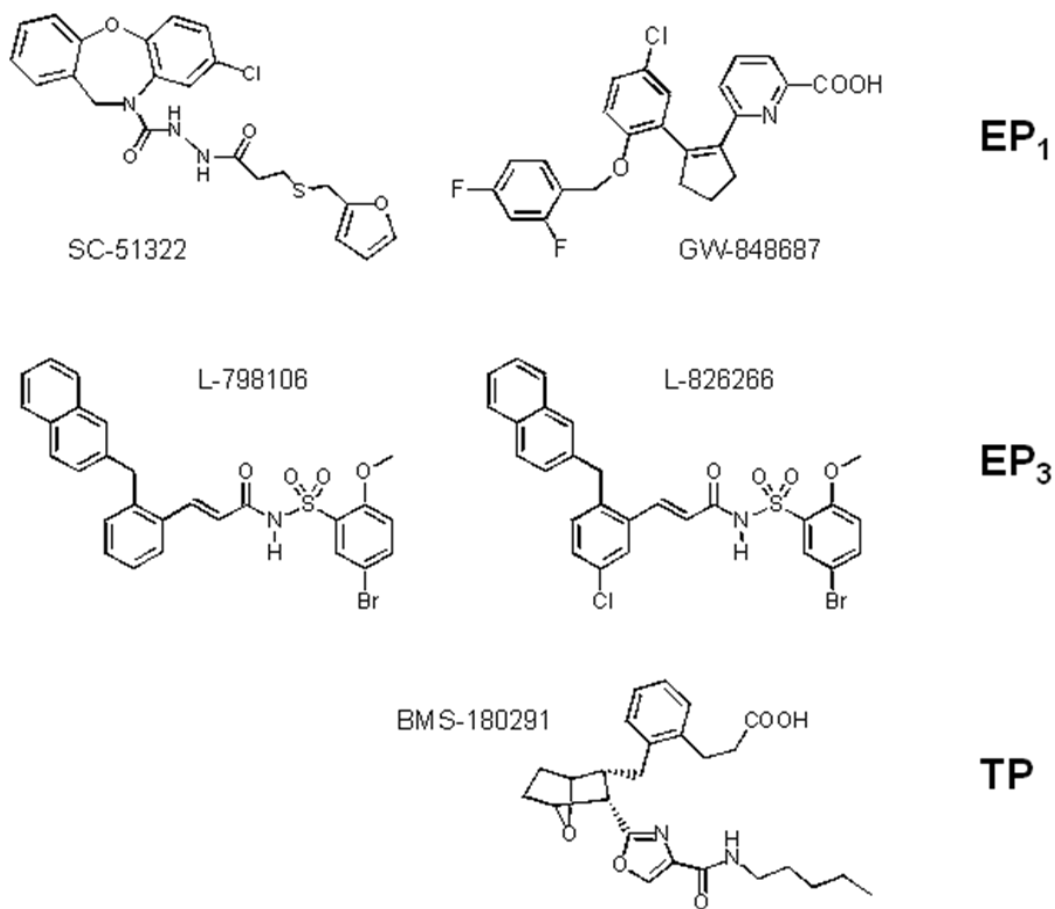
The TP antagonist available in the current study is BMS 180291 with pA<sub>2</sub> value between 9.2 – 10.0 (Zhang *et al.*, 1996). In the low nanomolar range, BMS-180291 has been reported slowly to reach a steady state, requiring up to 2 h contact in guinea-pig aorta (Jones *et al.*, 2008).

Selective IP receptor antagonists have been documented, but were not available in for the current study. RO-1138452 and RO-3244794 have been shown to selectively bind to human platelet IP receptor and to recombinant IP receptor system (Bley *et al.*, 2006).

Several selective FP receptor antagonists have been studied before, mainly for their effects on myometrium contraction. For example, the non-prostanoid AS-604872 inhibited spontaneous uterine contractions in pregnant rat near term (Cirillo *et al.*, 2007). It has K<sub>i</sub> values of 35, 158 and 323 nM for human, rat and mouse rc-FP receptors. Its selectivity for EP<sub>2</sub> receptor was 20-fold and more than 300-fold for other prostanoid receptors. However, no selective FP receptor antagonists were available for the current study.

**Table 1.3** Antagonists chosen for use in the current study

<b>Prostanoid receptor</b>	<b>Antagonists</b>	<b>Reported pA<sub>2</sub></b>	<b>References</b>
EP <sub>1</sub>	SC-51322	8.45	Hung <i>et al.</i> , 2006
	GW-848687	9.1	Giblin <i>et al.</i> , 2007
EP <sub>3</sub>	L-798106	7.12 – 8.03	Juteau <i>et al.</i> , 2001 Clarke <i>et al.</i> , 2004 Hung <i>et al.</i> , 2006 Jugus <i>et al.</i> , 2009
	L-826266	7.58 - 8.35	Clark <i>et al.</i> , 2008 Jones <i>et al.</i> , 2008
TP	BMS-180291	9.5	Zhang <i>et al.</i> , 1996



**Figure 1.5** Structures of antagonists for EP<sub>1</sub>, EP<sub>3</sub> and TP receptors.

## 1.5 Physiological role of prostanoids in the human body

Prostanoids are presumed to play important roles in a variety of physiological and pathophysiological processes in the body. When induced pathologically, prostaglandins can assume harmful or protective roles. These roles have been studied by use of the knockout (KO) mice. Mice deficient in each prostanoid receptor have been generated by the disruption of each receptor gene by homologous recombination. In the sections below, these roles will be discussed as they relate to the *in vitro* systems studied here.

### 1.5.1 Heart

Acute myocardial infarction (AMI) is still a leading cause of death in developed countries, and it is usually caused by acute thrombotic occlusion of a coronary artery that has become atherosclerotic. The underlying pathophysiology of AMI is ischemia-reperfusion (I/R) injury of the heart. PGI<sub>2</sub> and its analogues have been reported to attenuate cardiac I/R injury when used exogenously *in vivo* (Narumiya *et al.*, 1999). The proposed mechanisms of protection were because of their inhibitory effects on platelets and neutrophils and ill-defined membrane-stabilizing effects. TXA<sub>2</sub> synthase inhibitors and / or TP antagonists have been reported to reduce myocardial infarct size in animal studies *in vivo* (Xiao *et al.*, 2004). The effects of TXA<sub>2</sub> synthase inhibitors were attributed to enhanced generation of PGI<sub>2</sub> derived from increased availability of its precursor, resulting in inhibition of neutrophil adhesion to endothelial cells. The cardioprotective effects of TP antagonists, however, are variable (Xiao *et al.*, 2001).

Selective COX-2 inhibitors have replaced traditional NSAIDs for many inflammatory conditions. However, the withdrawal of certain COX-2 inhibitors due to increased likelihood of hypertension, myocardial infarction and stroke has cast a shadow over this therapy (Bombardier



*et al.*, 2000; Silverstein *et al.*, 2000; Mukherjee *et al.*, 2001). By inhibition of COX-2, biosynthesis of endogenous PGI<sub>2</sub> is reduced. Hence, the homeostatic response to accelerated platelet-vascular interactions, as in severe atherosclerosis and unstable angina, is abolished (Cheng *et al.*, 2002).

In a study, using IP-KO and TP-KO mice, it was shown that IP receptor deficiency significantly aggravates cardiac I/R injury *in vivo* (Xiao *et al.*, 2004). The IP-KO heart had greater I/R injury when assessed functionally and biochemically. However, the study showed that lack of TP receptors did not significantly alter either the myocardial infarct size *in vivo* or the degree of I/R injury *ex vivo*. This clearly demonstrated that TXA<sub>2</sub> has a small role, if any, in cardiac I/R injury.

It has been shown that up-regulation of COX-2 and increases in PGE<sub>2</sub> synthesis occur in the heart during I/R injury. This result suggested that PGE<sub>2</sub> plays some role in cardiac I/R injury (Calabresi *et al.*, 2003). Another study used EP<sub>4</sub>-KO mice to provide direct evidence that EP<sub>4</sub> deficiency significantly aggravates cardiac I/R injury *in vivo* (Xiao *et al.*, 2004). The result showed that endogenous PGE<sub>2</sub> is able to attenuate cardiac I/R injury and that the effect of PGE<sub>2</sub> is mediated by EP<sub>4</sub> receptor. Because there is significant production of PGE<sub>2</sub> during I/R injury and abundant expression of the EP<sub>4</sub> mRNA in the heart, PGE<sub>2</sub> should exert its cardioprotective action by acting directly on the cardiac tissue. Application of an EP<sub>4</sub> agonist significantly reduced the infarct size, thereby providing additional evidence for the involvement of EP<sub>4</sub> receptors.

## **1.5.2 Thrombosis and haemostasis**

It is now understood that atherosclerosis is an inflammation in the intima of large arteries that is triggered by high serum cholesterol and by the complex array interaction of various types of cells

including monocytes / macrophages, endothelial cells (ECs), smooth muscle cells (SMCs), T cells, and blood platelets (Ross, 1999). A variety of substances including cytokines, chemokines, and growth factors are suggested to induce, amplify, and modify this inflammatory process. Involvement of prostanoids in acute inflammation has been well documented based on the finding that aspirin-like NSAIDs are specific COX inhibitors. TXA<sub>2</sub> and PGI<sub>2</sub> are two major prostanoids in the cardiovascular system, being abundantly produced by blood platelets and vascular endothelium, respectively. Previous studies found that TXA<sub>2</sub> and PGI<sub>2</sub> biosynthesis is increased in patients with atherosclerosis (Belton *et al.*, 2000). Urinary levels of TXA<sub>2</sub> and PGI<sub>2</sub> metabolites were elevated in patients with atherosclerosis compared with normal patients. A study by Murata *et al.*(1997) showed that PGI<sub>2</sub> and TXA<sub>2</sub> do not work constitutively in regulation of the systemic circulation, but more likely work on demand in response to local stimuli. The presence of EP<sub>4</sub> receptors on macrophages in atheromatous plaques from human coronary and carotid arteries has been established by immunohistochemistry (Takayama *et al.*, 2002).

Earlier study on human platelets showed that sulprostone enhanced platelet aggregation *in vitro* even at low concentration (4 – 400 nM) (Matthews and Jones, 1993). PGE<sub>2</sub> and the selective EP<sub>3</sub> receptor agonist, AE-248 were shown to potentiate the platelet aggregation induced by U-46619 in WT mice (Ma *et al.*, 2001). In EP<sub>3</sub>-deficient mice, both PGE<sub>2</sub> and AE-248 lost this action completely. These results clearly show that the potentiating effect of PGE<sub>2</sub> on platelet aggregation was mediated by EP<sub>3</sub> receptor. In a recent study, DG-041 an EP<sub>3</sub> receptor antagonist has been demonstrated to antagonise the effects of sulprostone on platelet function (Heptinstall *et al.*, 2008). It may be useful in preventing platelet build-up in atherosclerotic lesions.

A study on IP-deficient mice showed that an enhanced thrombotic tendency was observed when endothelial damage was induced (Kobayashi *et al.*, 2004). This finding is consistent with the proposed role of PGI<sub>2</sub> as an endogenous antithrombotic agent. This antithrombotic system is activated in response to vascular injury to minimize its effects (Belton *et al.*, 2000). TP receptor activity has also been shown to have an important role in platelet homeostasis. Activation of TP

receptor lead to platelet aggregation (Arita *et al.*, 1989). Bleeding times were prolonged in TP-KO mice as compared to WT mice (Thomas *et al.*, 1998). Similarly, administration of U-46619 induced a rapid, irreversible platelet aggregation in WT mice but no detectable aggregation of platelets in TP-KO mice.

The balancing act between the opposing effects of PGI<sub>2</sub> and TXA<sub>2</sub> is important to vascular homeostasis. This was demonstrated in the study using apoE-KO mice with deletion of TP or IP receptors as well (Kobayashi *et al.*, 2004). Atherogenesis was significantly accelerated in apoE-KO / IP-KO mice compared with apoE-KO mice. In contrast, TP deficiency suppressed the extent of atherosclerosis. Deposition of human platelets onto damaged rabbit aorta *in vitro* is reduced in the presence of selective TP antagonist, GR-32191 which appears to inhibit aggregation of platelets (Hornby *et al.*, 1989). Similarly, the administration of the TP-receptor antagonist, S-18886 to rabbits strongly inhibited the atherogenic process in both uninjured and injured vessels (Worth *et al.*, 2005). These results suggest that the balance of the PGI<sub>2</sub> and TXA<sub>2</sub> is important for maintaining vascular homeostasis, prevent thrombosis and vasospasm while performing efficient homeostasis.

### **1.5.3 Hypertension**

PGE<sub>2</sub> elicits contractile and / or relaxant responses of vascular smooth muscles *in vitro*. A study in EP<sub>2</sub>-KO mice showed that intravenous infusion of PGE<sub>2</sub> and sulprostone (EP<sub>1/3</sub> agonist) induces hypertension (Kennedy *et al.*, 1999). When fed on a high-salt diet, the EP<sub>2</sub>-KO animals developed significant hypertension with concomitant increase in urinary excretion of PGE<sub>2</sub>. These results indicate that PGE<sub>2</sub> is produced in the body in response to a high-salt diet and work to negatively regulate the blood pressure via the relaxant EP<sub>2</sub> receptor. The dysfunction of this pathway may be involved in producing the salt-sensitive hypertension.

#### 1.5.4 Pulmonary vasculature

Pulmonary arterial hypertension (PAH) is a condition where there is a continuous high pressure in the pulmonary artery (Farber and Loscalzo, 2004). The average pressure in the normal situation is about 14 mmHg when the person is resting. In PAH, the average is usually greater than 25 mmHg at rest or 30 mmHg with exercise. Early in the disease, as the pulmonary artery pressure increases, thrombotic pulmonary arteriopathy occurs. As the pulmonary pressure continues to rise, plexogenic pulmonary arteriopathy develops. A remodelling of the pulmonary vasculature with intimal fibrosis and replacement of normal endothelial structure characterizes this.

In the human pulmonary arterial smooth muscle, IP and EP<sub>3</sub> receptors have been shown to be responsible for the prostanoid induced relaxation and contraction respectively (Qian *et al.*, 1994; Walch *et al.*, 1999). In PAH, the synthesis of PGI<sub>2</sub> is reduced, whereas the synthesis of TXA<sub>2</sub> is increased (Christman *et al.*, 1992). The relative deficiency of PGI<sub>2</sub> in PAH secondary to reduced PGI<sub>2</sub> synthase activity leads to vasoconstriction, proliferation, thrombosis, and inflammation in the affected vessels (Tuder *et al.*, 1999).

Long-term intravenous PGI<sub>2</sub> has become the most important specific therapy for PAH and associated diseases. However, this therapy is hampered by catheter complications and systemic side effects. Alternatively, inhalation of the PGI<sub>2</sub> analogues, iloprost (Olschewski *et al.*, 2001) and beraprost (Abe *et al.*, 2001) results in pulmonary vasodilation with few systemic side effects.

### 1.5.5 Kidney function

The role of PGI<sub>2</sub> in regulating renal and glomerular hemodynamics, renin secretion, as well as tubular transport processes has been documented. PGI<sub>2</sub> had a protective role in primary cultures of proximal tubule epithelial cells that were subjected to hypoxia and reoxygenation (Paller and Manivel, 1992). The idea that PGI<sub>2</sub> more than other prostanoids is relevant to the maintenance of renal function stem from various studies. A few studies showed that in recombinant mice lacking the IP receptor there was salt-sensitive hypertension and enhanced renin release following water deprivation (Fujino *et al.*, 2004).

Altered growth responses (proliferation and hypertrophy) contribute to changes in renal function characteristic of various nephropathies. A clear link between renal hypertrophy and changes in renal function has been established (Nasrallah and Hebert, 2005). Cell loss also plays an important role in renal disease progression. There are two distinct forms of cell death, apoptotic and necrotic. In many progressive renal diseases, the primary feature is glomerulosclerosis. Sclerotic glomerulosclerosis is characterized by progressive expansion of the extracellular matrix, which replaces glomerular cells. Apoptotic death in the kidneys is triggered by a disruption of matrix-cell interactions (Sugiyama *et al.*, 1996). Activation of IP receptor in the kidneys has been shown to prevent matrix-induced apoptosis. This prevents the accumulation of the matrix underlying progressive glomerular disease (Nasrallah and Hebert, 2005).

In the kidneys, all four known EP receptors are expressed in different regions (Jensen *et al.*, 1999; Jensen *et al.*, 2001). The natriuretic and diuretic effects of PGE<sub>2</sub> are due to interaction with EP<sub>1</sub> and EP<sub>3</sub> receptors situated on the ascending part of the loop of Henle and the collecting duct system. PGE<sub>2</sub> inhibits arginine vasopressin-stimulated water and sodium transport, thereby increasing water and sodium excretion from the kidneys. The EP<sub>2</sub> receptor is exclusively expressed in the rat kidneys medulla and EP<sub>4</sub> receptor mainly in the medulla. The EP<sub>2</sub> receptor

maintains renal blood flow associated with low-salt states. The EP<sub>4</sub> receptor in the other hand promotes salt excretion in response to a high salt intake (Jensen *et al.*, 1999).

### **1.5.6 Ductus arteriosus**

At birth, mammals including humans undergo a dramatic change in their circulation with the commencement of respiration, i.e., from the fetal circulation system, which shunts the blood flow from the main pulmonary artery directly to aorta via ductus arteriosus, to the pulmonary circulation system in the neonate. This adaptive change is caused by the closure of the ductus and is induced by the withdrawal of the dilator prostaglandins as well as active contraction exerted by increased oxygen tension (Smith, 1998). Elevated oxygen tension also inhibits PGI<sub>2</sub> synthase, which decreases the PGI<sub>2</sub> level in ductus, thus promoting contraction of the ductus arteriosus (Smith *et al.*, 1994).

A study using various synthetic PG analogues suggested that both IP and EP<sub>4</sub> receptors are present in the ductus and cause dilation of this vessel (Smith *et al.*, 1994). PGE<sub>2</sub> is the major prostaglandin that affects tone (Clyman *et al.*, 1980). The other prostanoids play smaller roles (Smith *et al.*, 1994; Smith, 1998). A study using knockout mice showed that disruption of the EP<sub>4</sub> receptor gene resulted in death within three days after birth, due to marked pulmonary congestion and heart failure (Segi *et al.*, 1998). These results suggest a critical role of the EP<sub>4</sub> system in the ductus and suggest that the compensatory mechanism maintains ductus patency not only in the fetal period but also after birth. No abnormality was detected in IP-KO mouse.

### 1.5.7 Intraocular pressure

Prostaglandins play a role in lowering intraocular pressure (IOP) especially in the case of glaucoma and ocular hypertension. Traditionally,  $\beta$ -blockers are used to lower IOP. FP receptors agonists are currently widely used because they are more effective and have fewer side effects than  $\beta$ -blockers. These agonists exert their effects through activation of FP receptor (Anthony *et al.*, 1998). Latanoprost did not lower IOP in the FP-KO mice (Crowston *et al.*, 2004). In contrast, IOP was reduced in the treated eye of the WT mice. In addition, the potent FP-agonist, tafluprost stimulated production of endogenous prostaglandins through activation of the FP receptor, which in turn act on the EP<sub>3</sub> receptor (Ota *et al.*, 2007). Sulprostone have been shown to decrease IOP in rabbit without any side-effect as compared to ocular irritation and transient increase in IOP caused by PGE<sub>1</sub> and PGE<sub>2</sub> (Waterbury *et al.*, 1990). Allergan Pharmaceuticals, a pharmaceutical company specializing in ophthalmic drugs has an interest in this IOP lowering effect through EP<sub>3</sub> receptors. Allergan Pharmaceuticals supplied novel prostanoid compounds including the selective EP<sub>3</sub> agonist ONO-AE-248 and the EP<sub>3</sub> antagonist L-826266 for use in the current studies.

The selective EP<sub>2</sub> agonists have been shown to have varies in effects on intraocular pressure (Woodward *et al.*, 1993). Butaprost and AH-13205 have a different mechanism from other EP<sub>2</sub> agonists on effect in lowering intraocular pressure (Woodward *et al.*, 1995; Nilsson *et al.*, 2006).

A novel selective EP<sub>4</sub> agonist, PF-04475270 (a pro-drug) has been shown to have an effect of lowering IOP in dogs (Prasanna *et al.*, 2009). The effect was sustainable for a period of time. Further work is in progress by the same author to elucidate the role of lowering IOP by EP<sub>4</sub> agonist in human eye. Recently, a selective EP<sub>2</sub> agonist butaprost has been shown to significantly blunts the detrimental influence of ischemia / reperfusion to the retina (Andrade da Costa *et al.*, 2009).

## **1.6 Diabetes**

### **1.6.1 Diabetes overview**

Diabetes mellitus is an endocrine disease characterized by the lack of insulin production (Type 1) or resistance of the target-organ to insulin (insulin resistance) (Type 2). In early stage of diabetes, the subject usually presents with increased passing of urine (polyuria) and increased water uptake (polydipsia). Weight loss, which was seen in the STZ-rat (see later), is due to an inability to utilise the carbohydrate due to lack of insulin. Instead, the body fat is utilised for the energy.

The effects of diabetes can be examined in various rat models. The Zucker fatty (ZF) rat (Tokuyama *et al.*, 1995) and Goto-Kakizaki (GK) rat (Goto *et al.*, 1976) are both developed by cross-breeding and manifest type 2 diabetes. A more convenient model is the streptozotocin (STZ) induced-diabetic rat (Junod *et al.*, 1969).

### **1.6.2 Diabetes and endothelium**

Endothelium dysfunction has been reported in diabetic people and experimental animals with diabetes (De Vriese *et al.*, 2000; Schofield *et al.*, 2002; Schalkwijk and Stehouwer, 2005). The dysfunction is caused by the high glucose content in tissues, which triggers a cascade of functional and structural alterations in vascular cells, leading to macrovascular and microvascular diseases (Schalkwijk and Stehouwer, 2005). Diabetes itself also promotes an inflammatory response (Schalkwijk and Stehouwer, 2005), with changes in the ultrastructure and



the function of endothelial cells (Sotnikova *et al.*, 2006). These changes are confined to the endothelial cells and do not affect the smooth muscle cells.

### **1.6.3 Diabetes and prostaglandins**

A hallmark of endothelial dysfunction in diabetes is alteration in the biosynthesis of prostaglandins and oxidative stress products and decreased release and / or bioavailability of nitric oxide (NO) (Tesfamariam *et al.*, 1989; Vanhoutte *et al.*, 2005). The hyperglycaemic state in diabetes alone is the stimulation for productions of prostaglandins (Tesfamariam *et al.*, 1990). In support of this view, there is production of vasoconstrictor prostaglandins other than TXA<sub>2</sub> in rat mesenteric arterial bed as shown by Peredo *et al.* (1999). Endothelial COX, which is upregulated in various pathological conditions including diabetes, is thought to be responsible (Tesfamariam *et al.*, 1989; Ge *et al.*, 1995; Shi *et al.*, 2006; Shi *et al.*, 2007b). In contrast, Peredo *et al.* (2001) showed impairment of prostanoid production in the mesenteric vascular bed of the diabetic rat.

The endogenous prostanoids enhanced by DM have been shown to have synergism with the  $\alpha_1$ -adrenoreceptor agonist, phenylephrine through the TP receptor system in the rat femoral artery (Shi *et al.* 2008). However, the possibility of other prostanoid receptor involvement in the rat femoral artery, in particular the EP<sub>3</sub> receptor has not been investigated. There was a pronounced synergism between the EP<sub>3</sub> system and other contraction, accompanied by pronounced synergy with other contractile systems that rely extensively on Ca<sup>2+</sup> influx through L-type Ca<sup>2+</sup> channels including phenylephrine (Hung *et al.*, 2006).

## 1.7 Pharmacological principles

### 1.7.1 Synergism and its measurement

Synergism occurs when the combination of two or more drugs produces a larger effect than the sum of the effects of each acting alone. This is in contrast to an additive effect, where the combined effect is simply the summation of the components.

In the prostanoid area, synergism between EP<sub>3</sub> and TP contractile systems was demonstrated in rat femoral artery (Hung *et al.*, 2006). On its own, PGE<sub>2</sub> induced a small contraction in the artery. However, strong contraction was seen when the preparation was primed with K<sup>+</sup>, phenylephrine or U-46619. Sulprostone also synergised with these strong contractile agents. U-46619 also has a synergistic interaction with adrenaline in human umbilical veins (Errasti *et al.*, 2007). U-46619 at 0.3 nM did not induce contractions but shifted the concentration–response curve for adrenaline to the left without modifying maximal contraction. In comparison, U-46619 at 3 nM increased the maximal response attained by sulprostone in rat femoral artery (Hung *et al.*, 2006). The selective TP receptor antagonist SQ-29548 blocked the potentiating effect of 0.3 nM U-46619.

PGE<sub>2</sub> at high concentration (> 10 μM) has a cross activity on TP receptor, based on binding studies (Dorn *et al.*, 1992). Similarly, other EP<sub>1/3</sub> agonists also show some TP agonism (Jones *et al.*, 1982; Errasti *et al.*, 2007). For this reason, a TP antagonist was routinely included in the bathing fluid in my studies to prevent potential synergism between the EP and TP contractile systems (see Methods chapter).

## 1.7.2 Types of antagonism and $pA_2$ measurement

A receptor antagonist does not provoke a biological response itself upon binding to a receptor, but blocks or reduces the agonist-mediated response. In competitive antagonism, the antagonist binds reversibly to (part of) the agonist binding domain without activating the effector mechanism (Neubig *et al.*, 2003). The antagonism is usually surmountable, that is, a maximum response can still be elicited by increasing the concentration of the agonist. Alternatively, agonist and antagonist can be simultaneously bound to different sites of the receptor; antagonist binding reduces or prevents the action of the agonist with or without any effect on the binding of the agonist. This antagonism is non-competitive; where no amount of agonist can completely overcome the inhibition once it has been established.

$pA_2$  is defined as the negative log of the concentration of the antagonist required to produce a two-fold shift to the right in the agonist-concentration-effect curve (Neubig *et al.*, 2003).  $pA_2$  is a measure of the affinity constant of an antagonist for the receptor. Binding constants (preferably  $pK_i$ ) are often given where functional information is not available.  $pA_2$  and  $pK_i$  values may not always agree: often the difference is due to processes affecting ligand concentration that are present in intact tissues, but not isolated cells or cell membranes.

The  $pA_2$  value of antagonist can be determined by several experimental protocols. The Schild protocol remains the gold standard for determining the  $pA_2$  of the antagonist. The Schild analysis requires the construction of full concentration-response curves (CRCs) for an agonist in the absence and in the presence of increasing concentrations of antagonist. For the Schild analysis to proceed the antagonism must be surmountable and the log agonist concentration-effect curve should be shifted to the right by the antagonist in a parallel fashion (Arunlakshana and Schild, 1959). The Schild protocol is based on this equation:

$$-\log K_b = \log (DR - 1) - \log B \quad (1)$$

in which DR (= A'/A) represents the ratio between equieffective agonist concentrations, respectively in the presence (A') and in the absence (A) of the concentration B of the competitive antagonist, and the  $K_b$  is the equilibrium dissociation constant. In the case of a competitive antagonist with a one-to-one relationship with the receptor, the regression of  $\log (DR - 1)$  on  $\log B$ , gives a straight line with slope (Schild slope) = 1 (e.g.: Figure 1.6). The  $pA_2$  is the value of  $x$ -intercept, which equals the value of  $-\log K_b$ . Clearly, when the Schild slope is different from unity, the evaluation of the dissociation constant is meaningless. Equation 1 can be re-arranged as follows:

$$-\log K_b = -\log \frac{B}{(DR - 1)} \quad (2)$$

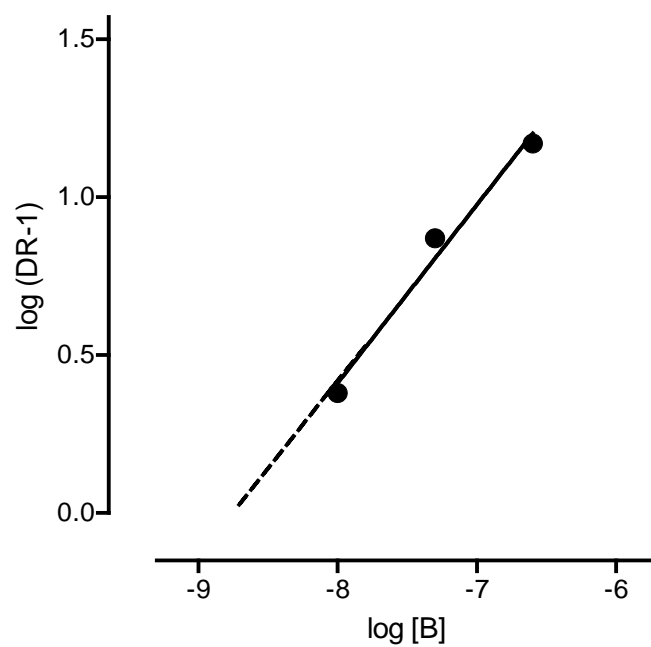
A negative / antilog transformation gives the original Gaddum equation:

$$K_b = B / (DR - 1) \quad (3)$$

Alternatively, the inhibition-curve (Cheng-Prusoff) protocol may be used. The Cheng-Prusoff protocol is derived from enzymatic system study, and describes the interaction between substrate, S and competitive inhibitor, I (Cheng and Prusoff, 1973). This interaction can be expressed by this equation:

$$K_i = \frac{IC_{50}}{1 + \frac{[S]}{K_m}} \quad (4)$$

where  $K_i$  is the antagonist dissociation constant, [S] is the fixed concentration of substrate and  $K_m$  is the dissociation constant of the substrate.  $IC_{50}$  is the concentration of the competitive inhibitor at half-maximal inhibition of the enzyme.



**Figure 1.6** An example of a Schild plot. The regression of  $\log (DR - 1)$  on  $\log B$  using simulated data, gives a straight line. The  $pA_2$  is the value of  $x$ -intercept, (8.73) which equals the value of  $-\log K_b$ .

Similarly, in ligand binding studies, the interaction of the ligand, L and the competitive inhibitor, I can be expressed by the related equation:

$$K_b = \frac{IC_{50}}{1 + \frac{[L]}{K_d}} \quad (5)$$

where  $K_b$  is the dissociation constant of the competitor,  $[L]$  is the fixed concentration of the (radio) ligand and  $K_d$  is the dissociation constant of the (radio) ligand.

Translating equation (5) to functional system gives the interaction between the agonist, A and the competitive antagonist as:

$$K_b = \frac{IC_{50}}{1 + \frac{[A]}{EC_{50}}} \quad (6)$$

where  $[A]$  is the fixed concentration of agonist,  $IC_{50}$  is the midpoint location parameter in the inhibition curve of the antagonist and  $EC_{50}$  is the midpoint location parameter in the agonist curve (Craig, 1993). However, this equation is only valid when the agonist CRC is rectangular hyperbolic in shape, which corresponds to a Hill slope of 1.0 for the logCRC.

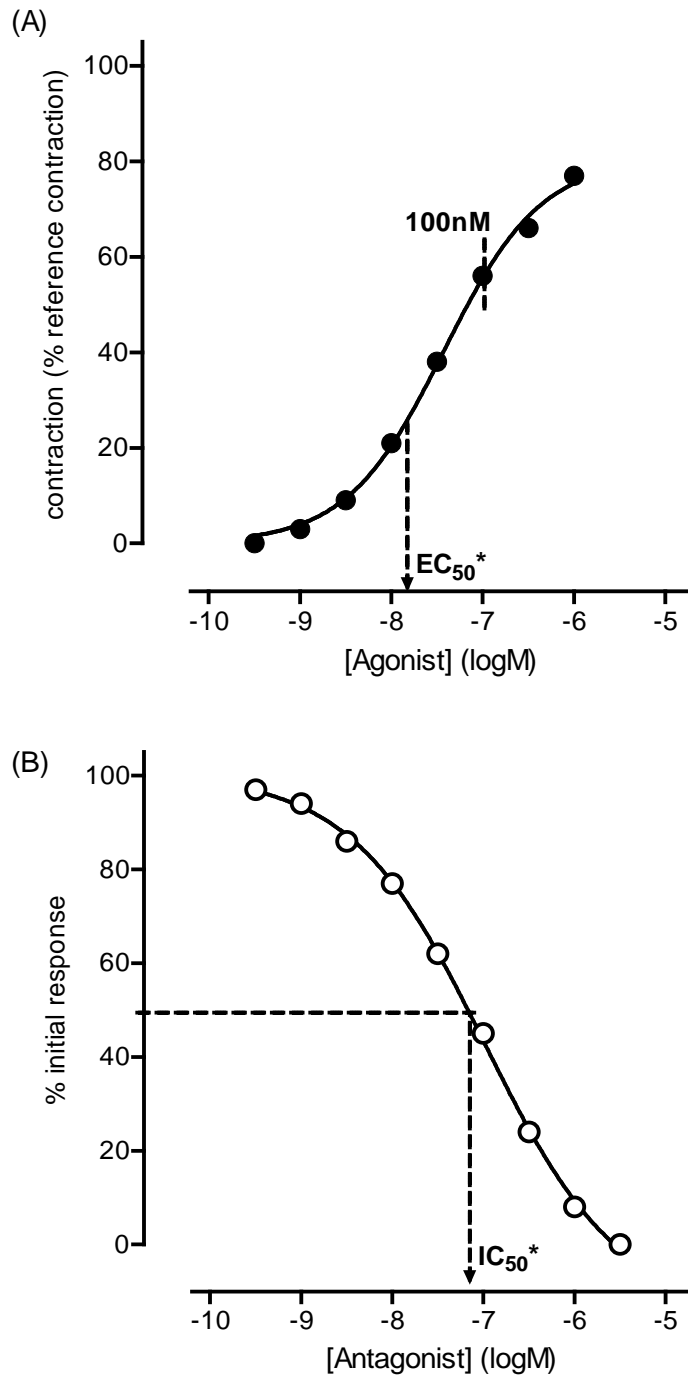
Commonly, the CRCs of the agonist in the majority of experiments tend to deviate from a rectangular hyperbola, thus  $n \neq 1$ . In the case of  $n \neq 1$ , it is necessary to use a modified form of the Cheng-Prusoff equation (Leff and Dougall, 1993):

$$K_b = \frac{IC_{50}}{\left[ 2 + \left[ \frac{[A]}{EC_{50}} \right]^n \right]^{1/n} - 1} \quad (7)$$

It has been suggested that the best method of analyzing the data from the inhibition-curve protocol is to simultaneously fit the antagonist inhibition curve and the agonist activation curve containing agonist concentrations up to the fixed concentration used in the inhibition curve. Both the agonist and antagonist concentration-effect curves constrained to have the same maximum. The dose-ratio (DR) is then calculated corresponding to the  $IC_{50}$  point on the inhibition curve (Lazareno and Birdsall, 1993a). The Gaddum-Schild equation now can be expressed as:

$$K_b = \frac{IC_{50}^*}{\frac{[A]}{EC_{50}^*} - 1} \quad (8)$$

where  $[A]$  is the fixed agonist concentration (example, at 100 nM). It should be noted that  $EC_{50}^*$  will not be the same as true  $EC_{50}$  unless the fixed agonist concentration itself produced the maximal response. The  $IC_{50}^*$  was estimated from an antagonist titration (inhibition curve) in the presence of 100 nM agonist (Figure 1.7B). The agonist concentration causing the same effect ( $EC_{50}^*$ ) is estimated from the agonist titration / CRC (Figure 1.7A).



**Figure 1.7** The example of basic inhibition-curve protocol used in the current experiments. (A) An agonist curve is first constructed from which the  $EC_{50}^*$  is obtained. (B) An antagonist inhibition curve is then obtained on the same preparation precontracted with submaximal agonist response (e.g. 100 nM agonist); the  $IC_{50}^*$  value is obtained.



Another method that was used in the current study, when the inhibition is incomplete is a ‘null method’ (Lazareno and Birdsall, 1993a). It makes no assumptions about the curves shapes. It is derived from the Gaddum equation (equation 3):

$$K_b = \frac{[B_i]}{\frac{[A_f] - 1}{[A_i]}} \quad (9)$$

At any level of effect, a horizontal line joining the agonist and antagonist curves defines two concentrations, [A] and [B], i.e. [A<sub>f</sub>] and [B<sub>i</sub>] such that [A<sub>f</sub>] alone has the same effect as [A<sub>i</sub>] in the presence of [B<sub>i</sub>]; so DR in the Gaddum equation above becomes [A<sub>f</sub>]/[A<sub>i</sub>] (Figure 1.8).

The inhibition-curve protocol and null method were preferred in the current study due to limited availability of the agonists and antagonists, as both are operate over a lower agonist concentration range than Schild protocol. It may also be easier to identify whether two receptors contribute to the measured response using the inhibition-curve protocol.

The inhibition-curve protocol has seen limited use in previous studies of prostanoid antagonists. In a recent study, the properties of two novel IP antagonists, RO1138452 and RO3244794 were studied at the humans rc-IP receptor (Bley *et al.*, 2006). IP receptor antagonism was assessed by the ability of both compounds to block cAMP accumulation stimulated with a fixed-concentration of carbaprostacyclin (cPGI<sub>2</sub>), a stable PGI<sub>2</sub> analogue. The results were consistent with selective antagonism on IP receptor, but the authors failed to use the modified form of the Cheng-Prusoff equation as described above.

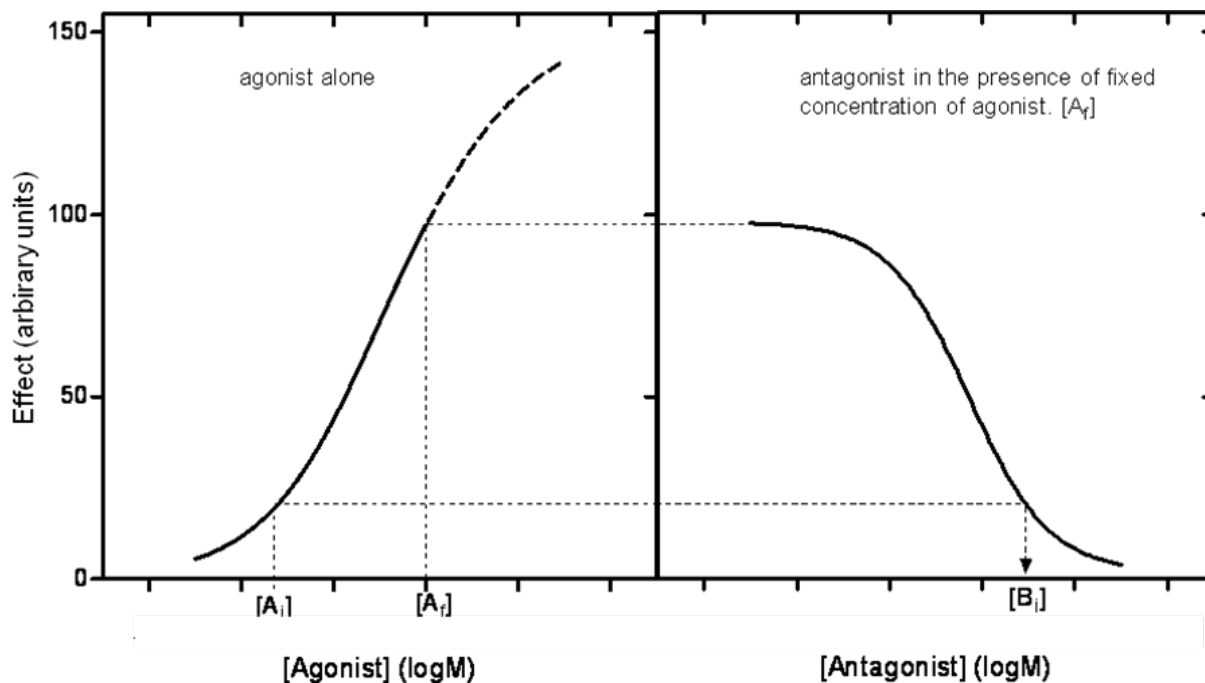


Figure 1.8 A null method for analysing antagonist inhibition curves. The dashed portion of the agonist concentration effect curve indicates data with agonist concentrations greater than the fixed agonist concentration  $[A_f]$  used in the inhibition curve and do not contribute to the analysis. At any level of effect, a horizontal line joining the agonist and antagonist curves defines two concentrations,  $[A]$  and  $[B]$ , i.e.  $[A_i]$  and  $[B_i]$  such that  $[A_f]$  alone has the same effect as  $[A_i]$  in the presence of  $[B_i]$ ; so DR in the Gaddum equation above becomes  $[A_f]/[A_i]$  (redrawn from Lazareno and Birdsall, 1993a).

## 1.8 Project aims

The main aim of the thesis was to use established and recently developed ligands to explore the nature of prostanoid receptor systems in several biological situations. These included:

1. The relaxant EP<sub>2</sub> system in guinea-pig trachea, with view to explaining why certain validated EP<sub>2</sub> agonists fail to produce their expected activities.
2. The characterization of contractile EP<sub>1</sub> and EP<sub>3</sub> receptors in vascular smooth muscle preparations where these receptors are both present. Published studies on this theme have serious deficiencies where the presence of both receptors has been proposed. It was also considered useful to identify preparations where prostanoid ligands had a rapid onset and offset of action; on-going studies by one of my supervisors had been blighted by the long incubation periods required for some ligands to reach steady-state receptor occupancy.
3. The nature of prostanoid receptors in the mesenteric artery of the rat, in view of the physiological control of the artery being similar to the human counterpart. This preparation may be a good model for pharmacological intervention studies.
4. The nature of contractile prostanoid receptors in the urinary bladder of the rat, in view of the current use of this animal to screen prostanoid ligands for potential as anti-incontinence activity.

5. The role of endogenous prostanoids activating contractile EP<sub>3</sub> and TP receptor systems in controlling vascular sensitivity in the rat streptozotocin model of diabetes. Previous studies on the TP system in the isolated femoral artery from the streptozotocin rat have failed to take into account the possibility of pronounced synergism with endogenous PGE<sub>2</sub> acting via the EP<sub>3</sub> receptor.

A particular focus of the work was the pharmacological utility of novel selective agonists and antagonists for EP<sub>1</sub> and EP<sub>3</sub> receptors. In studying the antagonists, inhibition-curve protocols were explored as an efficient means of establishing the onset kinetics and affinity of each antagonist, and of determining how many receptor types contribute to the agonist action. There has been little use of this type of protocol in the prostanoid receptor area.

**CHAPTER TWO**

**GENERAL METHODS**

**AND**

**MATERIALS**

## **2.1 Tissue isolation**

### **2.1.1 Guinea-pig trachea**

Male Dunkin-Hartley guinea pigs (400 - 450 g) were killed by CO<sub>2</sub> inhalation. The cervical trachea identified, removed immediately and dissected. The dissected trachea was pinned down on a Sylgard coated block, immersed in Krebs' solution and cleaned free of surrounding connective and fatty tissue. The trachea was cut into two rings 3 mm wide.

### **2.1.2 Rat mesenteric artery**

Male Sprague-Dawley rats (250 - 350 g) were killed by cervical dislocation. The abdomen was dissected open and the proximal part of the superior mesenteric artery was isolated from the abdominal aorta and tied off. The superior mesenteric artery was dissected out from the surrounding connective tissue. The artery was immersed in Krebs' solution and pinned down on a Sylgard coated block and cleaned free of surrounding fat and connective tissue. Care was taken not to stretch the artery, or to damage the endothelium. The artery was either used the same day or kept overnight at 4° C in Krebs solution (viability as demonstrated by McIntyre *et al.*, 1998). Four ring segments, 3 mm in length, were prepared from each rat.

### **2.1.3 Rat urinary bladder**

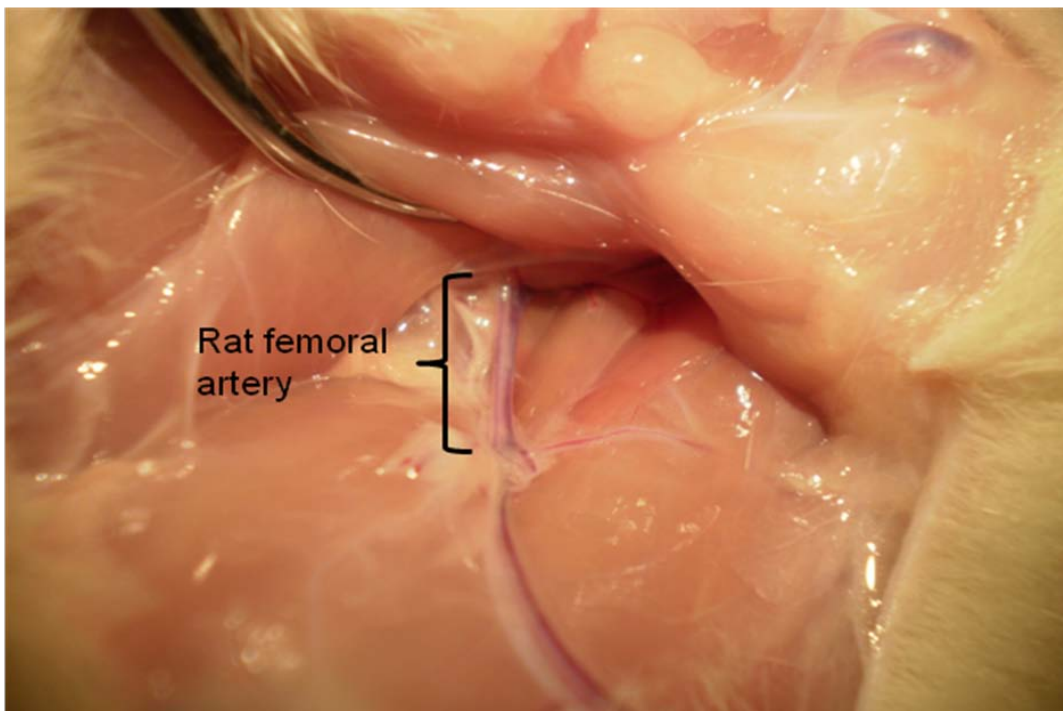
Male Sprague-Dawley rats (12 – 14 weeks old, 300 - 400 g) were killed by cervical dislocation and bleeding. The lower abdomen was dissected open, and the urinary bladder was isolated free from the surrounding structures and connective tissue. The whole bladder was excised and transported in oxygenated standard Krebs solution. The bladder was pinned down on a Sylgard coated block, immersed in Krebs' solution, and cleaned of connective tissue (Figure 2.1). The bladder was opened along the midline, and cut into longitudinal strips (7 mm long and 1.5 mm wide) (Maggi *et al.*, 1988; Schneider *et al.*, 2004). The bladder muscle strips were used immediately.

### **2.1.4 Rat femoral artery**

Right and left femoral arteries were obtained from 10 - 12 week old normal or diabetic, male Sprague-Dawley rats (200 - 250 g). The inguinal region was dissected open and the upper section of the femoral artery was isolated and tied off (Figure 2.2). The femoral artery was then gently freed from surrounding tissue by blunt dissection down to first branch of the femoral artery, kept in standard Krebs' solution, and pinned out on a Sylgard coated block (Figure 2.3). After cleaning free of surrounding fat and connective tissue, the artery was cut into rings, measuring 1 - 1.5 mm in length. Artery sections were used immediately. When necessary, the endothelium was removed mechanically from the artery by rolling the intimal surface against a piece of roughened wire slightly thinner than the lumen of the artery. This method has been described in the study of acetylcholine's mechanism of action in cat cerebral artery (Lee, 1982). The removal of endothelium was confirmed by the inability of acetylcholine to relax the artery precontracted with noradrenaline (Furchgott and Zawadzki, 1980; Furchgott, 1983) (See section 6.3.2 for the current experiment).



**Figure 2.1** Rat urinary bladder after dissection on Sylgard plate.

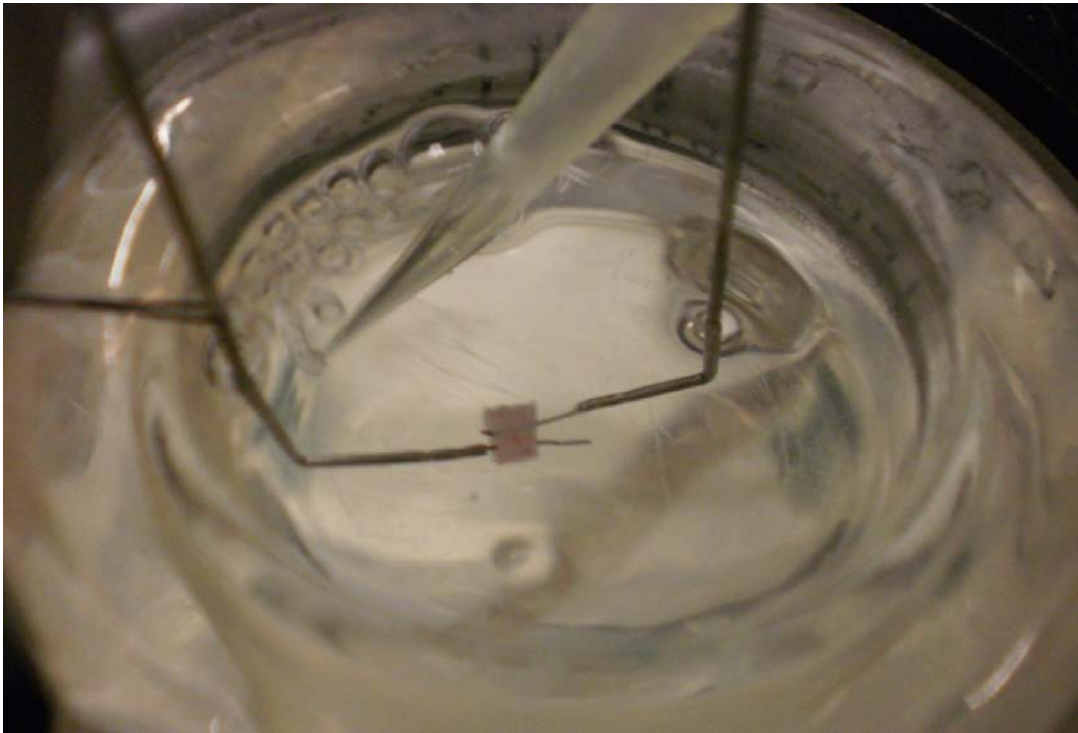


**Figure 2.2** Rat femoral artery *in-situ*.





**Figure 2.3** Rat femoral artery post dissection on Sylgard plate.



**Figure 2.4** Guinea-pig trachea mounted in 10 ml organ bath.

## **2.2 Wire myograph**

### **2.2.1 Krebs solution**

Krebs' solution was prepared fresh on the day of the experiment. The constitution was NaCl 118.4; NaHCO<sub>3</sub> 25.0; glucose 11.1; KCl 4.7; MgSO<sub>4</sub> 1.2; CaCl<sub>2</sub> 2.5 mM, bubbled with 95% O<sub>2</sub> and 5% CO<sub>2</sub> to maintain the pH at 7.4. Where indicated, indomethacin 1 μM was added to bath solution to prevent any influences from endogenous prostanoids (Buus *et al.*, 2000; Chauhan *et al.*, 2003).

### **2.2.2 Organ bath**

#### **2.2.2.1 Guinea-pig trachea**

The trachea ring was suspended between two intraluminal stainless steel hooks in Krebs-Henseleit solution aerated with 95% O<sub>2</sub> / 5% CO<sub>2</sub> (pH 7.4) in a conventional organ bath (10 ml) (Figure 2.4). The temperature of the Krebs was thermostatically maintained at 37 °C using continuous water circulation. Indomethacin 1 μM was added to the Krebs solution to block any influence from endogenous prostaglandin released during the course of the experiment (Undem and Adams, 1988). One hook was fixed while the other was connected to a Grass FT03 isometric force transducer, which in turn was connected to an ADInstruments PowerLab bridge amplifier / digitizer (Chart 5.23 software, sampling rate 400/s). Signals were saved to a Dell laptop computer. Tension was set at 1.5 g, determined from the preliminary experiments as an optimal resting tension, and within range of other published studies (McKenniff *et al.*, 1988; Undem and Adams, 1988; Schlemper *et al.*, 2005). The tracheal rings were allowed to equilibrate for 1 h

with repeated adjustment to ensure the 1.5 g resting tone was maintained. The ring was contracted with increasing concentrations of potassium chloride (KCl) (20, 40 and 60 mM) at intervals of 10 min to indicate tissue viability, and was then washed several times until baseline tone was restored. Any arterial rings not contracted by more than 0.5 g (representing 50% of normal 40 mM KCl response) were excluded from the study. Washout was done manually. The preparations were allowed a further 30 - 40 min to equilibrate. In the contractile studies, the prostanoid agonists were added cumulatively to construct concentration-response curves (CRC). After washout, the preparations were contracted with the agonist to 80% of maximal response ( $EC_{80}$ ), with the addition of the antagonist once the contraction had stabilised. In the relaxant sequences, histamine was used as the pre-contractile agent. Following pre-contraction, a cumulative dose-response to  $PGE_2$  or other prostanoids ( $EP_2$  agonists) was constructed in the presence of the  $EP_1$  receptor antagonist, SC-51322 (1  $\mu$ M) and the TP receptor antagonist, BMS-180291 (100 nM).

### **2.2.2.2 Rat mesenteric artery**

Arterial rings were suspended horizontally between two parallel stainless steel prongs in a Danish Myo Technology Model 610 M multichannel myograph for isometric force measurement in an organ bath. The tissue baths (10 ml) contained Krebs-Henseleit solution at 37 °C aerated with 95%  $O_2$  / 5%  $CO_2$  (pH 7.4); washout was done by draining through automatic suction and replacement of bathing solution from a 10 ml pipette. Indomethacin 1  $\mu$ M was added to bath solution to prevent any influences from endogenous prostanoids as demonstrated in previous studies (Buus *et al.*, 2000; Chauhan *et al.*, 2003). Changes in isometric force were recorded on a Dell laptop computer by use of Chart program (version 5.5.0) and a PowerLab data acquisition system (ADInstruments Ltd, Oxfordshire, UK). The resting tone was set at 1.0 g, determined by preliminary experiment and documented in other studies as optimal resting tension (Masset *et al.*, 1998). Followed a period of equilibration, the arterial rings were contracted by 40 mM of KCl to assess their viability. Any arterial rings not contracted by more than 0.5 g (representing 50% of 40 mM normal KCl response) were excluded from the study. The vessel rings were then

exposed to 10 nM PGE<sub>2</sub>, followed by washout. A concentration-response curve was constructed for each prostanoid agonists to determine the maximal response and EC<sub>50</sub> value. For inhibition curve studies, the arterial rings were precontracted with agonist (at 80% of maximal response; EC<sub>80</sub>), followed by construction of an inhibition curve for the antagonist. Selective prostanoid receptor antagonists used in the current study included SC-51322 (EP<sub>1</sub>), BMS-180291 (TP), and L-798106 (EP<sub>3</sub>). In agonist-induced relaxation studies, the preparation was precontracted with phenylephrine followed by a cumulative addition of particular agonist was constructed.

### **2.2.2.3 Rat urinary bladder**

Four bladder muscle strips were tied with cotton thread at each end and suspended in Krebs-Henseleit solution (containing 1 μM indomethacin) aerated with 95% O<sub>2</sub> / 5% CO<sub>2</sub> (pH 7.4) in a conventional organ bath (10 ml) and maintained at 37 °C. The tension signals were relayed from a Grass FT03 force transducer to an ADInstruments PowerLab bridge amplifier / digitizer (ADInstruments Ltd, Oxfordshire, UK) and recorded on a Dell laptop computer by use of Chart program (version 5.5.6). 1 g was set as the resting tension for this preparation, determined by preliminary experiments and has been documented in previous studies (Maggi *et al.*, 1988; Schneider *et al.*, 2004).

The bladder strips were allowed to equilibrate for 1 h, and were challenged twice with carbachol at 100 nM. After washout and an additional 30 min of equilibration, the preparations were exposed to 30 nM PGE<sub>2</sub> for 10 min. In contractile studies, a concentration-response curve was constructed by cumulative addition of the particular agonist. In successive sequences, the vehicle (serving as control) or antagonist at different concentrations was added at least 30 min before the cumulative addition of the agonist. For antagonist studies, the bladder strips were contracted with agonist, followed by construction of an inhibition curve for the antagonist.

#### **2.2.2.4 Rat femoral artery**

The arterial rings were mounted using the same technique outlined in Section 2.2.2.2. The resting tone was set at 1.0 g, determined in preliminary experiments. It has also been documented in previous studies (Shi *et al.*, 2007a; Shi *et al.*, 2007b). Reliable contractions to 60 mM KCl were used as an indication of tissue viability. The criteria of exclusion as described in Section 2.2.2.1 were applied. Phenylephrine was used as the precontractile agent. Where applicable, the antagonists used were preincubated for 30 min before cumulative addition of phenylephrine.

### **2.3 Streptozotocin-induced diabetes**

Diabetes was induced with a single dose of streptozotocin (STZ; 60 mgkg<sup>-1</sup>, i.p) after overnight fasting (Pfaffman *et al.*, 1982; Peredo *et al.*, 1999; Burke *et al.*, 2006). Control rats were injected with saline corresponding to the volumes of the STZ vehicle. 72 hours after STZ injection, a blood sample was taken from the tail artery and the fasting blood glucose level was measured using an Ascensia Breeze 2 glucometer (Bayer HealthCare, UK). The cut-off level for fasting blood glucose level to be considered as diabetic was 16 mmol<sup>-1</sup>. Any rat with blood level below 16 mmol<sup>-1</sup> was excluded from the study and sacrificed. Body weight was measured weekly and rats were kept under optimal conditions and given a normal diet. The rats were decapitated four weeks following the STZ injection. Blood glucose and body weight were measured just before the decapitation. Control rats with non-fasting blood glucose higher than 11.1 mmol<sup>-1</sup> were excluded from the study. All procedures conformed to Home Office Guidelines. Figure 2.5 summarizes the STZ-induced diabetes protocol in the current study.

## 2.4 Antagonist protocol and pA<sub>2</sub> estimation

In the current study, the inhibition-curve protocol was preferred to the Schild protocol due to limited availability of the agonists and antagonists and the shorter time period required (Lazareno and Birdsall, 1993b). Basically, the protocol involved initial construction of a concentration-response curve for the agonist. After washout, the preparation was submaximally pre-contracted with agonist (usually at 80% of maximal agonist response, EC<sub>80</sub>) followed by cumulative addition of the antagonist (Figure 2.6). The experimental data were then fitted separately to construct the antagonist inhibition curve and the agonist activation curve containing agonist concentrations up to the fixed concentration used in the inhibition curve.

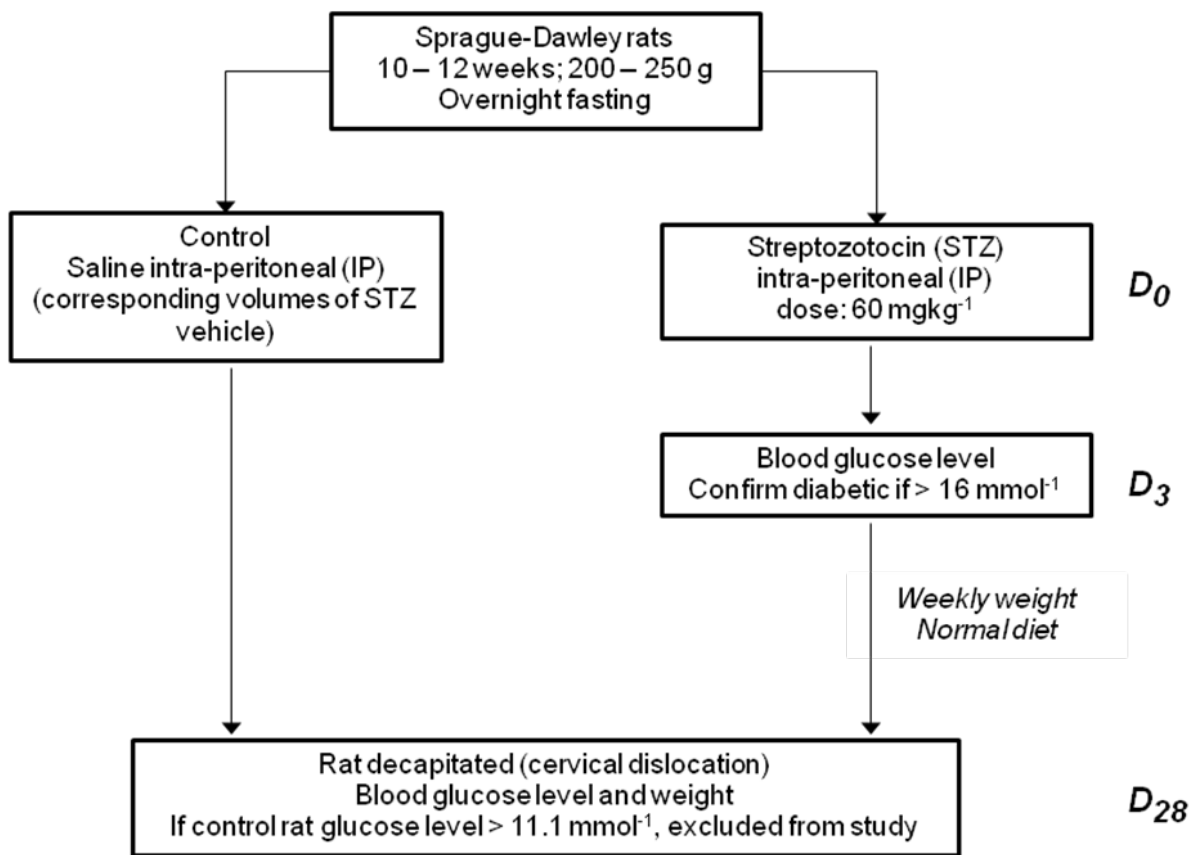
From both agonist and antagonist curve, pA<sub>2</sub> (= - log K<sub>b</sub>) of the antagonist was estimated using the equation below:

$$K_b = \frac{IC_{50}^*}{\frac{[A] - 1}{EC_{50}^*}}$$

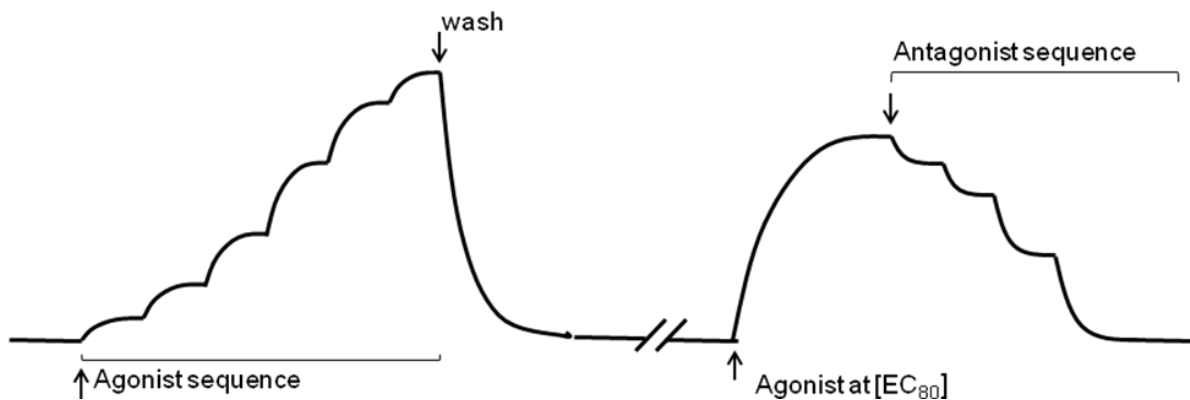
and

$$K_b = \frac{[B_i]}{\frac{[A_f] - 1}{[A_i]}}$$

as been described in detail in General Introduction chapter (section 1.7.2).



**Figure 2.5** The protocol for streptozotocin (STZ)-induced diabetes in rat.



**Figure 2.6** Inhibition curve protocol for estimating antagonist affinity.

## 2.5 Data Analysis

### 2.5.1 Statistical analysis

Contractile responses were measured as increases in tension (g) above the resting level, and normalized to the second 60 mM K<sup>+</sup> response (K-standard) in the case of rat femoral artery and to the second 100 nM carbachol in rat urinary bladder preparation. Relaxant responses were expressed as a percentage of the established tone induced by a pre-contractile agent. Log concentration-response curves were fitted by sigmoidal curves using GraphPad Prism v4.0 software (GraphPad, San Diego, CA). Where appropriate, the lower asymptote was constraint to zero for contractile agonists and the upper asymptote to 100% for relaxant agonists or antagonists. EC<sub>50</sub> and IC<sub>50</sub> values were computed from these curves. Data for individual preparations were used to obtain sigmoidal curve parameters, from which mean  $\pm$  SEM were calculated.

Data were analysed by 1-factor and repeated measures 2-factor ANOVA with GraphPad Prism. A pair of means was compared by planned orthogonal contrasts using SuperANOVA software (Abacus Concepts Inc). Planned contrasts represent a robust method of comparing multiple means (Glass and Hopkins, 1996; Bewick *et al.*, 2004). Initially, main effects were assessed for statistical significance by planned (orthogonal) contrasts; individual cell means were then compared as required. Unpaired Student's *t-test* was used to compare two mean values. All tests were two-tailed and the significance level was set at  $p < 0.05$ .



## 2.5.2 Two-site competition model

When two components / receptors comprise the response inhibited by the prostanoid, the data fitted by the nonlinear regression analysis (one-site competition equation) using GraphPad Prism will appear to deviate from the data points. The next step is to fit the inhibition data using the two-site competition equation in GraphPad Prism. The Hill slopes of both sigmoidal components are automatically constrained to 1.0. If the two  $IC_{50}$  values are at least a log apart with a narrow and significant 95% confidence interval (CI), then the sum-of-squares (SS) is compared statistically using the modified F test (refer to Motulsky and Christopoulos, 2004). The assumption is based on Null hypothesis that two-site model is better than one-site model in fitting the curves. The two models are compared for goodness-of-fit, with adjusting for difference in the number of degrees of freedom (df), and the F ratio between two models. From F table (Appendix), if  $p < 0.05$ , we can conclude that the two-site competition equation model fits the data significantly better than the one-site model.

The regression correlation coefficient ( $r^2$ -values) for the graph fits in nonlinear regression analysis were also analysed (refer to Motulsky and Christopoulos, 2004). The  $r^2$ -values can be obtained from result sheets of the fitted curve with GraphPad Prism. The values range from -1 to 1. A value of 1 implies that a linear equation describes the relationship between  $X$  and  $Y$  perfectly, with all data points lying on a line for which  $Y$  increases as  $X$  increases. A value of -1 implies that all data points lie on a line for which  $Y$  decreases as  $X$  increases. A value of 0 implies that suggests a poor model. The closer the  $r^2$ -value to 1, the better the data fitted by the line.

These two-site competition model comparisons with one-site model were applied in guinea-pig trachea, rat mesenteric artery and rat urinary bladder chapters.

## **2.6 Reagents and chemicals**

Table 2.1 lists the reagents and chemicals used in the current experiments.

**Table 2.1** Reagents and chemicals used in the current experiments

Agents	Source	Stock	Solvent
17-Phenyl PGE <sub>2</sub> 17-phenyl- $\omega$ -trilor prostaglandin E <sub>2</sub>	Cayman Chemical, USA	10 mM	Absolute ethanol
AH-13205 7-[2-[4-(1-hydroxyhexyl)phenyl]-5-oxo cyclopentyl]heptanoic acid	Cayman Chemical, USA	10 mM	Absolute ethanol
Acetylcholine chloride	Sigma-Aldrich Chemicals, USA	10 $\mu$ M	Distilled water
BMS-180291 [1s-( <i>exo,exo</i> )]-2-[[3-[4- [(pentylamino)carbonyl]-2-oxazolyl]-7- oxabicyclo[2.2.1]hept-2-yl]methyl]- benzenepropanoic acid	Bristol-Myers Squibb, USA.	1 mM	Absolute ethanol
Butaprost-FA ( $\pm$ )-15-deoxy-16S-hydroxy-17-cyclo butyl PGE <sub>1</sub>	Cayman Chemical, USA	10 mM	Absolute ethanol
Carbachol hydrochloride	Sigma-Aldrich Chemicals, USA	10 mM	Distilled water
CAY-10399 ( $\pm$ )-15-deoxy-16S-hydroxy-17-cyclo butyl PGE <sub>2</sub>	Cayman Chemical, USA	10 mM	Absolute ethanol
CAY-10441 (4,5-dihydro-1H-imidazol-2-yl)-[4-(4- isopropoxybenzyl)phenyl]amine	Cayman Chemical, USA	1 mM	Absolute ethanol
CP-533536 2-[3-[N-(4-tert-butylbenzyl)-N-(pyridin- 3-ylsulfonyl)aminomethyl]phenoxy]ace tic acid	Pfizer, UK	10 mM	Absolute ethanol
GW-848687 6-[2-(5-chloro-2-[(2,4- difluorophenyl)methyl]oxy}phenyl)-1-	GlaxoSmithKline Research & Development, UK	1 mM	Dimethylsulphoxide (DMSO)

<b>Agents</b>	<b>Source</b>	<b>Stock</b>	<b>Solvent</b>
cyclopenten-1-yl]-2-pyridinecarboxylic acid			
Histamine diphosphate	Sigma-Aldrich Chemicals, USA	10 $\mu$ M	Distilled water
Iloprost 6,9 $\alpha$ -methylene-11 $\alpha$ ,15S-dihydroxy-16-methyl-prosta-5E,13E-dien-18-yn-1-oic acid	Cayman Chemical, USA	10 $\mu$ M	Absolute ethanol
Indomethacin	Sigma-Aldrich Chemicals, USA	20 mM	Absolute ethanol
Isoprenaline hydrochloride	Sigma-Aldrich Chemicals, USA	100 $\mu$ M	Distilled water
L-798106 5-bromo-2-methoxy-N-[3-(naphthalen-2-yl-methylphenyl)-acryloyl]-benzene sulphonamide	GlaxoSmithKline Research & Development, UK	10 mM	Dimethylsulphoxide (DMSO)
L-826266 [(2E)-N-[(5-bromo-2-methoxyphenyl)-sulfonyl]-3-[5-chloro-2-(2-naphthylmethyl)phenyl]acrylamide	Dr David Woodward, Allergan Pharmaceuticals, USA	10 mM	Dimethylsulphoxide (DMSO)
Lataprost-FA 17-phenyl-13,14-dihydro prostaglandin F <sub>2<math>\alpha</math></sub>	Cayman Chemical, USA	10 mM	Absolute ethanol
ONO-AE1-259 11,15-O-dimethyl prostaglandin E <sub>2</sub>	Ono Pharmaceuticals, Japan	10 $\mu$ M	Absolute ethanol
ONO-AE-248 11,15-O-dimethyl-PGE <sub>2</sub>	Dr David Woodward, Allergan Pharmaceuticals, USA	10 mM	Absolute ethanol
ONO-DI-004 (17S)-2,5-ethano-6-oxo-17,20-dimethyl PGE <sub>1</sub>	Dr David Woodward, Allergan Pharmaceuticals,	10 mm	Absolute ethanol

<b>Agents</b>	<b>Source</b>	<b>Stock</b>	<b>Solvent</b>
	USA		
PGF <sub>2α</sub> 9α,11α,15S-trihydroxy-prosta-5Z,13E-dien-1-oic acid	Cayman Chemical, USA	10 μM	Absolute ethanol
Phenylephrine hydrochloride	Sigma-Aldrich Chemicals, USA	1 μM	Distilled water
Potassium chloride	Sigma-Aldrich Chemicals, USA	1 mM	Distilled water
Prostaglandin E <sub>2</sub>	Cayman Chemical, USA	10 mM	Absolute ethanol
SC-51322 8-chlorodibenz[b,f][1,4]oxazepine-10(11H)-carboxylic acid, 2-[3-[2-(furanyl methyl) thio]1-oxopropyl]hydrazine	Biomol International, USA	10 μM	Absolute ethanol
Streptozotocin 2-deoxy-2-(3-(methyl-3-nitrosoureido)-D-glucopyranose	Alexis Biochemical, UK	20 mg/ml	Sodium-Citrate
Sulprostone	Cayman Chemical, USA	5 mM	Absolute ethanol
U-46619 9,11-dideoxy-9α,11α-methanoepoxyprostaglandin F <sub>2α</sub>	Cayman Chemical, USA	10 μM	Absolute ethanol

# **CHAPTER THREE**

## **GUINEA-PIG TRACHEA**

### 3.1 Introduction

The guinea-pig isolated trachea (GPT) was used in the early part of my PhD study. In view of my lack of laboratory experience, this preparation was considered a suitable starting point to measure agonist and antagonist potency in smooth muscle preparations.

This robust preparation has been widely used in the past to characterize selective  $\beta_2$ -adrenoceptor agonists and for studies on autonomic innervation. The presence of prostanoid receptors in the guinea-pig trachea has been well documented. Activation of EP<sub>1</sub> and TP receptors has been shown to cause a contractile response (Jones *et al.*, 1982; Coleman and Kennedy, 1985; Eglen and Whiting, 1988; McKenniff *et al.*, 1988; Ndukwu *et al.*, 1997). On the other hand, a relaxant effect was demonstrated to be mediated by EP<sub>2</sub> receptor (Gardiner, 1986; Eglen and Whiting, 1988).

Initially, the activities of PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub>, sulprostone and U-46619 were investigated. PGE<sub>2</sub> is a prostanoid which activates both contractile and relaxant EP receptors, with weak affinity on the TP receptor (Kiriyaama *et al.*, 1997; Abramovitz *et al.*, 2000). 17-Phenyl PGE<sub>2</sub> has a moderate affinity to EP<sub>1</sub>, with some activity to EP<sub>3</sub> and minimal to EP<sub>2</sub> (Lawrence *et al.*, 1992; Breyer *et al.*, 2001). Sulprostone has a higher affinity to EP<sub>3</sub> more than EP<sub>1</sub>, with some activity on FP receptors (Kiriyaama *et al.*, 1997; Abramovitz *et al.*, 2000).

Subsequently, the activities of several EP<sub>2</sub> agonists were investigated. I was particularly interested in AH-13205, which does not show the IOP-lowering activity typical of others EP<sub>2</sub> agonists (Woodward *et al.*, 1995). The intention was to examine the nature of the AH-13205 log concentration-response curve (CRC) to see whether there is any evidence for partial agonism at the EP<sub>2</sub> receptor. Analogously, the ability of AH-13205 to relax under high contractile tone was

investigated; relative to full agonist, a EP<sub>2</sub> partial agonist should show a poorer relaxation when working against higher tone. In addition to PGE<sub>2</sub> and AH-13205, other EP<sub>2</sub> agonists examined were butaprost-FA; CAY-10399 and CP-533535 (Gardiner, 1986; Armstrong, 1995; Kiriya *et al.*, 1997; Abramovitz *et al.*, 2000; Breyer *et al.*, 2001; Tani *et al.*, 2001; Paralkar *et al.*, 2003; Jones, 2004). CP-533535 has a non-prostanoid structure (refer to Figure 1.3).

Table 3.1 lists the antagonists used in the current study with their respective affinities on the prostanoid receptors. McKeniff *et al.* (1988) have demonstrated that the contractile response to U-46619 in guinea-pig trachea is selectively antagonised by the TP antagonists EP-092 and AH-23848. The study also demonstrated that the EP<sub>1</sub> antagonist AH-6809 selectively antagonised the responses to 16,16-dimethyl PGE<sub>2</sub> and PGF<sub>2 $\alpha$</sub> . In the current study, the potent and selective EP<sub>1</sub> antagonist, SC-51322 was used to antagonise the EP<sub>1</sub> effect, while BMS-180291 was used to block the TP receptor (Zhang *et al.*, 1996; Clarke *et al.*, 2004; Hung *et al.*, 2006). Using agonists, direct contraction via EP<sub>3</sub> receptors has not been demonstrated in previous studies on guinea-pig trachea; however EP<sub>3</sub> receptors on parasympathetic nerve endings inhibit transmitter release (Spicuzza *et al.*, 1998; Clarke *et al.*, 2004). In the current study, L-798106, a selective EP<sub>3</sub> antagonist was used to demonstrate whether activation of the EP<sub>3</sub> receptor causes contraction of the tracheal smooth muscle (Clarke *et al.*, 2004).

**Table 3.1** Antagonists and concentrations chosen for use in the current study.

<b>Antagonists</b>	<b>Reported pA<sub>2</sub></b>	<b>Concentration</b>	<b>Expected dose-ratio</b>	<b>References</b>
SC-51322	8.45	1 $\mu$ M	280	Hung <i>et al.</i> , 2006
BMS-180291	9.5 – 9.8	0.1 $\mu$ M	320 – 630	Zhang <i>et al.</i> , 1996
L-798106	7.48 – 7.82	1 $\mu$ M	30 – 66	Clarke <i>et al.</i> , 2004



## 3.2 Methods

The basic methodology used has been described in detail in Chapter 2. Specific methodological points not discussed previously are addressed below.

### 3.2.1 Setting up of preparations

The cervical trachea was removed from guinea pig, mounted on myograph as described in Chapter 2. Resting tone was set at 1.5 g, determined from as an optimal resting tension in preliminary experiment, and within range of other published studies (McKenniff *et al.*, 1988; Udem and Adams, 1988; Schlemper *et al.*, 2005). After period of equilibration of 1 hour, viability of the tracheal ring was determined as described in Chapter 2. 1  $\mu$ M indomethacin was added in Krebs solution throughout the experiment with the tracheal ring.

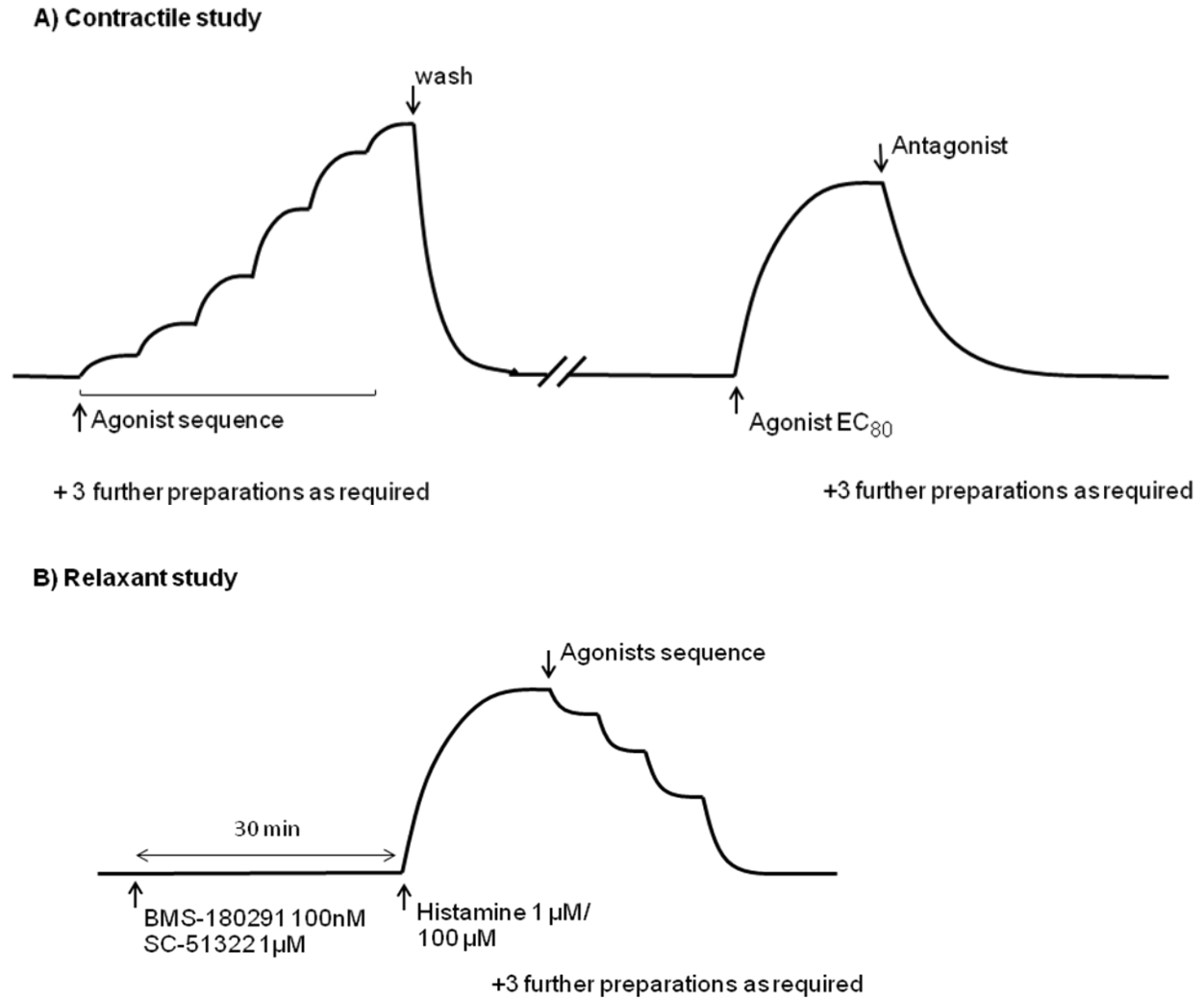
In the contractile studies, the prostanoid agonists were added to determine the maximal response ( $E_{max}$ ) with the construction of CRC's. Except when U-46619 used as agonist, 100 nM BMS-180291 was added 30 min before the addition of other prostanoid agonist. Following period of washout, the preparations were subsequently contracted with the agonist to 80% of maximal response ( $EC_{80}$ ), with the addition of the selective antagonist once the contraction had stabilised (Figure 3.1A).

In the relaxant sequences, 1  $\mu$ M SC-51322 and 100 nM BMS-180291 were added 30 min before addition of the pre-contractile agent, histamine (see results section). Histamine was chosen (at a concentration corresponding to 30 or 80% maximum response) as it is a non-prostanoid regulator of airways smooth muscle. Furthermore, the contraction produced by histamine is consistent and

does not fade throughout the experimental period. Once the tone is established and stable, PGE<sub>2</sub> or other prostanoids (EP<sub>2</sub> agonists) were cumulatively added to construct cumulative-inhibition curves (Figure 3.1B).

### **3.2.2 Statistical analysis**

Contractile responses were measured as increases in tension (g) above the resting level. The relaxant responses were expressed as a percentage loss of the initial contractile tone developed to the established tone of the contractile agonist or histamine. A variable-slope sigmoidal curve was fitted to log concentration–response data using GraphPad Prism software; the bottom asymptote was constrained to zero for contraction and to 100% for relaxation. Sigmoidal curve parameters were derived from data for individual preparations. Data were further analysed by 1-factor and repeated measures 2-factor ANOVA combined with comparison of selected means by planned (orthogonal) contrasts using SuperANOVA software; all tests were two-tailed and the significance level was set at  $p < 0.05$ . Where applicable, data were analysed by the two-site competition equation using the modified F test in combination with the  $r^2$ -values comparison (refer to Motulsky and Christopoulos, 2004). All data are presented as mean  $\pm$  SEM.



**Figure 3.1** Experimental protocols for guinea-pig trachea. A) The protocol used for contractile study. B) The protocol used in relaxant study, where GPT was precontracted with two different concentrations of histamine, corresponding to EC<sub>30</sub> and EC<sub>80</sub> of maximal histamine responses.

## **3.3 Results**

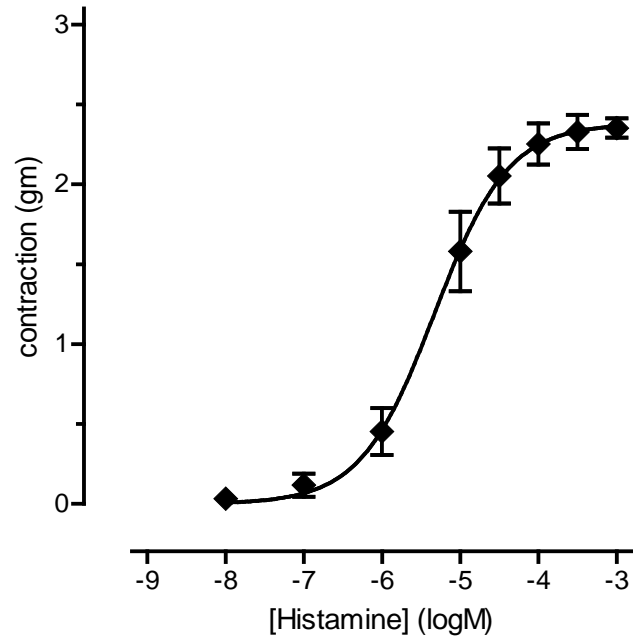
### **3.3.1 Effects of histamine**

Histamine contracted GPT in a concentration-dependent manner with maximal response of  $2.4 \pm 0.2$  g and  $pEC_{50}$  of  $5.32 \pm 0.17$  (Figure 3.2,  $n = 4$ ).

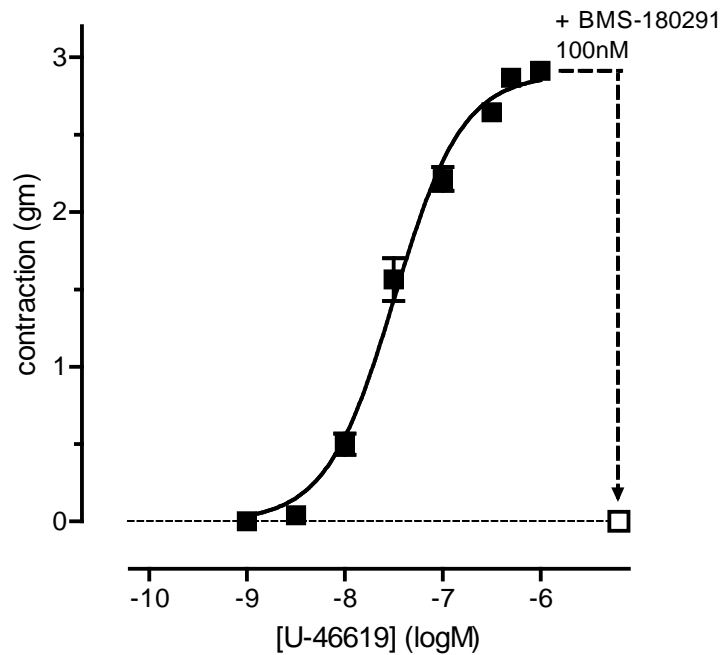
### **3.3.2 Effects of prostanoid agonists**

In early part of the study, the effect of U-46619, a selective TP agonist was studied. Increasing concentrations of U-46619 (1 nM – 1  $\mu$ M) were added cumulatively. Log concentration-responses curve (CRC) was plotted as shown in Figure 3.3 ( $n = 8$ ). U-46619 contracted the preparation to a maximum response ( $E_{max}$ ) of  $2.8 \pm 0.2$  g. The  $pEC_{50}$  was  $7.57 \pm 0.07$ . BMS-180291 at 100 nM completely blocked the established tone.

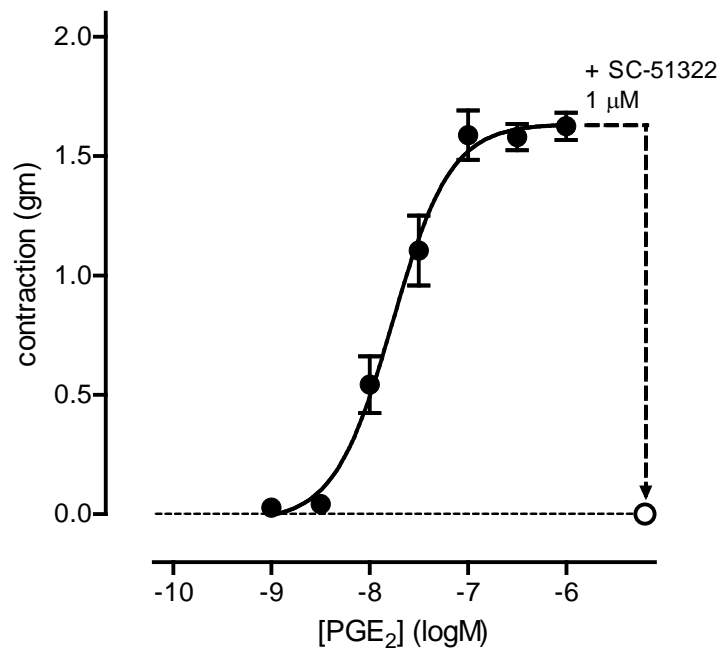
$PGE_2$ , a non-selective EP agonist was examined in the presence of 100 nM BMS-180291 to block any influence from activation of TP receptors.  $PGE_2$  (1 nM – 1  $\mu$ M) produced a concentration-dependent contractile response in GPT as demonstrated in Figure 3.4 ( $n = 4$ ). The maximal response was  $1.6 \pm 0.1$  g with  $pEC_{50}$  of  $7.76 \pm 0.09$ . Administration of 1  $\mu$ M SC-51322 to the established tone of  $PGE_2$  reduced the contractile response to resting tension.



**Figure 3.2** Contractile activity of histamine on guinea-pig trachea. Histamine contracted the trachea in concentration-dependent manner ( $n = 4$ ).



**Figure 3.3** Contractile activity of U-46619 on guinea-pig trachea. U46619 (1 nM – 1  $\mu$ M), a selective TP agonist contracted the trachea in a concentration dependent manner ( $n = 8$ ). 100 nM BMS-180291 completely antagonised the response.



**Figure 3.4** Contractile activity of PGE<sub>2</sub> on guinea-pig trachea. PGE<sub>2</sub> (1 nM – 1 μM), a non-selective agonist at EP receptors contracted the trachea in concentration-dependent manner ( $n = 4$ ). 100 nM BMS-180291 was present. 1 μM SC-51322 completely relaxed the tone.

In the presence of BMS-180291, 17-phenyl PGE<sub>2</sub> (1 nM – 1 μM) contracted GPT in concentration-dependent manner with E<sub>max</sub> 1.5 ± 0.1 g and pEC<sub>50</sub> 7.63 ± 0.09 (Figure 3.5, *n* = 4). Likewise, the established tone of 17-phenyl PGE<sub>2</sub> was completely inhibited by 1 μM SC-51322.

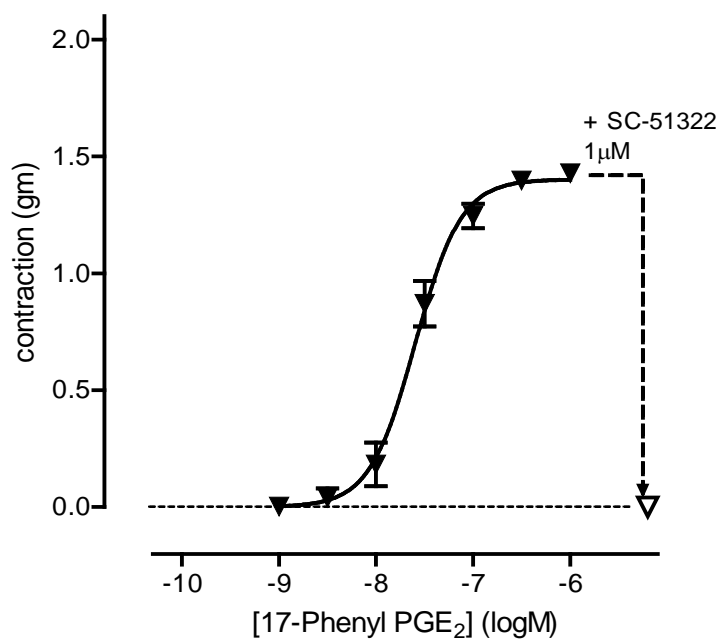
The effects of sulprostone, (affinity of EP<sub>3</sub> > EP<sub>1</sub>, minimal on EP<sub>4</sub> and EP<sub>2</sub>) on GPT can be observed as in Figure 3.6 (*n* = 4). Sulprostone (1 nM – 1 μM) produced a contractile response in dose-dependent manner with E<sub>max</sub> of 2.3 ± 0.2 g and pEC<sub>50</sub> of 7.88 ± 0.05. SC-51322 at 1 μM abolished the established tone by 100 nM sulprostone. In contrast, L-798106 at 1 μM had no effect on the 100 nM sulprostone-induced tone.

The contractile effects of the agonists used are summarized in Table 3.2.

The selectivity of SC-51322 on GPT using the inhibition-curve protocol is demonstrated in Figure 3.7. SC-51322 inhibited the established tone of 10 nM 17-phenyl PGE<sub>2</sub> in concentration-dependent manner. At 1 μM, SC-51322 completely blocked the 17-phenyl PGE<sub>2</sub> contraction. At 10 μM, SC-51322 relaxed of the 30 nM U-46619-induced tone by about 10% and had no effect at all on 10 nM carbachol-induced tone.

At this point, it was decided to include 100 nM BMS-180291 and 1 μM SC-51322, pre-incubated for 30 min for all agonist or antagonist sequences, to ensure that there was no interference from TP and EP<sub>1</sub> contractile component in the relaxant studies.





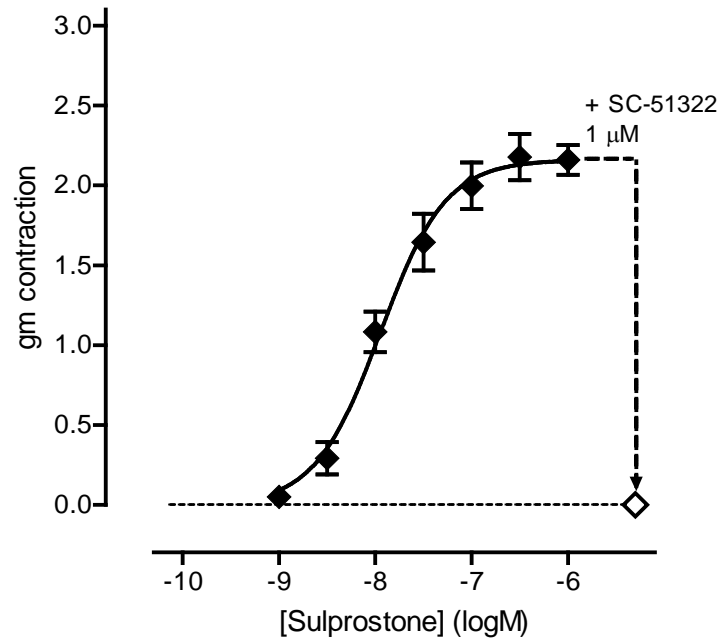
**Figure 3.5** Contractile activity of 17-phenyl PGE<sub>2</sub> on guinea-pig trachea. 17-Phenyl PGE<sub>2</sub> (1 nM – 1 μM) was used in the experiment (*n* = 4). 100 nM BMS-180291 was present. 1 μM SC-51322 completely relaxed the tone.

**Table 3.2** The pEC<sub>50</sub> and maximal response (E<sub>max</sub>) of the prostanoid agonists in the contractile study.

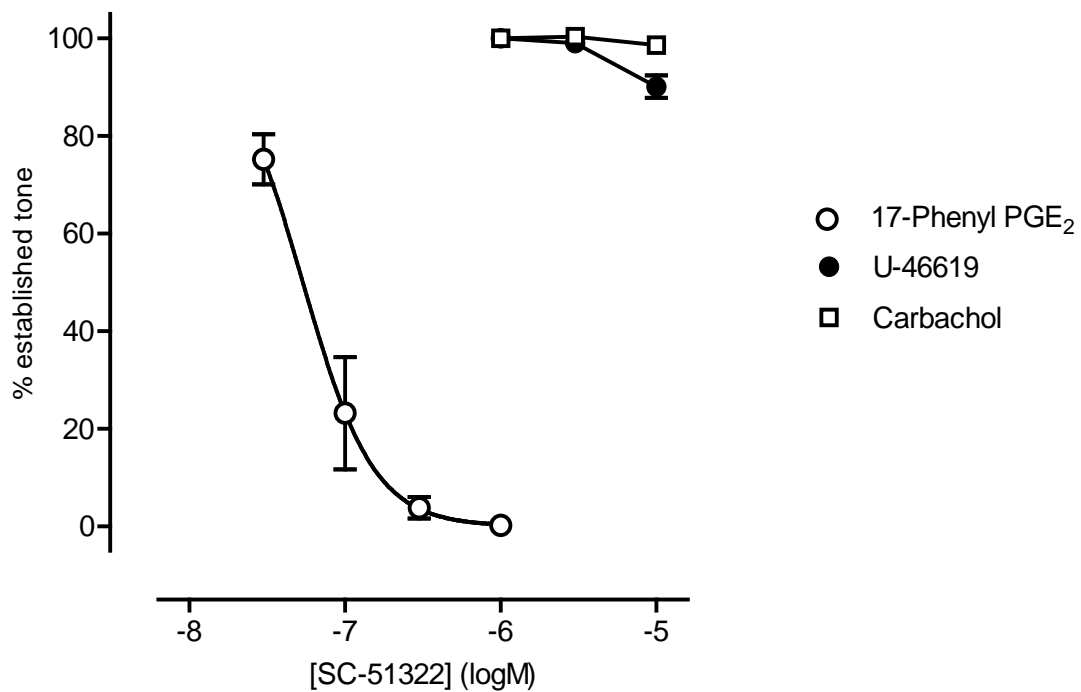
	pEC <sub>50</sub>	E <sub>max</sub> (g)
U-46619	7.37 ± 0.07	2.8 ± 0.2
PGE <sub>2</sub>	7.76 ± 0.09	1.6 ± 0.1
17-Phenyl PGE <sub>2</sub>	7.63 ± 0.09	1.5 ± 0.1
Sulprostone	7.88 ± 0.05	2.3 ± 0.2†

\* All the experiments were done in the presence of 1 μM indomethacin and 100 nM BMS-180291 (When U-46619 as the agonist, BMS-180291 was omitted).

† *P* < 0.05, sulprostone vs. PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub>.



**Figure 3.6** Contractile activity of sulprostone on guinea-pig trachea. Sulprostone (1 nM – 1 μM) was used in the experiment ( $n = 4$ ). 100 nM BMS-180291 was present. 1 μM SC-51322 completely relaxed the tone.



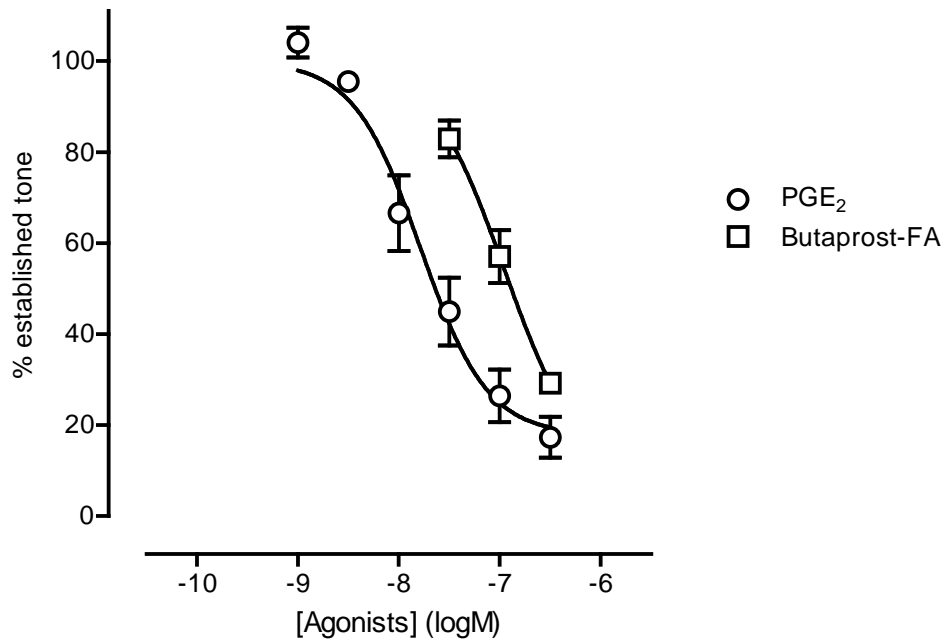
**Figure 3.7** Antagonism by SC-51322 of contraction induced by 10 nM 17-phenyl PGE<sub>2</sub>, 30 nM U-46619 and 10 nM carbachol in guinea-pig trachea ( $n = 4$ ,  $n = 2$ ,  $n = 3$ ; respectively). 1  $\mu$ M indomethacin is present.

### **3.3.3 Effects of PGE<sub>2</sub> and butaprost on precontracted guinea-pig tracheal ring under low tone**

Preliminary experiments established that histamine produced a stable contractile response in the GPT. Hence, histamine was chosen as the contractile agent. Once the tone was established, the prostanoid agonist was added cumulatively to construct an inhibition curve (Figure 3.1B).

Initially, GPT was precontracted with histamine 1  $\mu$ M (representing 30% of maximal histamine contraction). The response to cumulative doses of PGE<sub>2</sub> and the selective EP<sub>2</sub> antagonist, butaprost are shown in Figure 3.8 ( $n = 4$ ). PGE<sub>2</sub> and butaprost-FA (a selective EP<sub>2</sub> agonist) relaxed the preparation in a concentration-dependent manner. Butaprost-FA was about 6 times less potent than PGE<sub>2</sub>. Butaprost-FA is considerably less potent than PGE<sub>2</sub> on EP<sub>4</sub> receptors (Wilson *et al.*, 2004), and therefore at this stage, it was concluded that the relaxation by PGE<sub>2</sub> is most likely mediated by the EP<sub>2</sub> receptor. It is also clear that SC-51322 at 1  $\mu$ M is sufficient for inhibition of the EP<sub>1</sub> agonist activity of PGE<sub>2</sub>, thereby exposing its EP<sub>2</sub> relaxant activity. The other agonists studied were expected to have lower EP<sub>1</sub> agonist potencies than PGE<sub>2</sub>.

After further experimentation, it was decided that differences in receptor efficacy on the EP<sub>2</sub> system might be demonstrable if relaxation curves were obtained against a much higher tone level. Histamine concentrations of 1  $\mu$ M and 100  $\mu$ M were therefore chosen to contract the tracheal rings, corresponding to about 30 and 80% of the histamine maximum respectively.



**Figure 3.8** Relaxation activity of PGE<sub>2</sub> and butaprost-FA on pre-contracted guinea-pig trachea. GPT was pre-contracted with 1  $\mu$ M histamine. PGE<sub>2</sub> (1 – 300 nM) and butaprost (30 – 300 nM) were used as agonists ( $n = 4$ ). 100 nM BMS-180291 and SC-51322 1  $\mu$ M were present.

### **3.3.4 Comparison of EP<sub>2</sub> agonists on precontracted guinea-pig tracheal ring under low tone and high tone**

Table 3.3 listed the maximal relaxation and pIC<sub>50</sub> of the EP<sub>2</sub> agonists used in the current study.

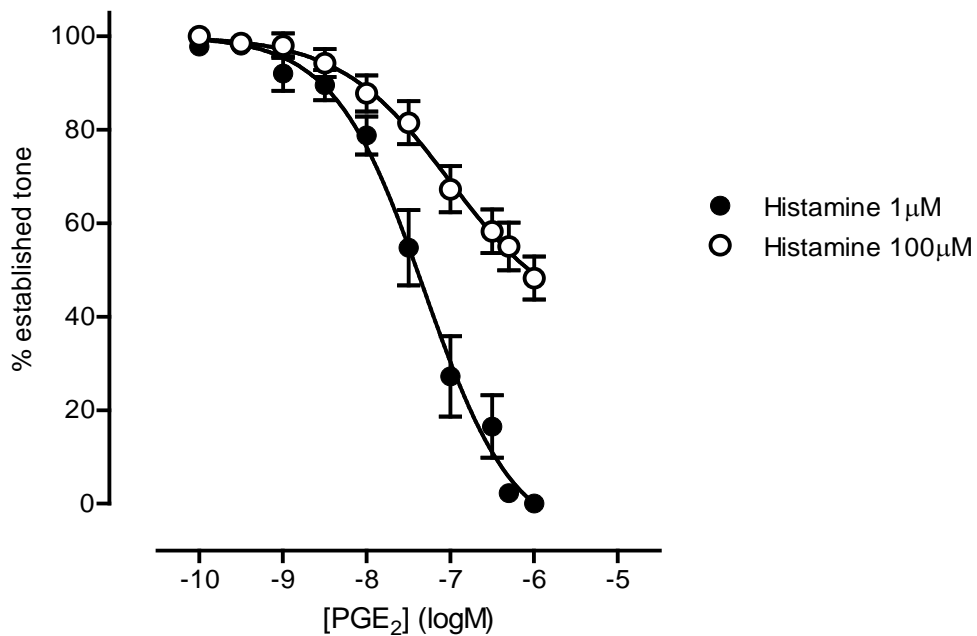
PGE<sub>2</sub> induced maximal relaxation of the histamine (1 μM)-contracted tracheal ring at a concentration of 1 μM (Figure 3.9; *n* = 4). Under higher tone, the relaxation curve to PGE<sub>2</sub> was shifted to the right and only 52% relaxation was found at 1 μM.

From Figures 3.10, 3.11, 3.12, and 3.13 and Table 3.3, it is clear that butaprost-FA, CAY-10399, CP-533536 and AH-13205 showed similar profiles to PGE<sub>2</sub>. There were inconsistencies however. Firstly, the slopes of the curves appeared to be different between the agonists. Instead of using the Hill slope (a sigmoidicity measure that is dependent on how the lower asymptote is fixed), the relaxation curves of these EP<sub>2</sub> agonists can be compared as the concentration interval for 20 - 80% relaxation of 1 μM histamine-induced tone as a measurement of slope (Figure 3.14, example on PGE<sub>2</sub> relaxation curve). The log intervals for 20 - 80% relaxation are: PGE<sub>2</sub> 1.45; CAY-10399 1.25; Butaprost-FA 1.40; CP-533536 1.10 and AH-13205 1.9. The steeper slope of the CP-533536 curve may be due to its slower onset at concentration ≤ 100 nM; it is possible that the corresponding responses have been under-estimated. The shallower slope of the AH-13205 curve appears to be a genuine result.

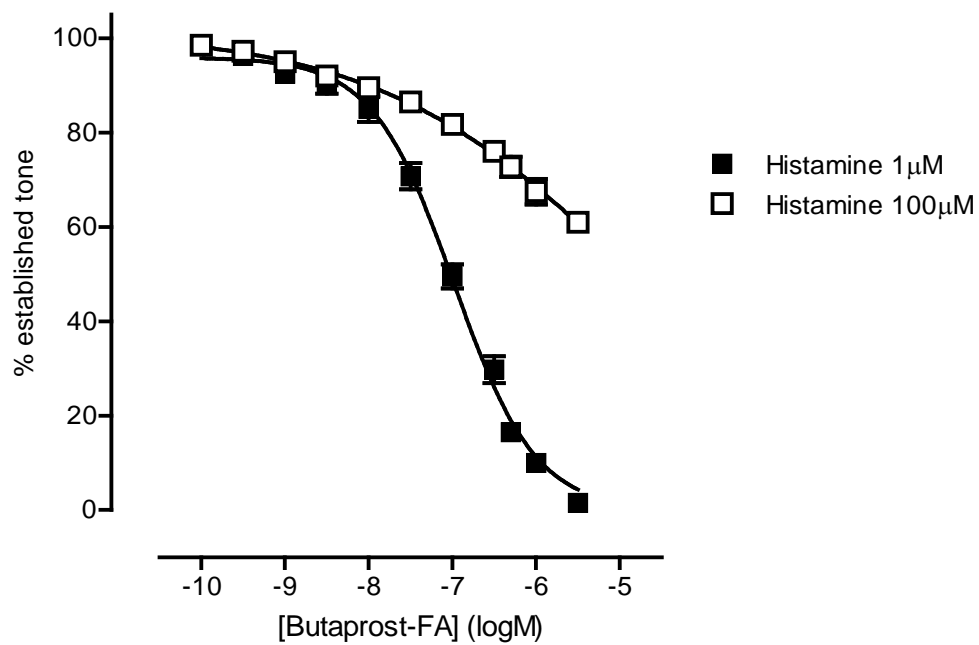
**Table 3.3** Guinea-pig trachea: pIC<sub>50</sub> values and maximal inhibitions of the prostanoid agonists under two different levels of histamine tone in the relaxant study.

	Histamine 1 μM		Histamine 100 μM
	pIC <sub>50</sub>	% relaxation	% relaxation
PGE <sub>2</sub>	7.30 ± 0.11	100 ± 0	52 ± 5
Butaprost-FA	6.98 ± 0.03	98 ± 1	32 ± 3
CAY-10399	7.64 ± 0.07	100 ± 0	39 ± 1
CP-533536	6.97 ± 0.06	100 ± 0	52 ± 5
AH-13205	6.84 ± 0.06	97 ± 2	53 ± 4

All the experiments were done in the presence of 1 μM indomethacin, 100 nM BMS-180291 and 1 μM SC-51322.

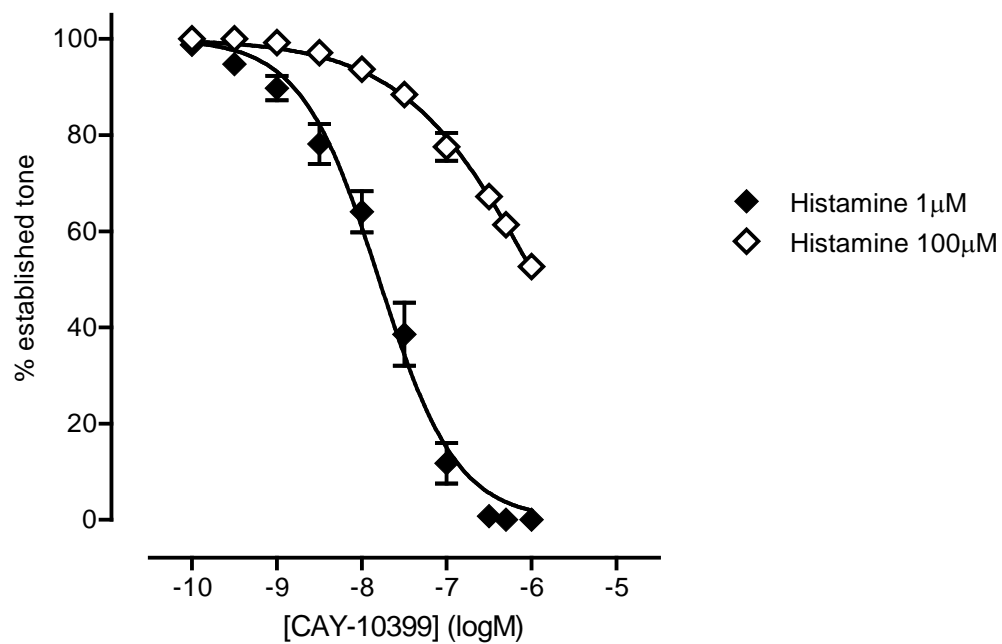


**Figure 3.9** Relaxant activity of PGE<sub>2</sub> on pre-contracted guinea-pig trachea. GPT were pre-contracted with histamine 1 μM and 100 μM (*n* = 4). 100 nM BMS-180291 and 1 μM SC-51322 were present.

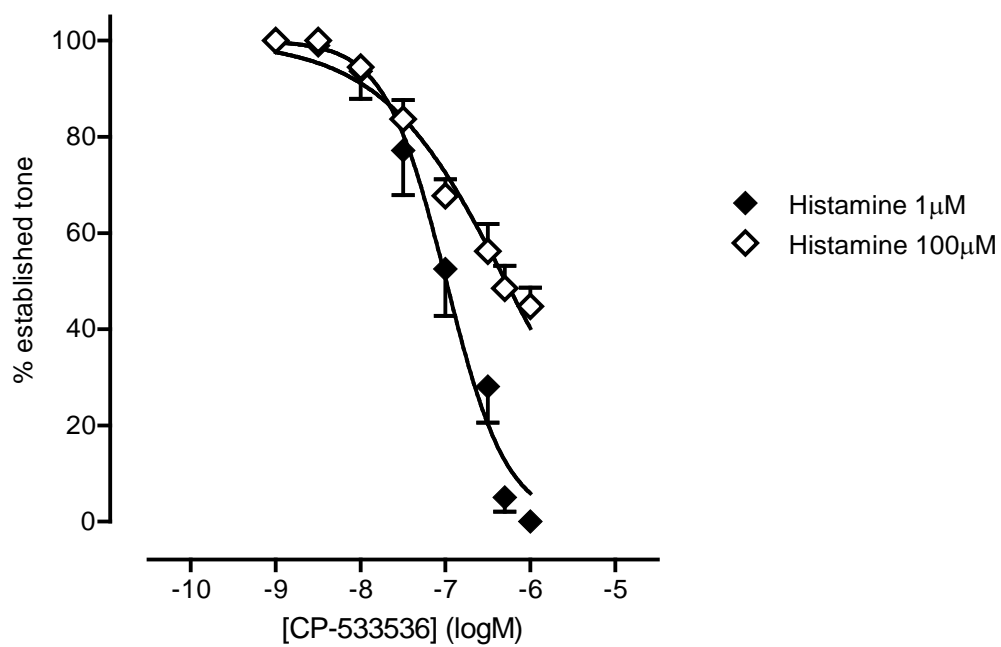


**Figure 3.10** Relaxant activity of butaprost-FA on pre-contracted guinea-pig trachea. GPT were pre-contracted with histamine 1  $\mu\text{M}$  and 100  $\mu\text{M}$  ( $n = 4$ ). 100 nM BMS180291 and 1  $\mu\text{M}$  SC51322 were present.

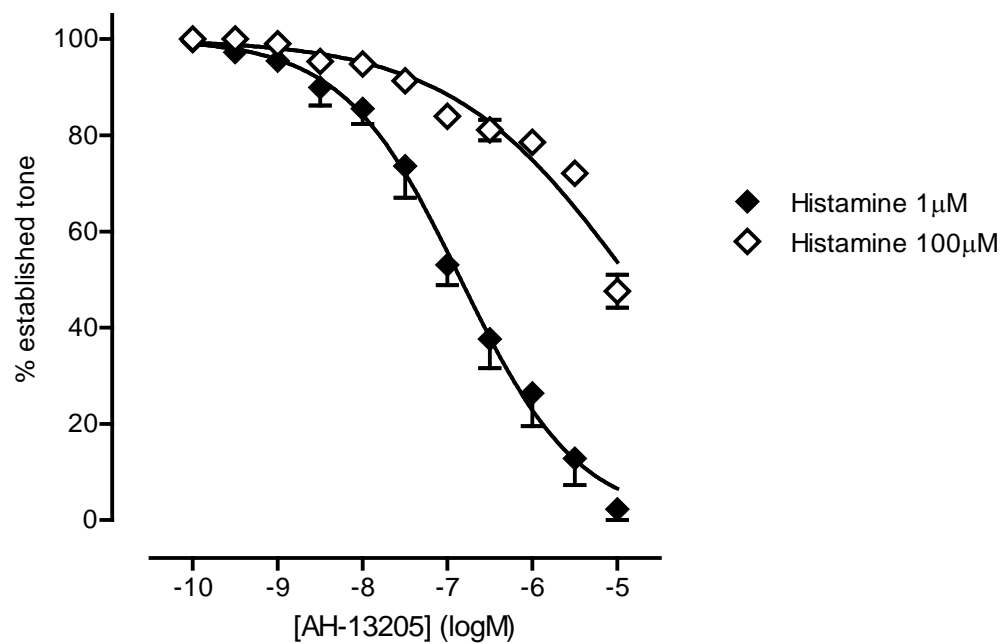




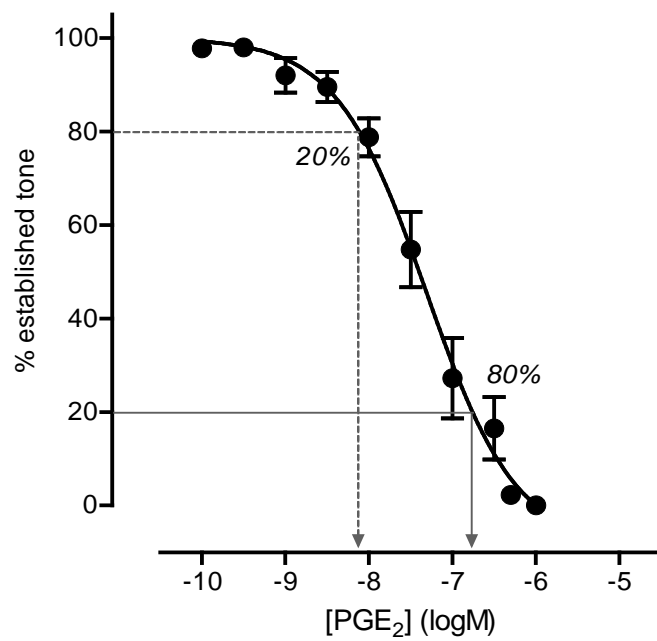
**Figure 3.11** Relaxant activity of CAY-10399 on guinea-pig trachea. The trachea was contracted with 1  $\mu$ M and 100  $\mu$ M histamine ( $n = 4$ ). 100 nM BMS-180291 and 1  $\mu$ M SC-51322 were present.



**Figure 3.12** Relaxant activity of CP-533536 on guinea-pig trachea. The trachea was contracted with 1  $\mu$ M and 100  $\mu$ M histamine ( $n = 4$ ). 100 nM BMS-180291 and 1  $\mu$ M SC-51322 were present.



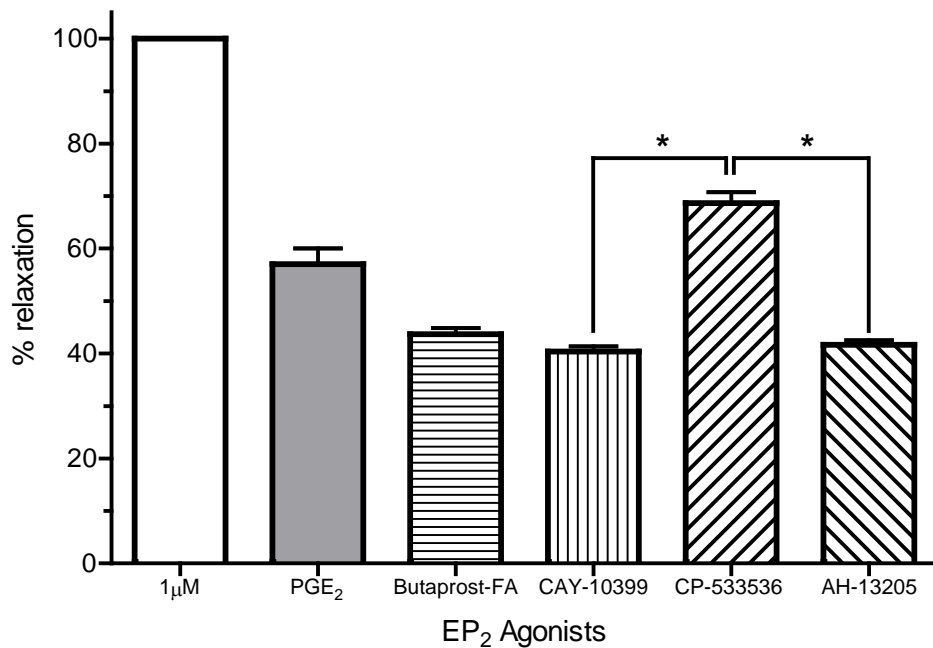
**Figure 3.13** Relaxant activity of AH-13205 on guinea-pig trachea. Tone was induced by 1  $\mu$ M ( $n = 4$ ) and 100  $\mu$ M ( $n = 4$ ) histamine. 100 nM BMS-180291 and 1  $\mu$ M SC-51322 were present.



**Figure 3.14** The logM interval for 20 - 80% relaxation of 1  $\mu$ M histamine-induced contraction as an alternative method measurement of curve slope (data from Figure 3.9).

In terms of estimating efficacy, it was decided to compare relaxations under high and low tone for each agonist at the level of 75% relaxation under low tone. The fraction of the 75% relaxation under low tone for the EP<sub>2</sub> agonists are: PGE<sub>2</sub> 57%; Butaprost-FA 47%; CAY-10399 43%; CP-533536 73% and AH-13205 44% (Figure 3.15). There were significant differences between CP-533536 in compared to CAY-10399 and AH-13205 ( $p = 0.003$ ). Otherwise, there were no large differences using this procedure.

The log concentration-response curve for AH-13205 under high tone was atypical (Figure 3.13) and it was decided to analyse these data further.



**Figure 3.15** Relaxation induced by EP<sub>2</sub> agonists under high tone (histamine 100 μM) corresponding to 75% relaxation under low tone (histamine 1 μM). Values expressed as a percentage of the low tone relaxation. \*  $P < 0.01$ , CP-533536 vs CAY-10399 and AH-13205, using one-factor ANOVA.

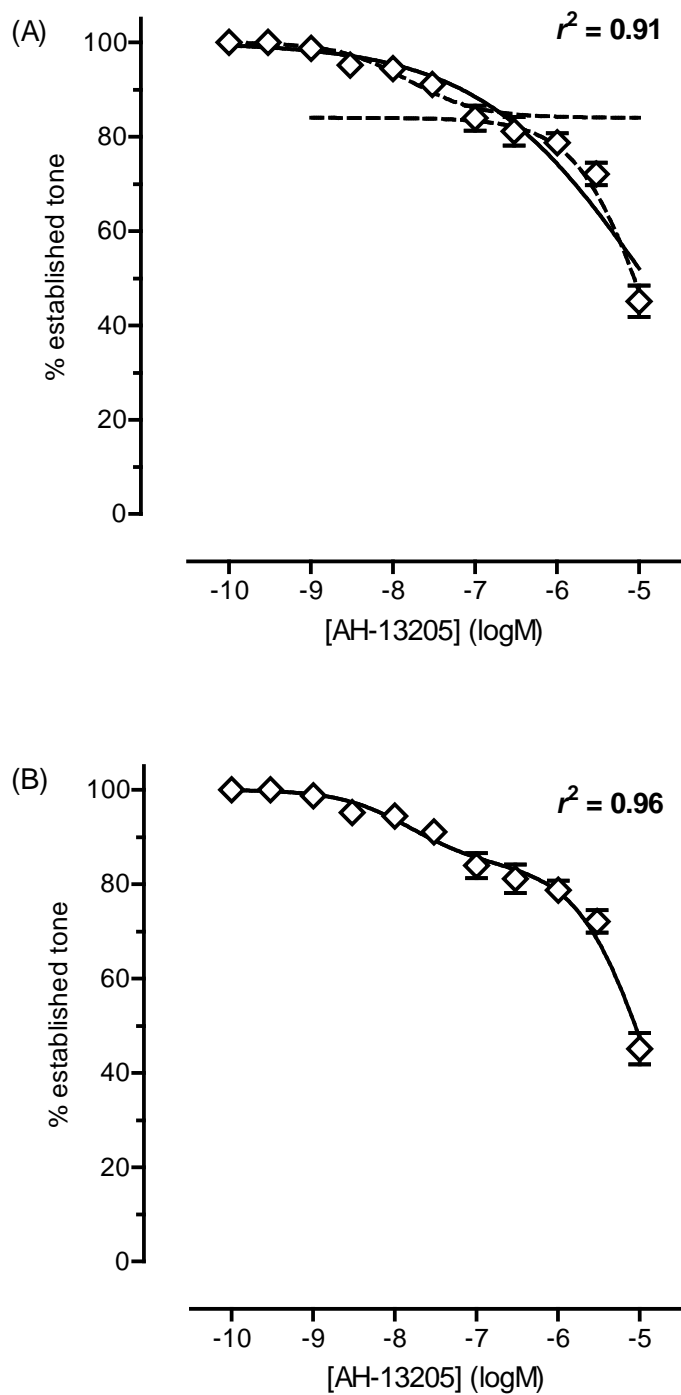
### 3.3.5 Two-site competition model

It appeared that there could be two components to the inhibition-curve for AH-13205 against 100  $\mu$ M histamine-induced tone, as the data points did not fit the standard sigmoidal curve well (Figure 3.13). Two theoretical curves were fitted to the inhibition data using the two-site competition equation in GraphPad Prism as described in Chapter 2 (Figure 3.16A). The combined curve is shown in Figure 3.16B. The  $r^2$ -values were compared as described in Chapter 2. The  $r^2$ -value was 0.96 for the two-component fit and 0.91 for the one-component fit.

The best-fit two-site curve has a high affinity site with a  $pIC_{50}$  of 7.72 and a low affinity site with  $pIC_{50}$  of 4.84. The fraction of high affinity site (the upper curve) is 16%, with 95% confidence intervals of 12 – 19%. These results are sensible to make a statistical comparison using the modified F test as described in Section 2.5.2. There are 44 data points in the curve (i.e. 4 replicates, with 11 different concentrations;  $4 \times 11 = 44$ ). Four parameters are fitted by the one-site equation fit:  $IC_{50}$ , slope, top and bottom and five parameters fitted for the two-site model:  $IC_{501}$ ,  $IC_{502}$ , top, bottom and common middle-level. Using these data, the calculation is summarized in Table 3.4. The F ratio is 28.78 with 1 (numerator) and 28 (denominator) degrees of freedom (df). With references to the F table (Appendix), the two-site model is significantly better than one-site model ( $p < 0.001$ ).

**Table 3.4** The F test of two-site model against one-site model of AH-13205 vs 100  $\mu$ M histamine.

Model	SS	df
Null hypothesis (1 site)	862	44
Null hypothesis (2 site)	456	43
Difference	406	1
<b>Fractional difference</b>	0.8904	0.0233
<b>Ratio (F)</b>	<b>38.21</b>	



**Figure 3.16** An attempt to establish a two-receptor mechanism for AH-13205 in guinea-pig trachea by applying the two-site competition equation to data from Figure 3.13. (A) AH-13205 curve vs. 100  $\mu$ M histamine, showing the two theoretical curves. (B) The two-site model fitted to the data.

## 3.4 Discussion

The guinea-pig tracheal ring has been shown to have contractile prostanoid receptors, namely TP and EP<sub>1</sub> (Jones *et al.*, 1982; Coleman and Kennedy, 1985; Eglen and Whiting, 1988; McKenniff *et al.*, 1988), while the relaxant activity of prostanoids has been demonstrated to be mediated by the EP<sub>2</sub> receptor (Eglen and Whiting, 1988). As an established preparation, it was a good point to gain experience of isolated smooth muscle preparations, especially since my data could be compared to previous studies.

### 3.4.1 The tracheal response to prostanoid agents

Table 3.2 lists the response of GPT to prostanoid agonists in the contractile study. The potent TP agonist, U-46619 (Coleman *et al.*, 1981) produced a contractile effect in the guinea-pig trachea. In the current study, the preparation were slightly less sensitive to the U-46619 (pEC<sub>50</sub> 7.37) than reported in previous studies (Jones *et al.*, 1982, 7.60; McKenniff *et al.*, 1988, 7.47; Tymkewycz *et al.*, 1991, 7.8). This may be due to difference in source of the animal and the techniques applied among the studies. The complete antagonism of U-46619-induced contraction by 100 nM BMS-180291 is consistent with the presence of TP receptor as demonstrated by previous studies (Jones *et al.*, 1982; Coleman and Kennedy, 1985; Eglen and Whiting, 1988; McKenniff *et al.*, 1988).

PGE<sub>2</sub> is a prostanoid agonist that has affinity to all the EP receptors; in the rank order: EP<sub>3</sub> = EP<sub>4</sub> > EP<sub>1</sub> > EP<sub>2</sub> with very low affinity for FP, DP, TP and negligible on IP (> 10,000) (Abramovitz *et al.*, 2000). In the current study, the response of GPT to cumulative addition of PGE<sub>2</sub> was similar to 17-phenyl PGE<sub>2</sub> (EP<sub>1</sub> > EP<sub>3</sub>, with minimal activity on EP<sub>2</sub>; Lawrence *et al.*, 1992).



However, PGE<sub>2</sub>-induced contraction is completely antagonised by the EP<sub>1</sub> antagonist SC-51322, while the EP<sub>3</sub> antagonist L798106 has no effect.

Sulprostone is an extremely potent EP<sub>3</sub> agonist, for example on guinea-pig vas deferens (Lawrence *et al.*, 1992) and aorta (Jones *et al.*, 1998). Sulprostone has higher affinity to EP<sub>3</sub> receptor than EP<sub>1</sub> receptor (Lawrence *et al.*, 1992; Abramovitz *et al.*, 2000). In the current study, the pEC<sub>50</sub> for sulprostone corresponded well to K<sub>i</sub> values obtained by radioligand binding using recombinant receptors. Similar to PGE<sub>2</sub>, the contractile action of sulprostone was blocked by SC-51322, but not by L-798106. The results indicate that EP<sub>3</sub> receptors are absent from GPT smooth muscle. Collectively, these results agree with the presence of EP<sub>1</sub> receptor in the GPT.

### **3.4.2 Relaxant response of the trachea to EP<sub>2</sub> agonists**

The initial relaxant studies with PGE<sub>2</sub> and butaprost-FA supported the presence of EP<sub>2</sub> receptor as opposed to EP<sub>4</sub> receptor in GPT. All the EP<sub>2</sub> agonists investigated induced a complete or nearly complete relaxation under 1 µM histamine tone (Figure 3.14). In other words, they are considered to be EP<sub>2</sub> full agonists. Using the data for 1 µM histamine, the pIC<sub>50</sub> values of PGE<sub>2</sub>, butaprost-FA, CAY-10399, CP-533536 and AH-13205 were 50, 105, 23, 107 and 145 nM respectively. The corresponding equipotent molar ratios are 1.0, 2.1, 0.46, 2.14 and 2.9. Thus the potency ranking of the EP<sub>2</sub> agonists on GPT demonstrated in the current study was: CAY-10399 > PGE<sub>2</sub> > butaprost-FA = CP-533536 > AH-13205. However, the slopes of the relaxation curves under low tone were different. In the case of CP533536, this may have been due to its slow onset of action. CP-533536 is much more lipophilic than the other agonist studied and this may underlie its slow onset (Jones RL, personal communication).

Under higher tone induced by 100  $\mu$ M histamine, all the prostanoid agonists were all much less effective relaxants. An agonist with lower efficacy would be expected to elicit relatively less relaxation under the condition of higher tone; this was seen with CP-533536 but not with AH-13205.

### 3.4.3 Dual receptor activity of AH-13205

The data for AH-13205 against 100  $\mu$ M histamine seems to be poorly fitted using one-site competition model (Figure 3.13). Using the two-site competition model, two theoretical curves were superimposed on the AH-13205 curve (Figure 3.16A). The curve for two-site competition refitted on the curve using GraphPad Prism is shown in Figure 3.16B. Statistical analysis established that the two-curve fitting was significantly better than the single-curve fitting, indicating that AH-13205 has two components to its relaxation curve.

One explanation of the biphasic relaxant effect of AH-13205 at high tone is activation of two receptors, the EP<sub>2</sub> receptor and another receptor that remain to be determined. AH-13205 has been used as a selective EP<sub>2</sub> agonist (Wheeldon and Vardey, 1993) and has a binding  $K_i$  of 240 nM for mouse recombinant EP<sub>2</sub> receptors (Kiriya *et al.*, 1997; Breyer *et al.*, 2001). However, AH-13205 also has affinity for EP<sub>3</sub> receptor, with  $K_i$  of 82 nM (Kiriya *et al.*, 1997; Breyer *et al.*, 2001). Moreover, in recent experiments on guinea-pig aorta by one of my supervisors, AH-13205 has been shown to be a low-potency EP<sub>3</sub> agonist (Jones RL, unpublished result). However, no EP<sub>3</sub> receptor has been documented in guinea-pig trachea smooth muscle in this and previous studies. Inhibitory EP<sub>3</sub> receptors are present on parasympathetic neurones in the guinea-pig trachea (Spicuzza *et al.*, 1998; Clarke *et al.*, 2004), but it seems unlikely that the contractile action of histamine involves activation of the parasympathetic innervation.

A second explanation relates to the chemical structure of AH-13205. Figure 1.3 shows a wavy line for the hydroxyl group attachment in the  $\omega$ -chain of AH-13205. This means that the compound is a mixture of isomers, one with the hydroxyl group down from the plane of the paper, the other above the plane. The ratio will be close to 1:1, since they are difficult to separate chromatographically. The other PGE<sub>2</sub> analogues studied all have a single configuration for this hydroxyl. It is usual for one of the two possible hydroxyl isomers to be more active than the other. For example, 15(R)-PGE<sub>2</sub> is 870 times less potent than natural 15(S)-PGE<sub>2</sub> in a recombinant EP<sub>1</sub> receptor-Ca<sup>2+</sup> flux assay (Ungrin *et al.*, 2001). It is possible that a similar scenario applies to AH-13205.

### **3.5 Conclusions**

The results from the current study are in agreement with the presence of TP, EP<sub>1</sub> and EP<sub>2</sub> receptors in the guinea-pig trachea. The selective EP<sub>2</sub> agonists used in the current study all behaved as full agonists.

In contrast to the other EP<sub>2</sub> agonists, AH-13205 showed a biphasic log concentration-response curve. This profile may be due to activation of the EP<sub>2</sub> receptor and a second receptor that needs to be elucidated. Alternatively, the presence of two isomers in AH-13205 may be responsible.

# **CHAPTER FOUR**

## **RAT MESENTERIC ARTERY**

## 4.1 Introduction

Essential hypertension (EH) is one of the most important treatable causes of premature death worldwide (Ezzati *et al.*, 2002). It is a major public health problem in many countries due to its high prevalence and its association with coronary heart disease, stroke, renal disease, peripheral vascular disease and other disorders. EH is a heterogenous, multi-factorial disorder. Both genetic and environmental factors influenced the onset and severity of EH (Gong and Hubner, 2006).

Maintenance of normal blood pressure is dependent on a balance between the cardiac output and peripheral vascular resistance. In humans, the elevated blood pressure in EH is associated with a normal cardiac output but raised peripheral resistance (Dustan *et al.*, 1972). Thus, the main contribution to the high blood pressure in EH is the elevation in arterial tone contributed to the changes in peripheral resistance. The smaller arterial resistance vessels (generally less than 200  $\mu\text{m}$  in diameter) are most involved in regulating blood flow and capillary pressure (Christensen and Mulvany, 2001). However, all the peripheral vessels, including aorta contribute to the peripheral resistance to some extent.

*In-vitro* study on an animal model of hypertension is an ideal method to study the pathophysiology and the effect of treatment in EH. The rat is the most common animal used in hypertension research. Previous work with isolated vessels in hypertensive rats has involved aorta, conductance arteries and resistance vessels. The rat superior mesenteric artery is considered to be a conductance artery having internal diameters greater than 300  $\mu\text{m}$ , but less than 1 mm (Mulvany and Halpern, 1977; Christensen and Mulvany, 2001). Structural abnormality in this conductance artery leading to wall stiffness, is considered to play an important role in the development of hypertension (Korner and Angus, 1997) and to be a predictor of cardiovascular events in essential hypertension (Mathiassen *et al.*, 2007). The contribution of the rat mesenteric bed in hypertension has been investigated using wire-

myography (Mulvany and Halpern, 1977; Garcia-Redondo *et al.*, 2009) and under various conditions of pressure-myography (Coats and Hillier, 1999; Bolla *et al.*, 2004). However, the main trunk of the artery has not frequently been used in previous experiments.

Noradrenaline has been shown to have a strong contractile effect on this artery (Masset *et al.*, 1998; Buus *et al.*, 2000). Phenylephrine also produced a good contractile response in the proximal part of the main trunk (Masset *et al.*, 1998). The superior rat mesenteric artery has been demonstrated to have a marked similarity in control of its vascular tone to the human counterpart (Hutri-Kahonen *et al.*, 1999). However, the scope of the study was limited to endothelium-derived mediators only.

In mesenteric artery preparations from normal male Wistar rats, arachidonic acid (AA) was shown to inhibit established contraction to noradrenaline in the presence and absence of endothelium and the effect was comparable to that of human small subcutaneous arteries (Buus *et al.*, 2000). AA may be converted to PGE<sub>2</sub> and / or PGI<sub>2</sub>, which could then activate EP<sub>2</sub> / EP<sub>4</sub> and IP receptors on smooth muscle cells. The same study also demonstrated that increasing the extracellular potassium concentration (more than 7 mM) caused a transient relaxation in rat mesenteric artery, which was absent in human small arteries. In contrast, a transient relaxation response to extracellular potassium was absent in male Sprague-Dawley rats despite the higher concentration of KCl, up to 100 mM (Masset *et al.*, 1998).

The response of the rat mesenteric artery to prostanoid agonists has been demonstrated in several studies. U-46619, a potent TP receptor agonist induced a strong contractile response with concentration ranging from 1 nM to 1 μM (Chauhan *et al.*, 2003). U-46619 also has been used to precontract rat mesenteric artery to study the effect of 17β-estradiol-induced relaxation in the presence of various antagonists (Tsang *et al.*, 2003). In another study, the presence of periadventitial adipose tissue have been demonstrated to have no effect on rat mesenteric artery response to 100 nM U-46619 (Verlohren *et al.*, 2004). Of particular relevance to the current

investigations is the demonstration of strong constriction of perfused second order mesenteric artery vessels from male Wistar-Kyoto rats by PGE<sub>2</sub> in the presence of the TP antagonist SQ-29548 (Bolla *et al.*, 2004).

The rat mesenteric artery was used in the current study to study the role of the prostaglandins in the control on the vascular tone for three reasons. Firstly, the distribution of prostanoid receptors in rat mesenteric artery has never been documented before. Secondly, its wall thickness is small and it was hope that responses to prostanoids would be conveniently faster than those previously seen in more robust preparations such as rabbit and guinea-pig aorta and human pulmonary artery. Thirdly, there is a marked similarity in the function of mesenteric arterial rings of corresponding size *in vitro* between humans and rats (Hutri-Kahonen *et al.*, 1999). Therefore, the rat mesenteric artery is a useful model to study the physiology and pathophysiology of arterial function. The superior rat mesenteric artery has been chosen in the current study due to its easy accessibility and it provided a good and consistent response to prostanoid agonists in preliminary experiments.

In order to classify prostanoid receptors in rat mesenteric artery, the antagonist protocol described in detail in Chapter 2 was used in the current study (Figure 4.1B). Basically, the protocol involved construction of an initial agonist concentration-response curve (CRC) followed by a concentration-inhibition curve for a selective antagonist on established response of the agonist. Prostanoid agonists (selectivity specified) that usually induce smooth muscle contraction include: 17-phenyl PGE<sub>2</sub> (EP<sub>1</sub> > EP<sub>3</sub>), sulprostone (EP<sub>3</sub> > EP<sub>1</sub>), ONO-D1-004 (EP<sub>1</sub>), ONO-AE-248 (EP<sub>3</sub>), PGF<sub>2α</sub> (FP) and U-46619 (TP). Prostanoid agonists inducing relaxation include: butaprost-FA (EP<sub>2</sub>), ONO-AE1-259 (EP<sub>2</sub>) and iloprost (IP). A selective EP<sub>4</sub> agonist was not available during the period of the study.

Prostanoid receptor antagonists that were used in the current study include a well-established EP<sub>1</sub> antagonist, SC-51322 (Hallinan *et al.*, 1994). L-798106 was the only selective EP<sub>3</sub> receptor antagonist available for the study. It has been reported to show high selectivity on EP<sub>3</sub> receptors based on ligand binding at recombinant prostanoid receptors ( $K_i$  1.1 nM; Juteau *et al.*, 2001). Previous functional study on EP<sub>3</sub> receptor shows L-798106 blocked the inhibitory action of sulprostone pre-synaptically in guinea-pig vas deferens and trachea with pA<sub>2</sub> values of 7.48 and 7.82, respectively (Clarke *et al.*, 2004). At the time of the current study, there was no other report of L-798106 activity on other isolated tissue or vessel preparation. Recently, L-798106 (CM9) was shown to inhibit the PGE<sub>2</sub>-induced Ca<sup>2+</sup> influx in a rat EP<sub>3</sub> receptor-Ca<sup>2+</sup> flux assay with pK<sub>i</sub> value of 7.12 (Jugus *et al.*, 2009). Another selective EP<sub>3</sub> receptor antagonist, L-826266 also was used in the current study (Clark *et al.*, 2008).

BMS-180291 was used as the selective TP antagonist in the study (Ogletree *et al.*, 1993). Concentrations were chosen to give large rightward shifts of the log concentration-response curves of the appropriate agonists, based on previously published affinity data (Table 4.1).

**Table 4.1** Antagonists and concentrations chosen for use in the studies

Antagonists	Reported pA <sub>2</sub>	Concentrations used in the current study	References
SC-51322	8.45	1 nM - 1 μM	Hung <i>et al.</i> , 2006
BMS-180291	9.5	1 – 300 nM	Zhang <i>et al.</i> , 1996
L-798106	7.48, 7.82	1 μM	Clarke <i>et al.</i> , 2004
L-826266	8.35	100 nM – 1 μM	Clark <i>et al.</i> , 2008



## 4.2 Methods

The basic methodology used has been described in detail in Chapter 2. Specific methodological points not discussed previously are addressed below.

### 4.2.1 Setting up of preparations

The superior mesenteric arteries were obtained as described in Chapter 2. The artery was either used the same day or kept overnight at 4 °C in Krebs solution (viability as demonstrated by McIntyre *et al.*, 1998). The resting tone was set at an optimal tension of 1.0 g, determined from preliminary experiments and documented in previous study (Maggi *et al.*, 1988; Schneider *et al.*, 2004). In the early part of the study, the resting tone of the rat mesentery artery was erratic. It was found that setting the resting tension in the presence of 100 nM isoprenaline produced more stable preparations. Drugs were added directly to the tissue bath and the concentrations of drugs were the final concentrations in the bath. Thirty minutes after being set up in organ baths, the arterial rings were constricted by 40 mM of potassium chloride to assess the contractility and were then washed several times until baseline tone was restored. Any arterial rings not contracted by more than 0.5 g (representing 50% of normal 40 mM potassium chloride response) were excluded from the study. The vessel rings were then exposed to 10 nM PGE<sub>2</sub> for 10 min followed by a washout period of 15 min and equilibration for 30 – 40 min; increasing agonist concentrations were added cumulatively to construct CRC.

Each preparation was used to obtain a concentration-response curve for one prostanoid only. The selective antagonist was either applied as a single dose before the construction of agonist CRC (Figure 4.1A) or as cumulative doses when the inhibition-curve protocol was utilised (Figure 4.1B).

In inhibition curve studies, the arterial rings were precontracted with agonist (at 80% of maximal response; EC<sub>80</sub>), followed by cumulative addition of antagonist (Figure 4.1B). Preparation without the antagonist served as a control.

For studies of agonist-induced relaxation, phenylephrine 300 nM (representing 35% of maximal response, EC<sub>35</sub>) was chosen as the precontractile agent. The selective antagonists for contractile prostanoid receptor were incubated 30 min before the addition of phenylephrine. Once the contraction had stabilised, the agonist was added cumulatively (Figure 4.2). Phenylephrine was used as the precontractile agent in antagonist studies, in the presence of selective antagonist pertaining to the agonist used. Once the phenylephrine tone was established, a cumulative-dose response curve to the particular agonist was constructed.

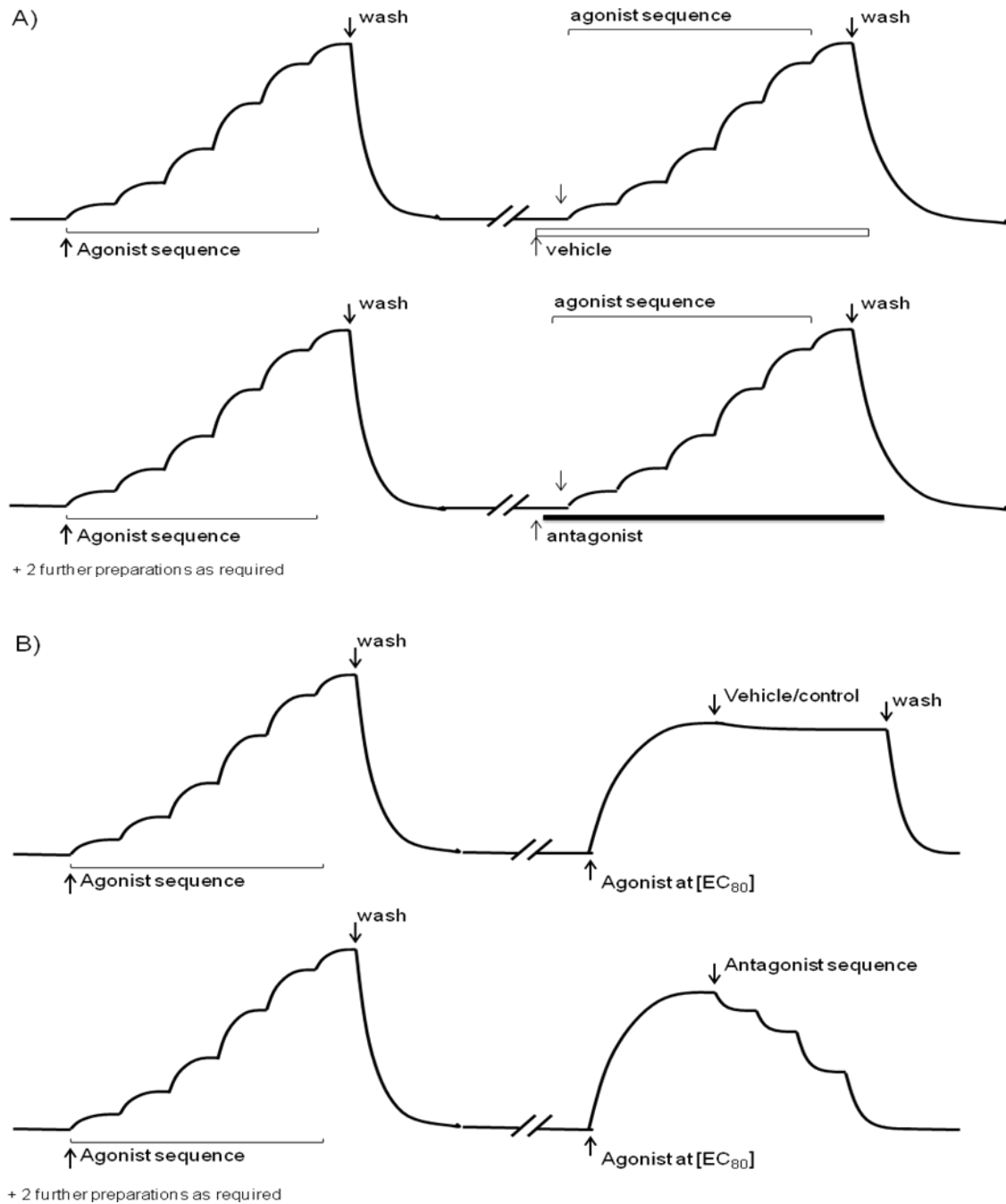
#### 4.2.2 Antagonist protocol and pA<sub>2</sub> estimation

Antagonist affinity was determined by calculating the pA<sub>2</sub>. In this study, the basic inhibition-curve protocol is used. The details have been described in Section 1.7.2. The pA<sub>2</sub> (= -logK<sub>b</sub>) is estimated from the equation (Lazareno and Birdsall, 1993b):

$$K_b = \frac{IC_{50}^*}{\frac{[A]}{EC_{50}^*} - 1}$$

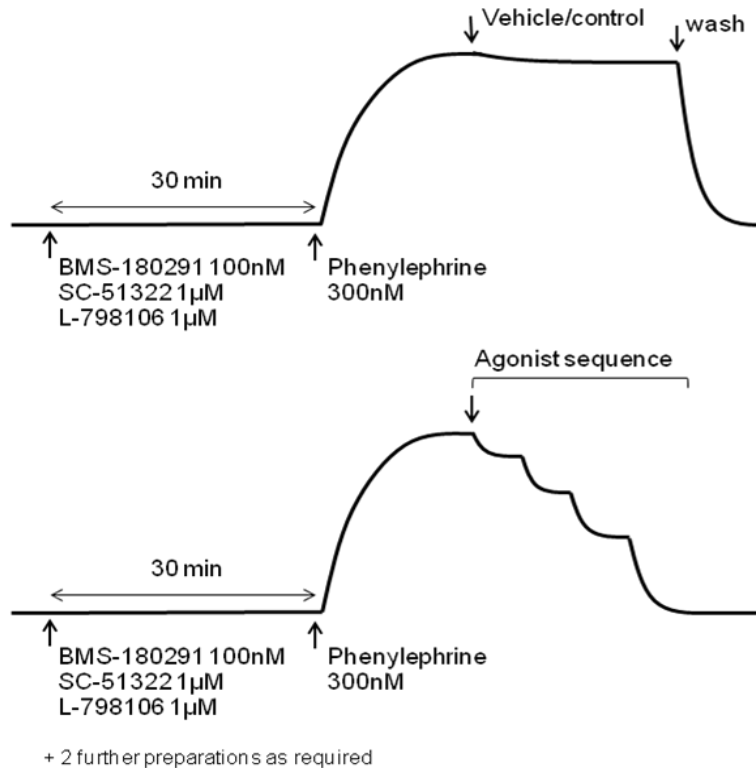
and

$$K_b = \frac{[B_i]}{\frac{[A_f]}{[A_i]} - 1}$$



**Figure 4.1** Protocols used for rat mesentery artery to determine the profile of an antagonist and where appropriate its affinity constant. A) The basic protocol: antagonist at different concentrations was added before the cumulative addition of prostanoid agonist in the second

sequence. B) The inhibition-curve protocol: an initial agonist sequence followed by cumulative inhibition of the response to an EC<sub>80</sub> concentration of the same agonist.



**Figure 4.2** Protocol used in rat mesenteric artery to determine the presence of relaxant prostanoid receptor. All three selective antagonists were added 30 min before pre-contraction with phenylephrine 300 nM, followed by the cumulative addition of the agonist.

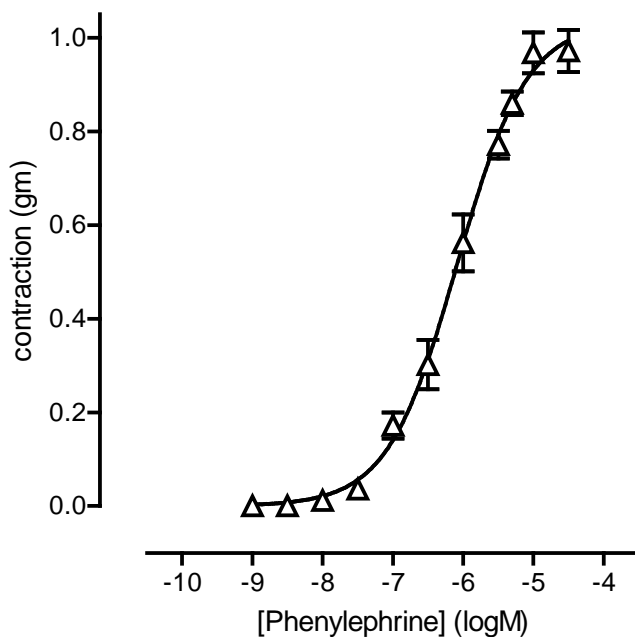
### 4.2.3 Statistical analysis

Contractile responses were measured as increases in tension (g) above the resting level. Relaxant responses were expressed as a percentage loss of the initial contractile tone developed to the established tone of the contractile agonist. A variable-slope sigmoidal curve was fitted to log concentration–response data using GraphPad Prism software; the bottom asymptote was constrained to zero for contraction and to 100% for relaxation. Sigmoidal curve parameters were derived from data for individual preparations. Data were further analysed by 1-factor and repeated measures 2-factor ANOVA combined with comparison of selected means by planned (orthogonal) contrasts using SuperANOVA software; all tests were two-tailed and the significance level was set at  $p < 0.05$ . Where applicable, data were analysed by the two-site competition equation using the modified F test in combination with the  $r^2$ -values comparison. All data are presented as mean  $\pm$  SEM.

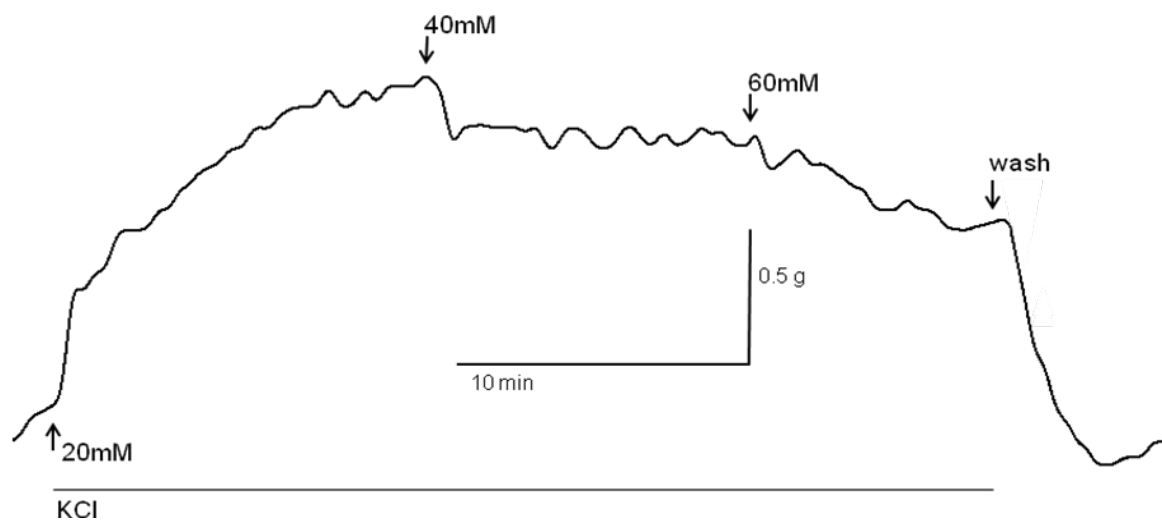
## 4.3 Results

### 4.3.1 Initial investigations

All experiments on rat mesenteric artery rings were conducted in the presence of 1  $\mu\text{M}$  indomethacin to suppress prostanoid biosynthesis. Phenylephrine produced a sustained contraction with the maximal response of  $0.97 \pm 0.04$  g and a  $\text{pEC}_{50}$  of  $6.31 \pm 0.20$  (Figure 4.3,  $n = 5$ ). The contraction was not affected by addition of 100 nM BMS-180291, 1  $\mu\text{M}$  SC-51322 or 1  $\mu\text{M}$  L-798106. Potassium chloride contraction was not sustained and started to fade at concentration of 40 mM and above (Figure 4.4). Histamine up to 10  $\mu\text{M}$  did not contract the ring.



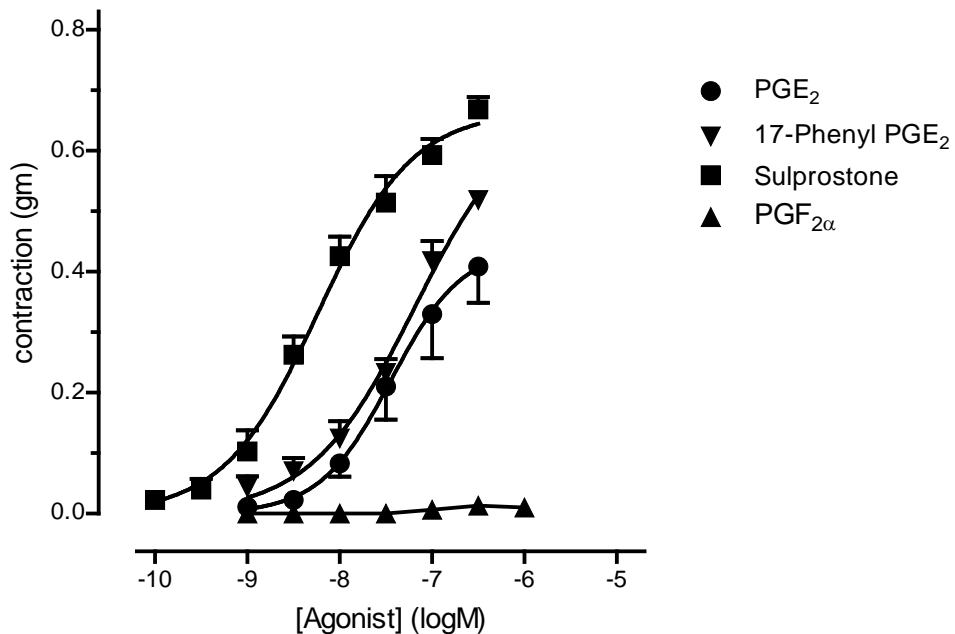
**Figure 4.3** Contractile activity of phenylephrine in rat mesenteric artery. Phenylephrine (1 nM – 30  $\mu\text{M}$ ) contracted the artery in a concentration dependent manner ( $n = 5$ ).



**Figure 4.4** Trace illustrating the effect of increasing potassium chloride concentration in the rat mesenteric artery. At concentration of 40 mM and above, the contractile response starts to fade.

Initial experiments showed that three prostanoid agonists, PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub> and sulprostone, produced a contractile response on the arterial rings (Figure 4.5, *n* = 7). The pEC<sub>50</sub> values for sulprostone, 17-phenyl PGE<sub>2</sub> and PGE<sub>2</sub> were 7.80, 7.41 and 6.87, affording equi-effective molar ratios (EMR) of 1.0, 2.46 and 8.48, respectively. The sulprostone curve appeared to be parallel to that of 17-phenyl PGE<sub>2</sub>. The absolute contractile responses produced by the prostanoid agonists were consistent throughout the study and normalisation was not considered necessary.

PGF<sub>2α</sub>, the natural ligand for the FP receptor was also tested on the rat mesenteric artery. No contraction was observed at concentrations ranging from 1 – 300 nM (Figure 4.5, *n* = 4).



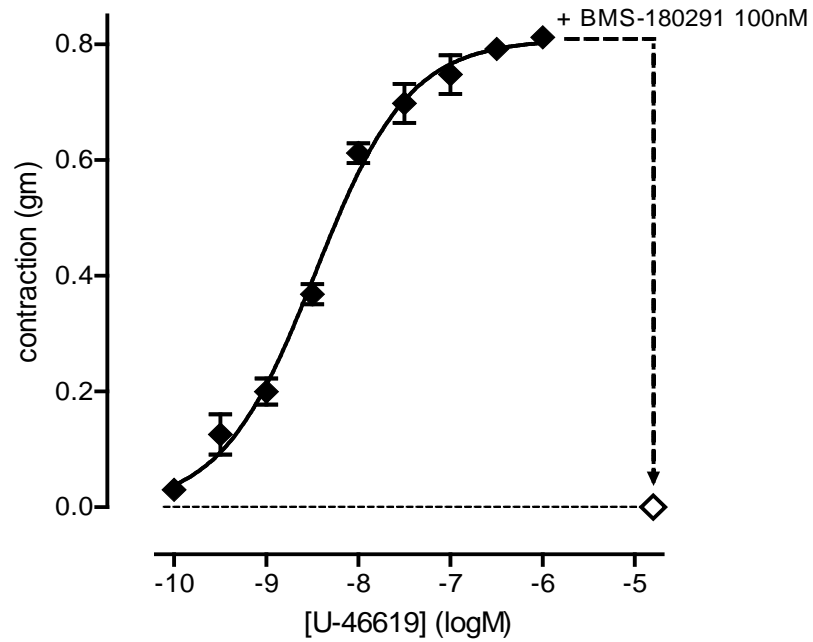
**Figure 4.5** Contractile activities of prostanoid agonists in rat mesentery artery. PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub> and sulprostone contracted the artery in a concentration dependent manner with different potency ( $n = 7$ ). PGF<sub>2α</sub> (1 nM – 1 μM) did not contract the artery ( $n = 4$ ). There was no prostanoid antagonist present.



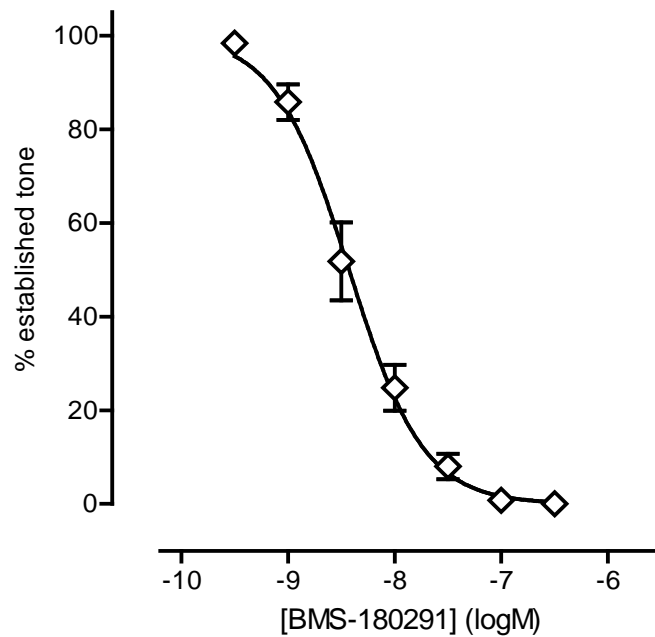
### 4.3.2 Effects of U-46619 and BMS-180291

As it was still unclear about the distribution of prostanoid receptor in the rat mesentery artery, the next step was to use U-46619 and BMS-180291 to investigate the presence of the TP receptor. U-46619 (selective TP agonist) produced a contractile response with maximal response of  $0.81 \pm 0.01$  g (Figure 4.6;  $n = 5$ ).

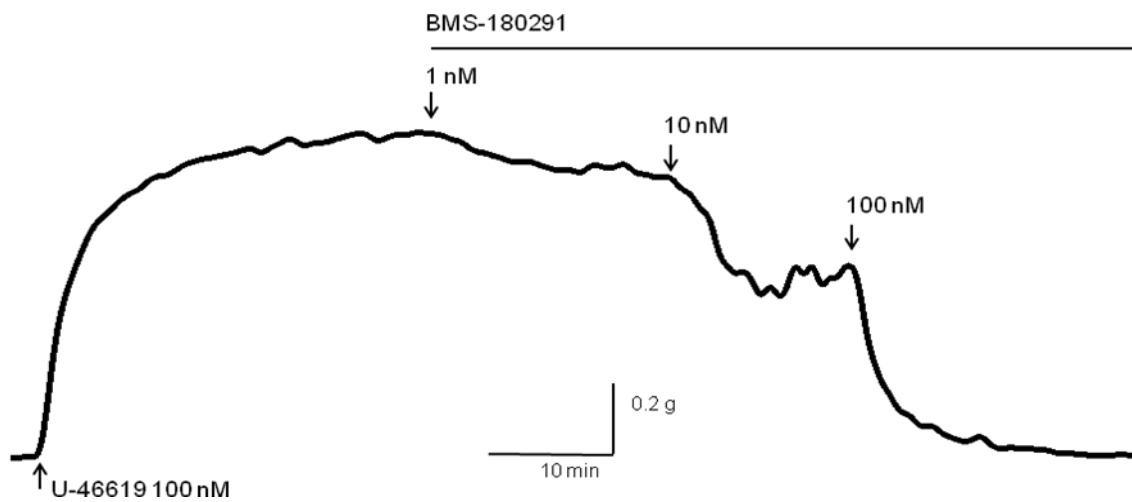
After the period of washout and the preparation had returned to resting tension, the arterial rings were precontracted with 100 nM U-46619, corresponding to  $EC_{90}$ . Once the contractile tone was stable and established, BMS-180291 was then added cumulatively (protocol in Figure 4.1B). BMS-180291 started to relax the arterial ring at 1 nM and completed relaxation was achieved at 100 nM (Figure 4.7,  $n = 5$ ). The antagonism of BMS-180291 on rat mesenteric artery was quick as shown in Figure 4.8. The  $pIC_{50}$  for BMS-180291 was  $8.31 \pm 0.14$ ; the dose-ratio for this level was 28 affording a  $pA_2$  of  $9.83 \pm 0.25$ . At this stage, it was concluded that TP receptors are present in the rat mesenteric artery. In further experiments, 100 nM BMS-180291 was added 30 min before the cumulative addition of other agonists; the expected dose-ratio is about 680.



**Figure 4.6** Contractile activity of U-46619 in rat mesenteric artery. U-46619 (0.1 nM – 1  $\mu$ M), a selective TP agonist contracted the artery in concentration dependent manner ( $n = 5$ ). 100 nM BMS-180291 completely antagonised the response.



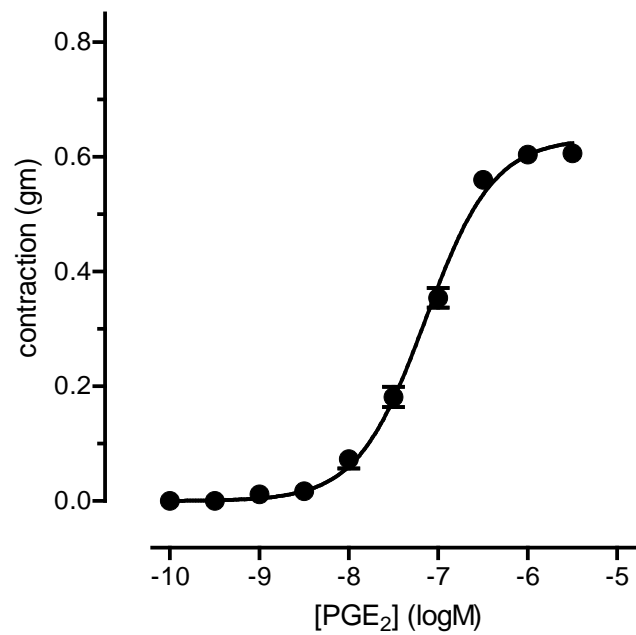
**Figure 4.7** Antagonism of 100 nM U-46619-induced contraction by BMS-180291 in rat mesenteric artery ( $n = 5$ ). After 40 - 45 min agonist contact, BMS-180291 (1 – 300 nM) was added cumulatively.



**Figure 4.8** Experimental trace illustrating the effect of BMS-180291 on 100 nM U-46619-induced contraction by in rat mesenteric artery. The inhibition by BMS-180291 was quick and at concentration of 100 nM, the response to U-46619 was abolished

### 4.3.3 Effects of prostaglandin E<sub>2</sub>

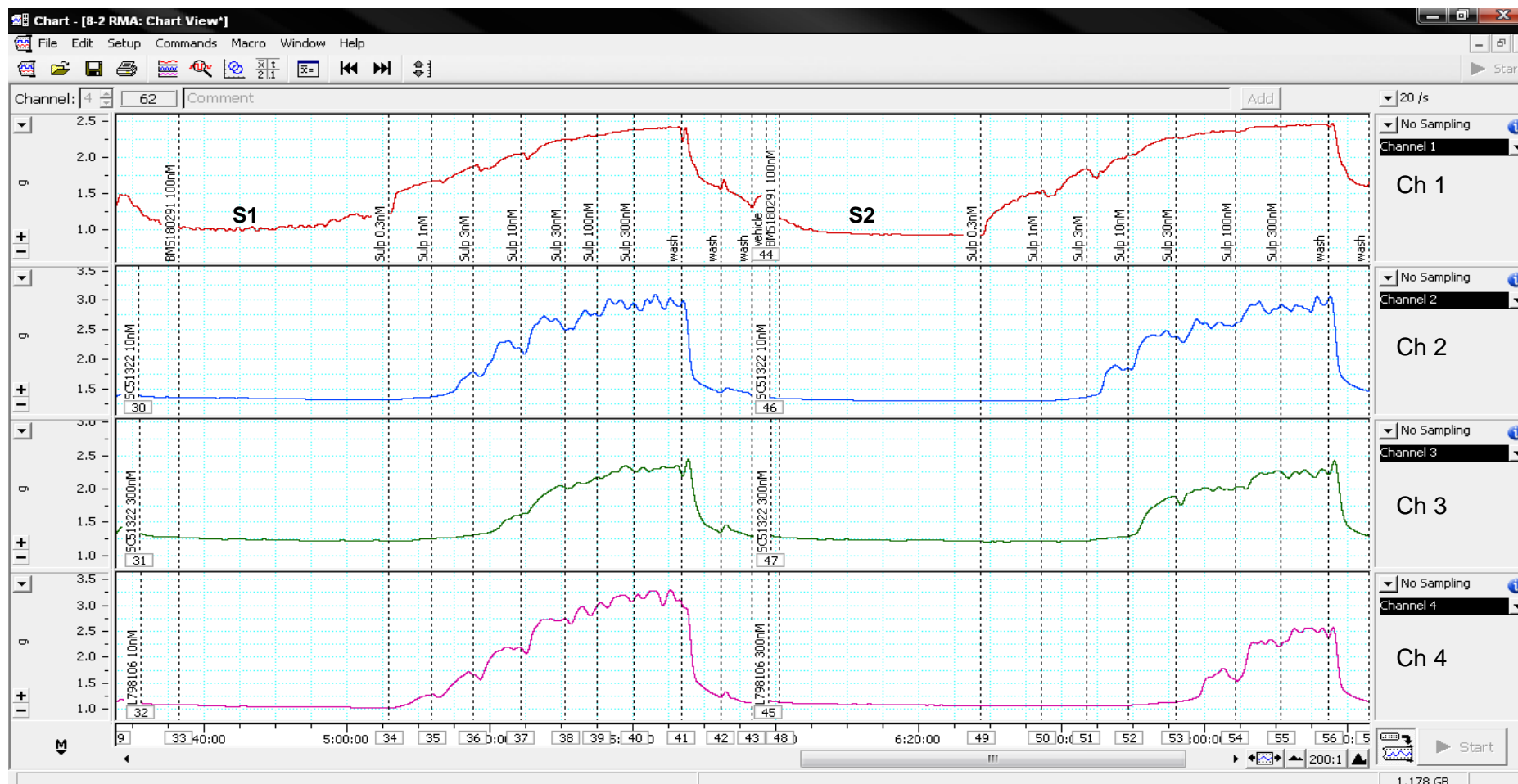
Initial investigation with PGE<sub>2</sub> in the presence of 100 nM BMS-180291 showed a contractile response in the rat mesenteric artery (Figure 4.9,  $n = 7$ ). The maximal response was  $0.60 \pm 0.01$  g with pEC<sub>50</sub> of  $7.14 \pm 0.06$ .



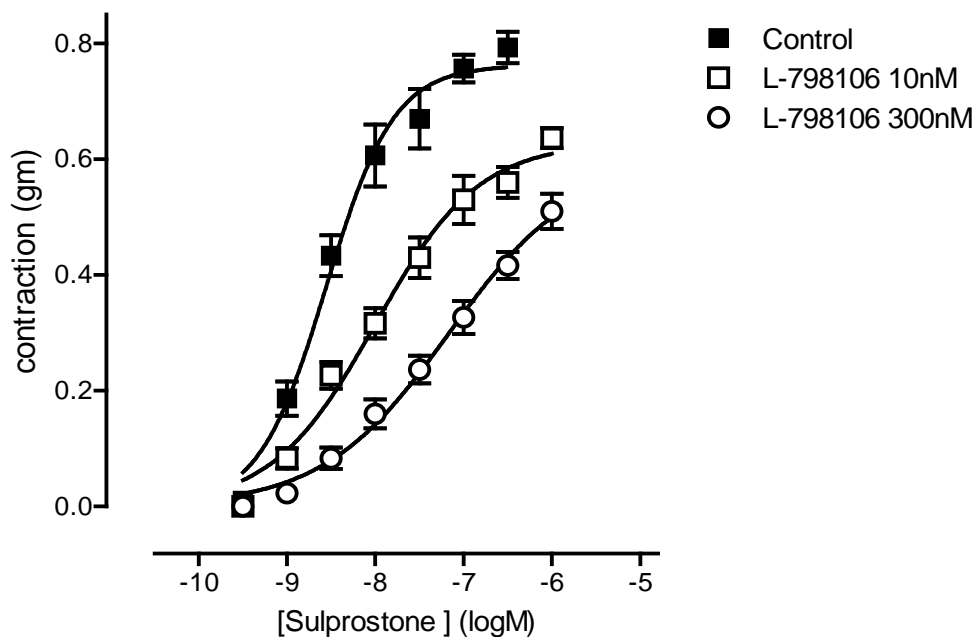
**Figure 4.9** Contractile activity of PGE<sub>2</sub> in rat mesenteric artery. PGE<sub>2</sub> (0.1 nM - 3  $\mu$ M) a non-selective agonist at EP receptors contracted the artery in concentration dependent manner ( $n = 7$ ). 100 nM BMS-180291 was present.

#### 4.3.4 Interaction of L-798106, L-826266 and SC-51322 with sulprostone

In the presence of BMS-180291 (100 nM), the EP<sub>3</sub> receptor antagonist L-798106 and EP<sub>1</sub> receptor antagonist SC-51322 were tested against sulprostone-induced contraction (Figure 4.10). L-798106 at 10 and 300 nM shifted the log concentration-response curve for sulprostone in non-parallel manner (Figure 4.11,  $n = 4$ ). This result clearly precludes calculation of a meaningful pA<sub>2</sub> value. However, in order to obtain some measure of antagonist affinity, dose-ratios were measured at the 25% response level for each L-798106 concentration, where a predominance of the EP<sub>3</sub> system is expected. The dose-ratios for 10 and 300 nM L-798106 were 3 and 14, giving pA<sub>2</sub> values of 8.30 and 7.65, respectively.



**Figure 4.10** Experimental records showing the effects of SC-51322 and L-798106 on sulprostone-induced contractile responses in the rat mesenteric artery. All antagonists were added 30 min before the addition of sulprostone. 100 nM BMS-180291 was present.



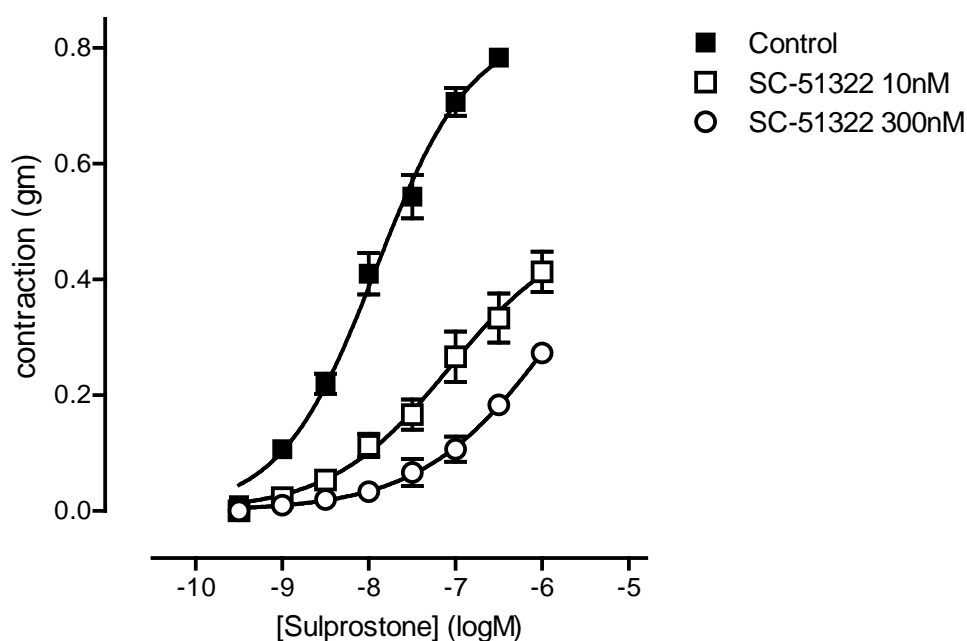
**Figure 4.11** Antagonism of sulprostone-induced contraction by L-798106 in rat mesenteric artery. Log concentration-response curves for sulprostone following treatment with 10 nM and 300 nM L-798106 are shown ( $n = 4$ ).

SC-51322 also shifted the log concentration-response curve of sulprostone in non-parallel manner (Figure 4.12,  $n = 3$ ). By the same reasoning, dose-ratios for 10 nM and 300 nM SC-51322 measured at the 25% response level were 71 and 148, with the estimated  $pA_2$  of 9.2 and 8.69, respectively.

Inhibition-curve protocols were also utilised to study the antagonist profile of L-798106 and L-826266. The arterial rings started to contract to sulprostone at 0.1 nM (Figure 4.13,  $n = 6$ ). The maximal response was  $0.76 \pm 0.02$  g with  $pEC_{50}$  of  $7.62 \pm 0.09$ . Established contraction to sulprostone (100 nM,  $EC_{80}$ ) was inhibited by L-798106 over the concentration range of 10 - 1000 nM. (Figure 4.14,  $n = 6$ ). However, relaxation was only  $56 \pm 9\%$  at 1  $\mu$ M. Given I am proposing that two receptor systems contribute to the contractile action of sulprostone (see discussion section), the use of the modified Cheng-Prusoff equations (section 4.2.2) to

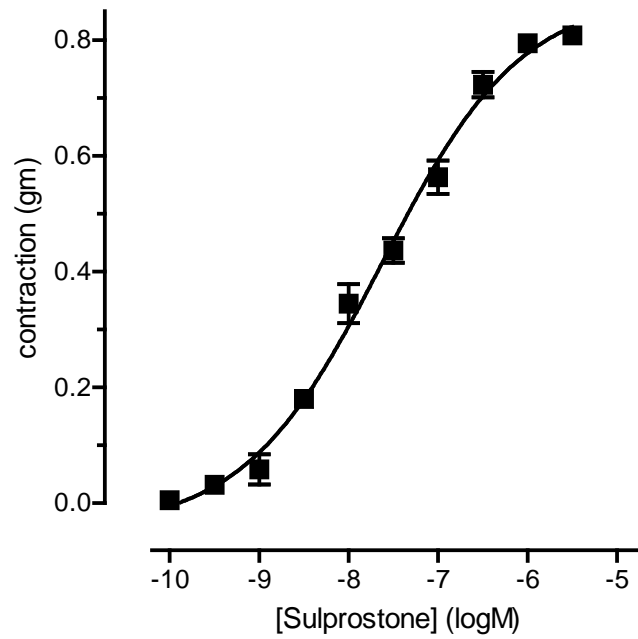
calculate a  $pA_2$  value from the dose-ratio corresponding to the  $IC_{50}$  value for L-798106 seems problematical. However, calculation of a dose-ratio for a modest inhibition (20%) by L-798106 ought to give a rough estimate of its affinity for the  $EP_3$  receptor. Thus, for 100 nM L-798106 in Figure 4.14 the dose-ratio is about 2.8, affording a  $pA_2$  of 7.41, which is comparable with the value from Figure 4.11.

L-826266 at 1  $\mu M$  also produced a similar partial inhibition of the established contraction to sulprostone. However, antagonism by L-826266 was slower than that for L-798106, with contact time of up to 2 hours required for steady state to be reached (see next section for similar response in 17-phenyl  $PGE_2$ ).

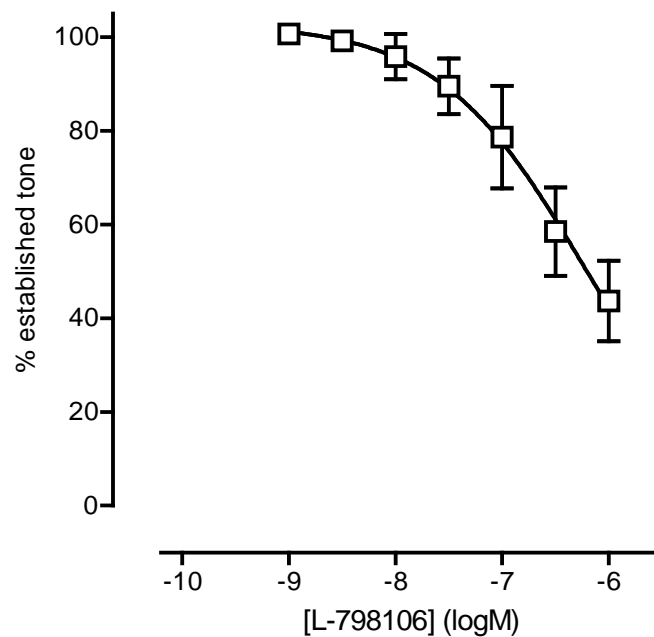


**Figure 4.12** Antagonism of sulprostone-induced contraction by SC-51322 in rat mesenteric artery. Log concentration response curves for sulprostone following treatment with 10 nM and 300 nM SC-51322 are shown ( $n = 3$ ).





**Figure 4.13** Contractile activity of sulprostone on rat mesenteric artery. Sulprostone (0.1 nM - 3  $\mu$ M) contracted the artery in concentration dependent manner ( $n = 6$ ). 100 nM BMS-180291 was present.



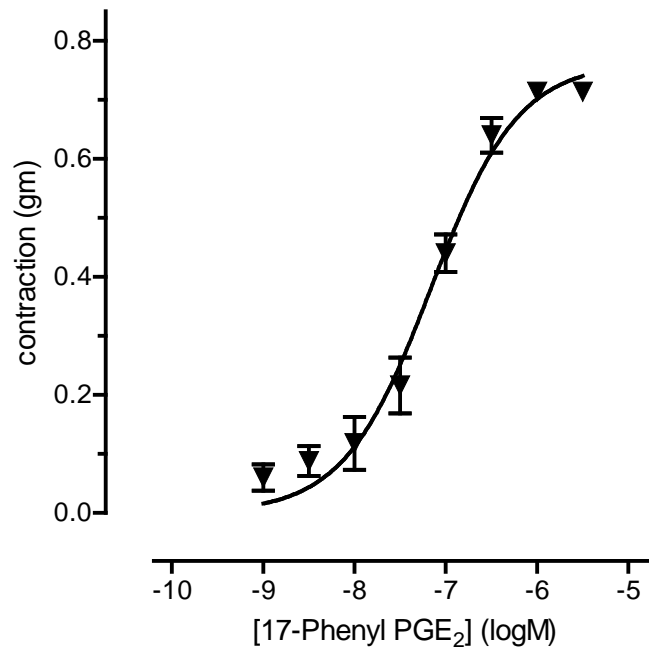
**Figure 4.14** Antagonism of sulprostone-induced tone by L-798106 in rat mesenteric artery ( $n = 6$ ). The arterial rings were precontracted with 100 nM sulprostone ( $EC_{80}$ ). L-798106 (1 nM - 1  $\mu$ M) was added cumulatively after 40-45 min. 100 nM BMS-180291 was present.

### 4.3.5 Interaction of 17-phenyl PGE<sub>2</sub> with SC-51322 and L-826266

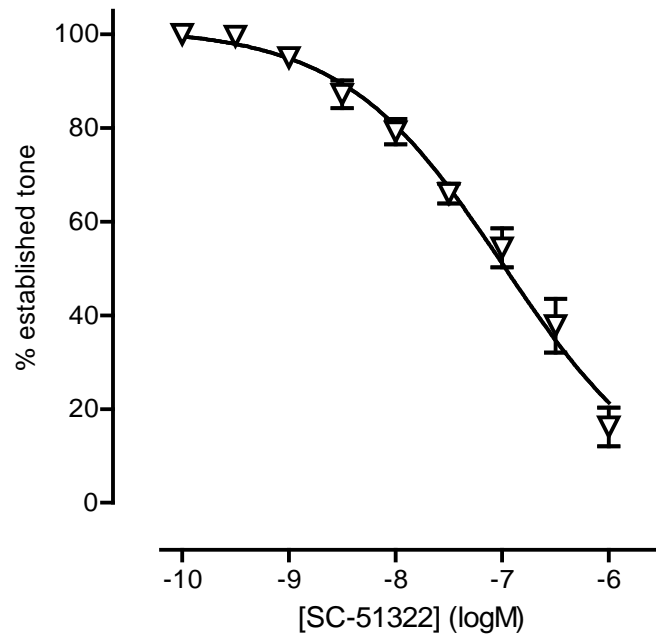
17-Phenyl PGE<sub>2</sub> (which has moderate EP<sub>1</sub> / EP<sub>3</sub> selectivity) was used in later studies on rat mesenteric artery. The experiments were done in the presence of 100 nM BMS-180291. 17-Phenyl PGE<sub>2</sub> contracted the arterial rings, with E<sub>max</sub> of 0.71 ± 0.02 g and pEC<sub>50</sub> of 7.06 ± 0.08 (Figure 4.15, *n* = 5).

Under the inhibition-curve protocol (Figure 4.1B), the arterial rings were contracted with 100 nM of 17-phenyl PGE<sub>2</sub>, corresponding to 80% of maximal response (EC<sub>80</sub>). SC-51322 started to relax the arterial ring at 1 nM (Figure 4.16, *n* = 5). At 1 μM, the maximal relaxation by SC-51322 was 84 ± 4%. Similar to sulprostone, the modified Cheng-Prusoff equations cannot be properly utilised in a two-receptor system. With reference to Figure 4.15, the dose-ratio at 25% relaxation was 2.0, affording a pA<sub>2</sub> of 7.80 for SC-51322.

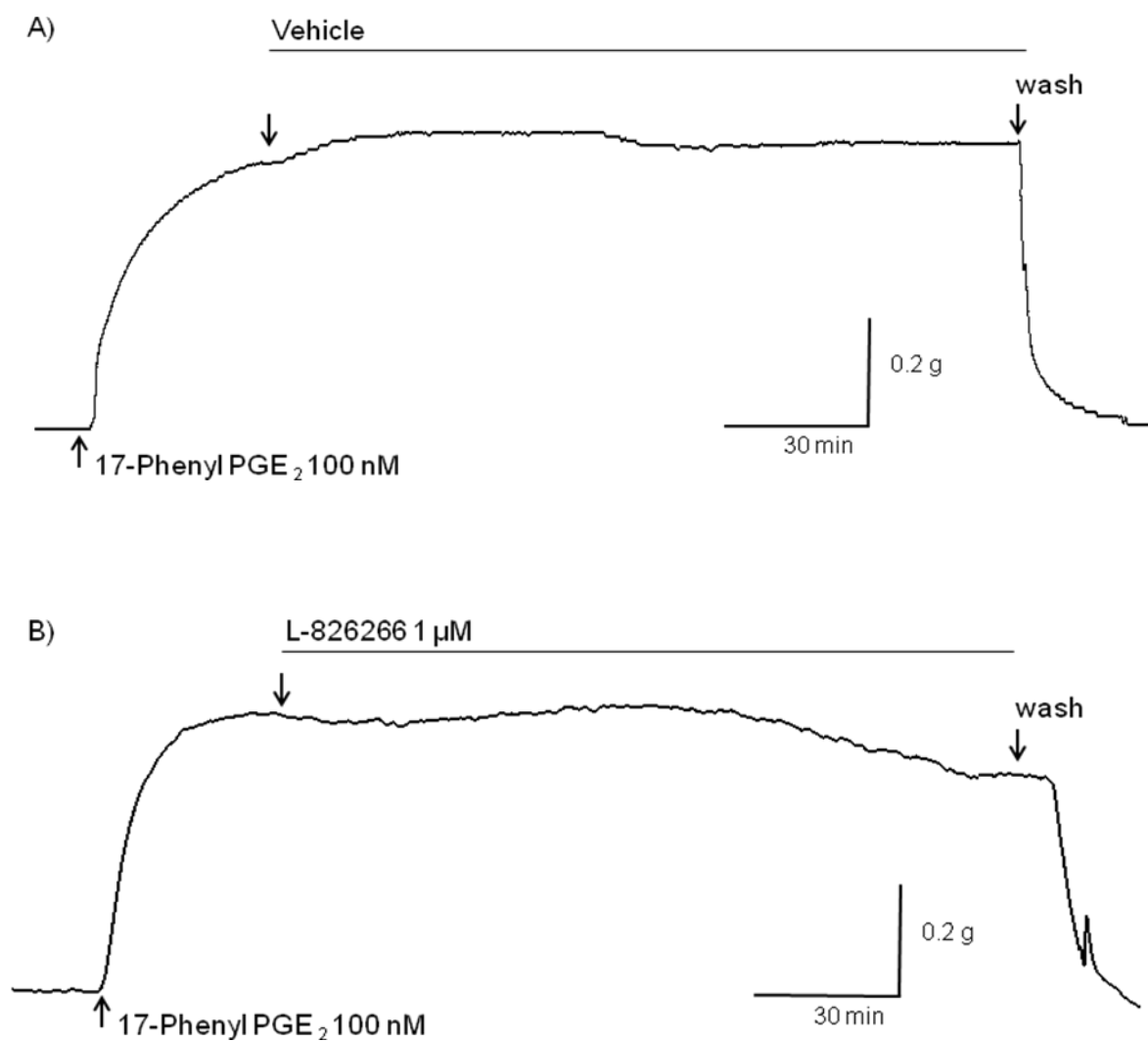
It has been observed that L-826266 was slow to reach equilibrium blockade as demonstrated in Figure 4.17. At 1 μM, L-826266 took approximately two hours to inhibit the established tone of 100 nM 17-phenyl PGE<sub>2</sub>. Therefore in inhibition-curve studies, each concentration (0.1, 0.3, 1.0 μM) was allowed 90 - 120 min contact (Figure 4.18, *n* = 5). L-826266 induced 30.2 ± 13.3% relaxation at 1 μM. Similarly, the modified Cheng-Prusoff cannot be utilised due to incomplete inhibition. The dose-ratio at 20% relaxation was 1.78, with pA<sub>2</sub> of 6.21 for L-826266.



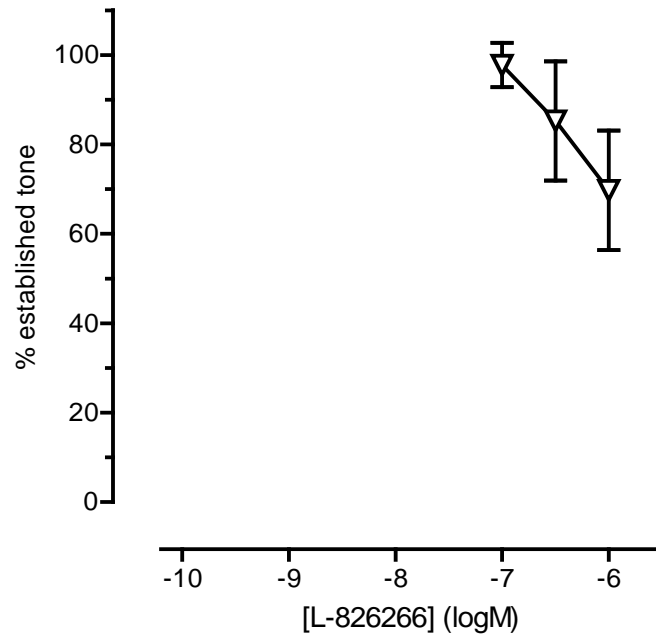
**Figure 4.15** Contractile activity of 17-phenyl PGE<sub>2</sub> on rat mesenteric artery ( $n = 5$ ). 17-Phenyl-PGE<sub>2</sub> (1 nM – 3  $\mu$ M) contracted the artery in concentration dependent manner. 100 nM BMS-180291 was present.



**Figure 4.16** Antagonism of 17-phenyl PGE<sub>2</sub>-induced contraction by SC-51322 in rat mesenteric artery ( $n = 5$ ). The artery was contracted with 100 nM of 17-phenyl-PGE<sub>2</sub> (EC<sub>80</sub>). SC-51322 (1 nM – 1  $\mu$ M) was added cumulatively after 40 - 45 min. 100 nM BMS-180291 was present.



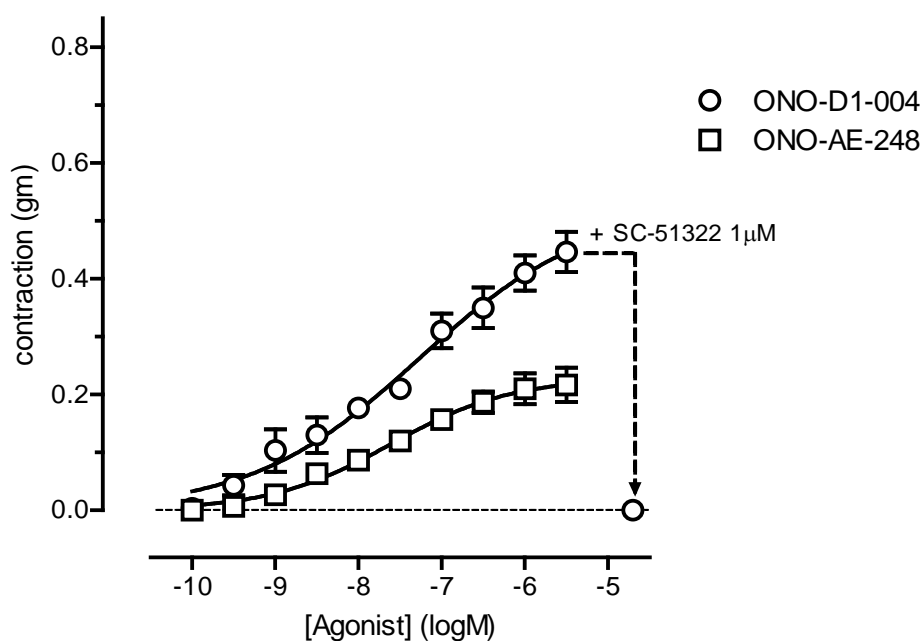
**Figure 4.17** Trace illustrating the effect of 1 μM L-826266 on 100 nM 17-phenyl PGE<sub>2</sub> - induced contraction in comparison with control (vehicle). 100 nM BMS-180291 was present.



**Figure 4.18** Antagonism of 17-phenyl PGE<sub>2</sub>-induced tone by L-826266 in rat mesenteric artery ( $n = 5$ ). The artery was contracted with 100 nM of 17-phenyl-PGE<sub>2</sub> (EC<sub>80</sub>). Once the tone was established, L-826266 (100 nM – 1  $\mu$ M) was added cumulatively. 100 nM BMS-180291 was present.

### 4.3.6 Effects of ONO-D1-004 and ONO-AE-248

ONO-D1-004, a selective EP<sub>1</sub> agonist, was tested on the arterial rings in the presence of 100 nM BMS-180291. A contractile response was seen with maximal response of  $0.50 \pm 0.01$  g at 3  $\mu$ M and pEC<sub>50</sub> of  $7.21 \pm 0.23$  (Figure 4.19,  $n = 4$ ). ONO-AE-248, a selective EP<sub>3</sub> agonist, produced a much weaker response; maximal response of  $0.21 \pm 0.01$  g at 3  $\mu$ M and pEC<sub>50</sub> of  $7.63 \pm 0.15$  (Figure 4.19,  $n = 4$ ). SC-51322 at 1  $\mu$ M completely inhibited the contractile response of ONO-D1-004. In the presence of 100 nM BMS-180291, L-798106 at 1  $\mu$ M did not have any effect on established contraction to ONO-AE-248.

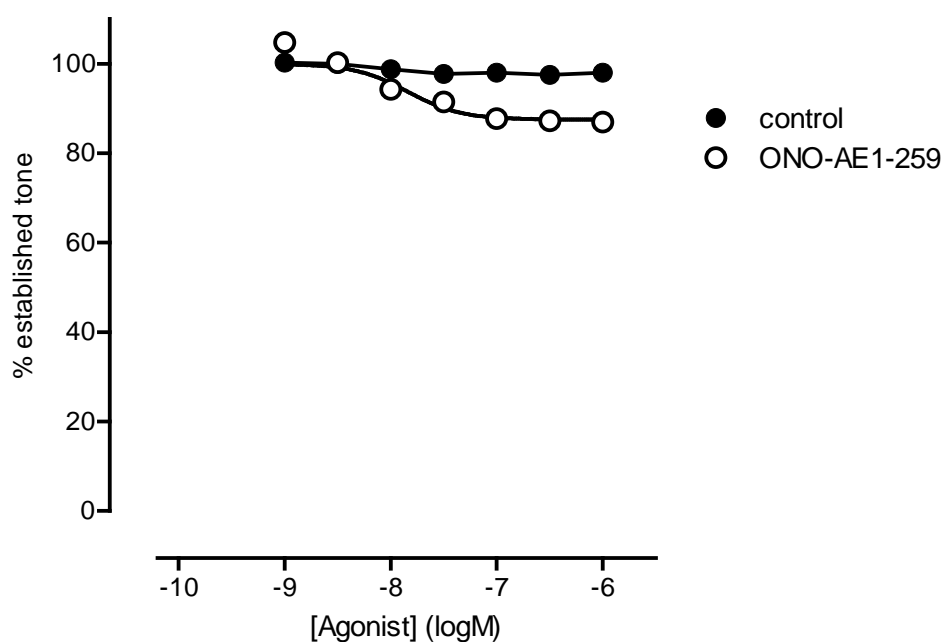


**Figure 4.19** Contractile activities of ONO-D1-004 and ONO-AE-248 on rat mesenteric artery ( $n = 4$ ). 1  $\mu$ M SC-51322 completely antagonised the response by ONO-D1-004. 100 nM BMS-180291 was present in all tests.

### 4.3.7 Relaxant studies

The arterial rings were precontracted with 300 nM phenylephrine in the presence of 1  $\mu$ M SC-51322, 1  $\mu$ M L-798106 and 100 nM BMS-180291. The selective EP<sub>2</sub> receptor agonist ONO-AE1-259 started to relax the rings at 10 nM (Figure 4.20,  $n = 4$ ). However, the relaxant responses were not significantly different compared with the control and no further relaxation was seen from 100 nM onwards. Butaprost-FA at the same concentration range also did not produce any relaxation ( $n = 4$ ; data not shown).

The IP agonist iloprost (1 – 300 nM) did not relax the arterial rings precontracted with 300 nM phenylephrine ( $n = 4$ ; data not shown).



**Figure 4.20** Relaxant activity of ONO-AE1-259 (1 nM – 1  $\mu$ M) in comparison with control in rat mesentery artery ( $n = 4$ ). The artery was precontracted with 300 nM phenylephrine. 100 nM BMS-180291, 1  $\mu$ M L-798106 and 1  $\mu$ M SC-51322 were present in all tests.



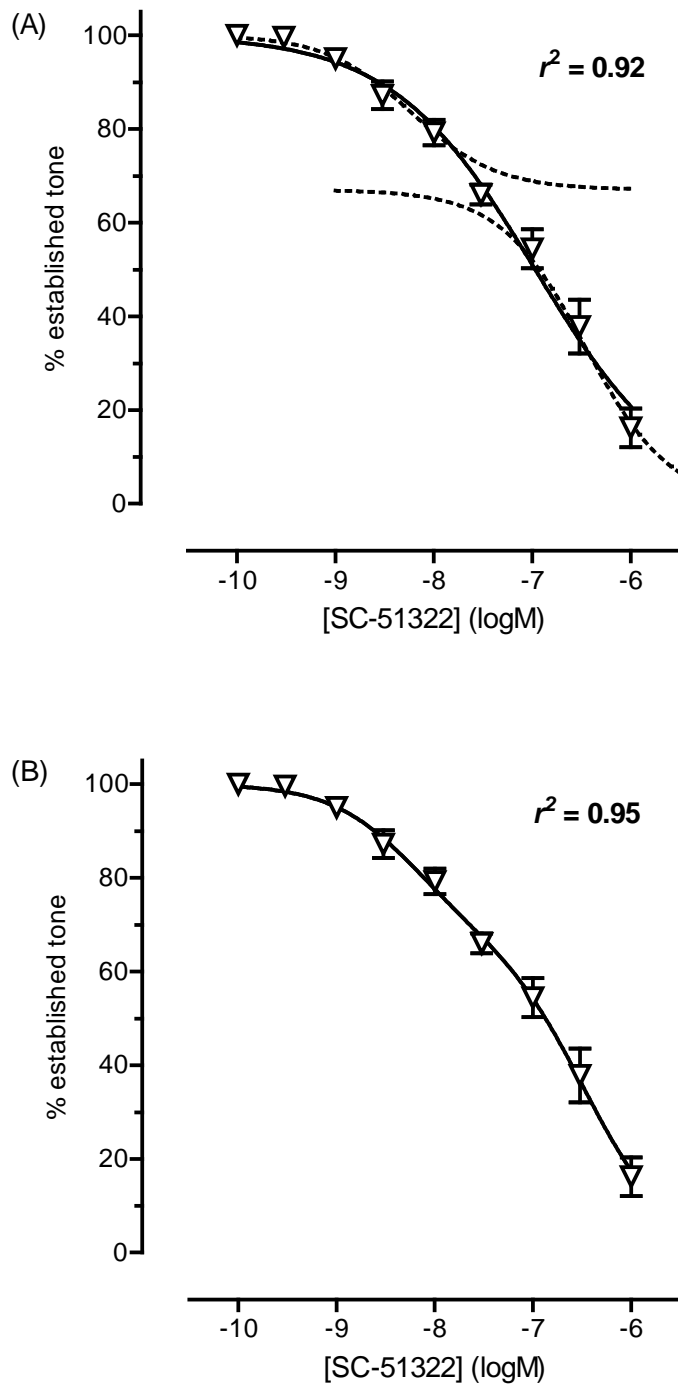
### 4.3.8 Two-site competition model

It appeared that there could be two components to the inhibition-curves for SC-51322 against 17-phenyl PGE<sub>2</sub>, as some of the data seems to deviate from the monotonic curve (Figure 4.16). Two theoretical curves were fitted in the inhibition curve (Figure 4.21A). Using the two-site competition equation in GraphPad Prism, the curve was refitted as described in Section 2.5.2. The data in graph of the two-site fit looks better (Figure 4.21B) than one-site model. The  $r^2$ -values were compared as described in Section 2.5.2. The  $r^2$ -value for one-site model is 0.92 and for two-site is 0.95.

The best-fit two-site curve has a high affinity site with a pIC<sub>50</sub> of 8.23 and a low affinity site with pIC<sub>50</sub> of 6.46 (Figure 4.21B). Based on the two theoretical curves, the fraction of high affinity sites is 33% with a 95% confidence interval that is reasonably narrow (20 – 45%). These results are acceptable; and it makes sense to compare the sum-of-squares of both models using the modified F test.

There are 45 data points to compare in 17-phenyl PGE<sub>2</sub> against SC-51322 curve. There are four parameters are fitted by the one-site equation fit and five parameters fitted for the two-site model. Using these data, the calculation is summarized in Table 4.2. The F ratio is 5.27 with 1 (numerator) and 40 (denominator) degrees of freedom. Using the F tables (see Appendix), the two-site model is significantly better than one-site model ( $p < 0.05$ ).

Despite no big difference between the  $r^2$ -values between both models, the F test result concluded that the two-side model fits the data significantly better than one-site model.



**Figure 4.21** The two-site competition model vs one-site model on SC-51322 and 17-phenyl PGE<sub>2</sub> inhibition curve (from data in Figure 4.16) using GraphPad Prism software. A) The one-site model with two theoretical curves superimposed. B) The two-site model fitted to the same data.

**Table 4.2** The F test for two-site model against one-site model of 17-phenyl PGE<sub>2</sub> vs. SC-51322.

<b>Model</b>	<b>SS</b>	<b>df</b>
Null hypothesis (1 site)	2139	41
Null hypothesis (2 sites)	1890	40
Difference	249	1
<b>Fractional difference</b>	0.1317	0.025
<b>Ratio (F)</b>	<b>5.27</b>	

#### **4.4 Discussion**

The superior rat mesenteric artery is part of a conductance system. Although resistance arteries present the greatest contribution to peripheral resistance, previous study has demonstrated the role of other conductance arteries, the femoral artery (Holloway and Bohr, 1973) and external iliac artery (Hansen and Bohr, 1975) in the control of blood pressure in the rat. The control of vascular tone in rat small artery is markedly similar to control in human artery of similar size (Hutri-Kahonen *et al.*, 1999). Thus, the effects of prostaglandins on the rat mesenteric artery may be relevant to the human artery.

The main interest of this study is to characterize prostanoid receptors that potentially control vascular tone in the rat mesenteric artery. It has been demonstrated that thromboxane A<sub>2</sub> (TXA<sub>2</sub>) plays a role in the artery through TP receptor. (Bolla *et al.*, 2004). In agreement, TXA<sub>2</sub> together with PGE<sub>2</sub> and prostacyclin (PGI<sub>2</sub>) were released from the pressurised artery in response to noradrenaline infusion (Soma *et al.*, 1996).

#### **4.4.1 Defining optimum conditions for studying prostanoid systems**

The rat mesentery exhibits a good viability over duration of the experiment, on average 8 hours. McIntyre *et al.* (2008) demonstrated that storage of the mesenteric artery in physiological salt solution (PSS) at 4° C preserved the artery response to noradrenaline up to two days of storage. In our study, we used the artery on the same day or after overnight storage in Krebs solution at 4 °C.

The use of isoprenaline in setting the resting tone of a smooth muscle preparation has been described previously for the guinea-pig trachea preparation (Farmer *et al.*, 1974). The guinea-pig trachea tube was exposed to 20 nM of isoprenaline to obtain the maximal relaxation of the preparation, during which time fluid was allowed to flow into the lumen. On re-sealing the system, a consistent level of intraluminal pressure was attained for the remainder of the experiment. Isoprenaline (1 nM – 10 µM) has been demonstrated to relax rat mesentery artery precontracted with KCl, with a maximal relaxation of  $66.8 \pm 2.4\%$  (Graves and Poston, 1993). In other studies, the relaxation with isoprenaline (10 nM – 100 µM) relaxed the 40 mM KCl precontracted artery to 80% of initial tone (Hutri-Kahonen *et al.*, 1999). In the current study, the arterial rings were exposed once to 100 nM isoprenaline for 10 min, followed by setting of the resting tone and then washout. Despite the absence of relaxation to isoprenaline in the majority of preparations, the resting tension was more stable and not prone to oscillate.

#### **4.4.2 The artery response to non-prostanoid agents**

Buus *et al.* (2000) demonstrated transient relaxation with exogenous KCl in ring preparations from Wistar rats at concentrations above 7 mM. The study was conducted in the presence of L-NOARG and indomethacin. In another study on rat mesenteric artery, KCl induced transient relaxation at concentration of 10 mM and above (Edwards *et al.*, 1998). Ba<sup>2+</sup> and oubain inhibited this transient relaxation. However, in the current study, transient relaxation

was not seen (only indomethacin 1  $\mu\text{M}$  was present in bath). Instead, the contractile response to exogenous KCl slowly faded at concentrations above 40 mM. An explanation for these different responses of KCl in the rat mesenteric artery is not readily apparent, although Edwards *et al.* (1998) has postulated  $\text{K}^+$  may act as an EDHF, through an increase in extracellular concentration of  $\text{K}^+$  causing hyperpolarisation and vasorelaxation of the artery. For this reason, KCl was not used as the contractile agent for the relaxant study.

In the current study, phenylephrine was demonstrated to produce a strong and sustained contractile response with a maximal response attained at a concentration of 10  $\mu\text{M}$ . The concentration of 300 nM (representing 35% of maximal response) was chosen to precontract the arterial rings in the relaxant study. Other workers have used 3  $\mu\text{M}$  as the precontractile dose (Edwards *et al.*, 1998).

#### **4.4.3 Defining the prostanoid receptors involved in the contractile response**

Table 4.3 listed the maximal responses and  $\text{pEC}_{50}$  values of the contractile prostanoid agonists examined on the rat mesenteric artery.

The TP analogue, U-46619 produced a concentration-dependent contractile response. This agreed with a previous study where U-46619 produced a stable contractile response and was used as a pre-contractile agent in a study of steroid hormone-induced relaxation on the rat mesenteric artery (Tsang *et al.*, 2003).

The selective TP receptor antagonist, BMS-180291 at 100 nM completely inhibited the U-46619 response in the current study. In previous studies in our laboratory, BMS-180291 in the low nanomolar range had a very slow onset on guinea-pig aorta and a cumulative

inhibition curve could not be obtained (unpublished observation by Jones RL). In comparison, the antagonism of BMS-189291 in rat mesenteric artery on U-46619 has a fast onset even at concentration of 1 nM as demonstrated in Figure 4.8. The difference may be due to the greater wall thickness of the aorta, such that diffusion through the extracellular space is markedly retarded by high affinity binding to TP receptors located on the cell surface (limited diffusion theory, Colquhoun *et al.*, 1972; Colquhoun and Ritchie, 1972).

Under the inhibition-curve protocol, the pA<sub>2</sub> for BMS-180291 against U-46619 was estimated to be 9.83 ± 0.25 for the rat mesenteric artery, which translates roughly into a dose-ratio of about 700 for 100 nM concentration. This is clearly sufficient to abolish any weak TP agonist activity of the EP agonists under study. The pA<sub>2</sub> estimated is comparable to pA<sub>2</sub> for BMS-180291 against U-46619 in rat aorta, reported as 9.5 (Zhang *et al.*, 1996) and 9.3 (Ogletree *et al.*, 1993). However the pA<sub>2</sub> reported in an early study in human platelets was 8.0 (Webb *et al.*, 1993). Since platelet-rich plasma (PRP) was used and BMS-180291 is a moderately lipophilic ligand, plasma protein binding will tend to reduce the estimate of its affinity.

**Table 4.3** pA<sub>2</sub> values for prostanoid receptor antagonists on rat mesenteric artery estimated from the inhibition-curve protocol.

Agonists	pEC <sub>50</sub>	E <sub>max</sub> (g)	Antagonists	pA <sub>2</sub>
U-46619	8.31 ± 0.14	0.81 ± 0.01	BMS-180291	9.83 ± 0.25
Sulprostone	7.62 ± 0.09	0.76 ± 0.02	L-798106	7.41 <sup>†</sup>
17-Phenyl PGE <sub>2</sub>	7.06 ± 0.08	0.71 ± 0.02	SC-51322	7.80 <sup>†</sup>
17-Phenyl PGE <sub>2</sub>	7.06 ± 0.08	0.71 ± 0.02	L-826266	6.21 <sup>†</sup>

\* 100 nM BMS-180291 was present in all experiments except when U-46619 was used as the agonist.

<sup>†</sup> Incomplete inhibition of established agonist response; pA<sub>2</sub> values are rough estimates from dose-ratios measured at the 20 - 25% inhibition level – see text.

PGF<sub>2α</sub>, a potent FP agonist did not produce any effect on the rat mesenteric artery, indicating the absence of FP receptors.

In initial experiments on the rat mesenteric artery, PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub> and sulprostone contracted the artery in the absence of any prostanoid antagonist. The order of potency was as follows: sulprostone > 17-phenyl PGE<sub>2</sub> = PGE<sub>2</sub>, which in the first instance suggests the presence of EP<sub>3</sub> and possibly also EP<sub>1</sub> receptors. Sulprostone is usually about 3 - 5 times more potent than PGE<sub>2</sub> on well established EP<sub>3</sub> preparations (e.g. guinea-pig vas deferens; Lawrence *et al.*, 1992). However, in this experiment, sulprostone was only about 1.5 times potent than PGE<sub>2</sub>. As expected, BMS-180291 did not affect the profiles of these three agonists.

Probably the most important observation in relation to the 'two EP receptors proposal' is the inability of the EP<sub>1</sub> antagonist SC-51322 and the EP<sub>3</sub> antagonist L-798106 to completely inhibit submaximal contractions elicited by 17-phenyl PGE<sub>2</sub> and sulprostone respectively. The rough estimate of the pA<sub>2</sub> for L-798106 under the inhibition-curve protocol was 7.41. The study by Clark *et al.* (2004) on EP<sub>3</sub> preparations using sulprostone as agonist, gave a pA<sub>2</sub> value of 7.82 in guinea-pig trachea and 7.42 in vas deferens of the same species. In rat femoral artery, L-798106 caused a parallel displacement of log concentration-response curve by sulprostone (Hung *et al.*, 2006) The pA<sub>2</sub> estimated in this EP<sub>3</sub> preparation was 7.43 - 8.03. Recently, L-798106 has been shown to inhibit PGE<sub>2</sub>-induced Ca<sup>2+</sup> flux in a rat recombinant assay EP<sub>3</sub> receptor, with pK<sub>i</sub> of 7.12 ± 0.04 (Jugus *et al.*, 2009). The pA<sub>2</sub> value obtained in the current study is comparable to the other 'pure' EP<sub>3</sub> preparation and suggests the presence of EP<sub>3</sub> receptor in the rat mesenteric artery.

Against 17-phenyl PGE<sub>2</sub>, the pA<sub>2</sub> value for SC-51322 estimated from the experiment was 7.80, which is somewhat lower than corresponding values from Schild protocol studies in EP<sub>1</sub> preparations (guinea-pig trachea, 8.45; Hung *et al.*, 2006). This could be due to species difference or the selectivity of 17-phenyl-PGE<sub>2</sub>, which can activate both EP<sub>1</sub> and EP<sub>3</sub> receptors in this case. Despite the high affinity for rat EP<sub>1</sub> receptors (rat hepatocytes, pIC<sub>50</sub> of

8.0; Kimura *et al.*, 2001), SC-51322 does have measurable affinity for the human recombinant EP<sub>3</sub> receptor ( $pK_i$  of 6.16; Abramovitz *et al.*, 2000). Moreover, the slope of the SC-51322 inhibition curve in Figure 4.16 is much less than the slope of the agonist curve for 17-phenyl PGE<sub>2</sub> (Figure 4.15); this is consistent with activation of two receptors by the fixed concentration of 17-phenyl PGE<sub>2</sub> and subsequent inhibition of both receptors by SC-51322, but with considerable difference in affinity. Fitting of the inhibition-curve for SC-51322 against 17-phenyl PGE<sub>2</sub> using the two-site competition equation in GraphPad Prism is shown in Figure 4.22. The Hill slopes of both sigmoidal components are automatically constrained to 1.0. The first phase of inhibition curve gave a  $pIC_{50}$  of 8.22 and the second phase a  $pIC_{50}$  of 6.45. Whether the two-component fit better than the one-component fit will be discussed in next section.

The slow onset of L-826266 may be attributed to its high lipophilicity, where the predicted *n*-octanol / water partition coefficient (ClogP) for L-826266 is 7.4 (ChemAxon software). In the current study, due to limited availability and slowly developing block of L-826266 (need 2-h contact before the inhibition is observed), only concentrations from 100 nM to 1  $\mu$ M were used. This slow inhibition against 17-phenyl PGE<sub>2</sub> (used as EP<sub>3</sub> agonist) in guinea-pig aorta, documented by Jones *et al.* (2008), afforded a  $pA_2$  of 7.58 after 3-h contact. In the current study, the estimated  $pA_2$  was 6.21, which is lower than in guinea-pig aorta.

The presence of two contractile EP receptors, EP<sub>1</sub> and EP<sub>3</sub> receptor in a vascular smooth muscle preparation has not often been reported. In large cerebral arteries of the adult pig, the contractile response to the prostanoid agonists used were attributed to the presence of both EP<sub>1</sub> and EP<sub>3</sub> receptors (Jadhav *et al.*, 2004). The EP<sub>1</sub> antagonist AH-6809 at 30  $\mu$ M and 100  $\mu$ M attenuated the PGE<sub>2</sub> contraction, with dose-ratios of 2.85 and 7.94 affording  $pA_2$  values of 4.78 and 4.84 respectively. However these values are considerably lower than  $pA_2$  values obtained in authentic EP<sub>1</sub> preparations. For example AH-6809 inhibited the contractile response to PGE<sub>2</sub> at 10  $\mu$ M in guinea-pig ileum with  $pA_2$  of 7.42 (Eglen and Whiting, 1988). Furthermore, 11-deoxy-16,16-dimethyl PGE<sub>2</sub> (DX-DM PGE<sub>2</sub>; a selective EP<sub>2</sub> / EP<sub>3</sub> agonist) has been shown by Jadhav *et al.* (2004) to be a potent contractile agonist in the porcine cerebral artery. The DX-DM PGE<sub>2</sub>-induced contraction was not inhibited by AH-6809 (3 –



300  $\mu\text{M}$ ), and the EP<sub>3</sub> receptor was concluded to be responsible for the response. However, the antagonist was tested against an established *supra-maximal* contraction to DX-DM PGE<sub>2</sub>, which casts doubt on the conclusions drawn.

Guinea-pig ileum also has a mixed EP<sub>1</sub> with other contractile EP receptors (Lawrence *et al.*, 1992). AH-6809 (0.2 - 5  $\mu\text{M}$ ) shifted the logCRC of the EP<sub>1</sub> agonist with pA<sub>2</sub> corresponding to its reported EP<sub>1</sub> affinity. The morphine / AH 6809-resistant contractile effect for sulprostone and other EP<sub>3</sub> agonists confirm the existence of a EP<sub>3</sub> receptor on the guinea-pig ileum.

The porcine cerebral artery study has however raised several issues related to ligand selectivity. The contractile response of DX-DM PGE<sub>2</sub> was attributed to EP<sub>3</sub> receptor activation despite no direct evidence documented for the present of functional EP<sub>3</sub> receptor in the artery. At higher concentration of EP receptor antagonists, the inhibition effect may not be specific. Thus, AH-6809 at 300  $\mu\text{M}$  completely blocked (3 of 6 preparations) the sulprostone-induced contractions. Given the rather low specificity of AH-6809 at high concentration (Keery and Lumley, 1988), it is not clear whether EP<sub>1</sub> or EP<sub>3</sub> receptors are involved. Moreover, could DX-DM PGE<sub>2</sub>-contraction in the cerebral artery be mediated by EP<sub>1</sub> receptor? DX-DM PGE<sub>2</sub> itself has been shown to have relatively high affinity for the EP<sub>1</sub> receptor in radio-ligand binding studies in HEK cells, with  $K_i$  of 3.8, normalised to PGE<sub>2</sub> (Ungrin *et al.*, 2001).

The involvement of EP<sub>1</sub> and EP<sub>3</sub> receptor has not been clearly defined in the functional studies. The coexistence of the two receptors in the porcine cerebral artery was concluded by the positive immunoreactivities and receptor bands in Western blot studies though the findings in the functional studies are not consistent. Thus, DX-DM-PGE<sub>2</sub> induced contractions can be due to other contractile prostanoid receptor despite being shown to have a potent contractile effect on EP<sub>3</sub> receptor in human respiratory tract smooth muscle (Karim *et al.*, 1980).

#### **4.4.4 The two-site competition model versus one-site competition model in relation to EP receptor-mediated contraction**

As discussed earlier, the study proposed the existence of two receptors mediating contractions to PGE<sub>2</sub> analogues. The graph of the one site fit model for SC-51322 against 17-phenyl PGE<sub>2</sub> seems to deviate systematically from the data point (Figure 4.16). The possibility of two contractile receptor systems is demonstrated by re-fitting the curve on Figure 4.16 using the two-site model in GraphPad Prism program (Figure 4.21A). The two-site curve-fitting procedure coupled with statistical evaluation by F test supported this proposal. In general, this procedure requires reproducible inhibition curves between individual preparations and a difference in pIC<sub>50</sub> values for the two components of at least 1.5 log units. In the current case, the difference was 1.77 log units. It would be of value to test another EP<sub>1</sub> antagonist with greater EP<sub>1</sub> / EP<sub>3</sub> selectivity than SC-51322. GW-848687 may serve this purpose; Giblin *et al.* (2007) reported > 400 fold selectivity for EP<sub>1</sub> over other EP receptors in human recombinant receptor assays.

#### **4.4.5 Utility of the novel EP agonists**

The specificities of ONO-DI-004 and ONO-AE-248 to the EP<sub>1</sub> and EP<sub>3</sub> receptor respectively, have been demonstrated in a study on intestinal smooth muscle preparations from EP knock-out mice (Okada *et al.*, 2000). However, the sensitivities of the EP<sub>1</sub> and EP<sub>3</sub> systems and the true potencies of these novel agonists in the rat mesenteric artery preparation have never been documented before. The maximal response (comparing at 3 μM) of ONO-D1-004 was significantly higher than ONO-AE-248 on rat mesenteric artery preparation, while their pEC<sub>50</sub> were similar for, at  $7.65 \pm 0.06$  and  $7.65 \pm 0.01$  respectively. On the basis that both compounds are full agonists the EP<sub>1</sub> system would appear to be the marginally dominant system in the rat mesenteric artery. In addition, ONO-D1-004 contractile response in rat mesenteric artery was 71% of the 17-phenyl PGE<sub>2</sub> response at similar maximum concentration (3 μM). Importantly, SC-51322 abolished contraction induced by ONO-DI-004, which argues strongly for ONO-DI-004 contracting the artery by solely activating EP<sub>1</sub>

receptors. Conversely, L-798106 (selective EP<sub>3</sub> antagonist) did not block the contraction induced by ONO-AE-248. This raised a question of specificity of ONO-AE-248 on EP<sub>3</sub> receptor. Further studies need to be done to elucidate the true sensitivities of these compounds in the rat mesenteric artery.

The previous study of the selectivity of these ONO compounds has produced a mixed results. The human pulmonary veins have been documented to have a TP and EP<sub>1</sub> contractile system (Walch *et al.*, 2001). ONO-D1-004 activity has been compared with sulprostone in the concentration range of 1 nM – 10 µM in this venous preparation (Norel *et al.*, 2004). However, ONO-D1-004 potency was very low and a pEC<sub>50</sub> was not calculable, while sulprostone-induced contraction was much more pronounced. Established responses to sulprostone were partially inhibited by the selective EP<sub>1</sub> antagonists, ONO-8711 and ONO-8713 at concentration of 30 µM and 10 µM, respectively. However, from the reported  $K_i$  values for these antagonists (1.7 and 0.3 nM respectively; Watanabe *et al.*, 1999) one would have expected abolition of sulprostone action. Finally, the same study also demonstrated the ability of ONO-AE-248 to contract the vein consistent with its moderate affinity for the EP<sub>3</sub> receptor ( $K_i$  value of 7.5 – 15 nM; Suzawa *et al.*, 2000). Collectively, these results suggest that the human pulmonary veins contains either an EP<sub>3</sub> system only or EP<sub>1</sub> and EP<sub>3</sub> systems, but not EP<sub>1</sub> system alone as implied by Norel *et al.* (2004).

#### **4.4.6 Relaxant EP and IP receptors**

Iloprost, a selective IP receptor agonist did not shown any relaxant effect on phenylephrine contraction. From radio-ligand binding studies, iloprost has some affinity for EP<sub>1</sub> and EP<sub>3</sub> receptors, with  $K_i$  of 21 and 22 nM, respectively (Kiriyaama *et al.*, 1997). Also functional studies indicate the high agonist potency of iloprost on EP<sub>1</sub> receptors (Dong *et al.*, 1986). However, the influence from both EP receptors has been eliminated, as the respective selective antagonists were added 30 min prior to the phenylephrine pre-contracted sequence (Figure 4.2).

ONO-AE1-259 has been shown to relax the non-pregnant porcine uterus selectively through EP<sub>2</sub> receptor activity (Cao *et al.*, 2002). In rat mesenteric artery, the activity of ONO-AE1-259 was not significantly different from the control. Butaprost-FA did not have any relaxant activity on the artery. Thus, evidence from the current study demonstrated the absence of relaxant EP<sub>2</sub> receptors in the rat mesenteric artery.

## 4.5 Conclusions

The current evidence has shown that only the contractile prostanoid receptors are present in the rat mesenteric artery. The receptors are TP, EP<sub>1</sub> and EP<sub>3</sub> receptors. Several PGE<sub>2</sub> analogues simultaneously activated both the EP<sub>1</sub> and EP<sub>3</sub> contractile receptor systems. Their inhibition curves could be fitted to a two-component inhibition equation and statistical testing showed that this fitting process was better than the one-component fit. Both receptor systems are likely to contribute to the contractile activity of the natural agonist PGE<sub>2</sub>, and activation of either alone appears insufficient to cause maximal contraction of the tissue.

The rank order of potency of EP agonists was sulprostone > 17-phenyl PGE<sub>2</sub> = PGE<sub>2</sub>.

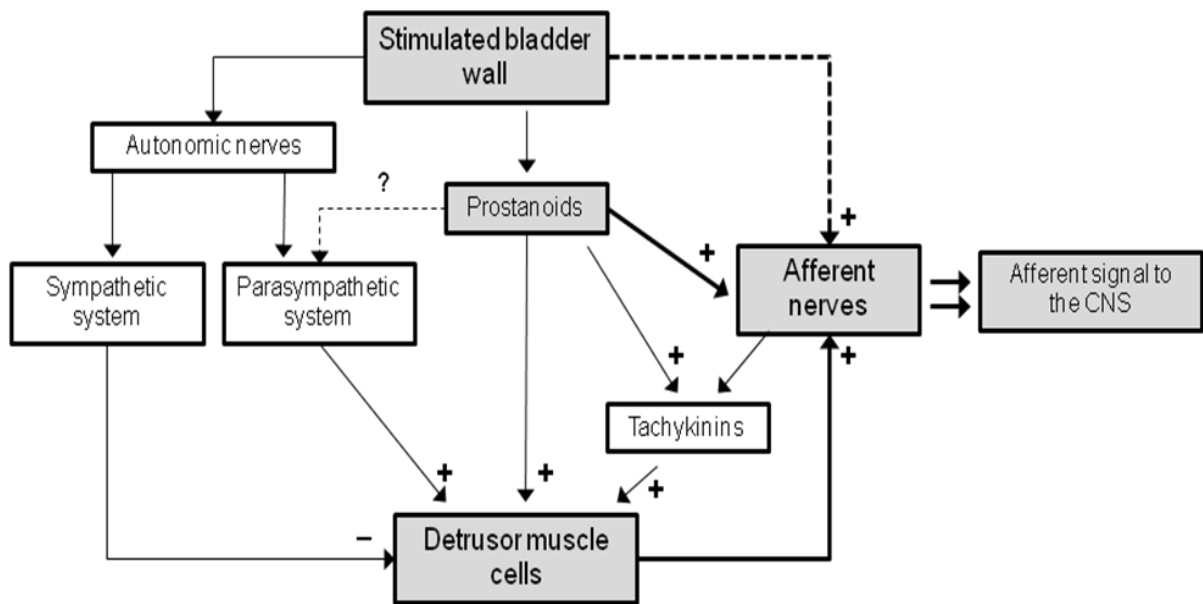
# **CHAPTER FIVE**

## **RAT URINARY BLADDER**

## 5.1 Introduction

The main function of the urinary bladder is to store and periodically release the urine into the urethra. The 'micturition reflex pathway' governs emptying of the bladder, which is voluntary in nature. There are various forms of bladder dysfunction; urgency and frequency are particularly common in older males and usually not controlled satisfactorily by drug administration. The control of the bladder involves interplay of neuronal and chemical factors. There are two main neuronal systems in the control of the bladder; the autonomic nervous system and somatic nerves (de Groat and Yoshimura, 2001; Andersson, 2002). The autonomic nervous system is responsible for the control of the bladder wall smooth muscle, whereas the somatic nerve is responsible for the control of the bladder neck sphincter.

Various chemical factors are involved in the control of the bladder wall, including histamine, neuropeptides and prostaglandins (prostanoids). The role of prostanoids is mainly to modulate the function and to maintain the basal tone of the urinary bladder (Ishizuka *et al.*, 1995; de Groat and Yoshimura, 2001; Schroder *et al.*, 2003). Prostanoids can affect the bladder wall function directly or indirectly. Prostanoids produce a contraction in the bladder directly through activation of prostanoid receptors on the smooth muscle cells. Prostanoids also have a potent stimulating action on sensory fibres (Schroder *et al.*, 2003). This will sensitise the afferent fibres of the bladder to natural stimuli. Intravesical infusion of PGE<sub>2</sub> in the conscious rat increased the levels of neurokinins, which in turn induced the micturition reflex (Ishizuka *et al.*, 1995). The interplay between neuronal and chemical controls of the bladder wall is summarized in Figure 5.1 (modified from Andersson, 2002).



**Figure 5.1** The neuronal and chemical controls of the urinary bladder, *CNS* – Central Nervous System (modified from Andersson, 2002).

It has been shown that various stimuli including mechanical stretching, inflammation and injuries to the bladder wall may initiate the synthesis and release of prostaglandins by the bladder wall (Ishizuka *et al.*, 1995; Park *et al.*, 1999; de Groat and Yoshimura, 2001; Wheeler *et al.*, 2001; Schroder *et al.*, 2003). Park *et al.* (1999) demonstrated the over-expression of COX-2 in the bladder smooth muscle cells of the Lewis rat after stretch-induced stimulation. COX-1 basal level remained unchanged. The study also demonstrated that NS-398 (COX-2 specific inhibitor) almost completely attenuated the increase in PGE<sub>2</sub> levels caused by stretching. Non-steroidal anti-inflammatory drugs (NSAIDs) also reduced prostaglandin levels and improved bladder function in the rat (Takagi-Matsumoto *et al.*, 2004).

The type of prostanoids and the relative amounts synthesized and released by the urinary bladder vary between species. In the rat urinary bladder, PGE<sub>2</sub>, 6-keto-PGF<sub>1α</sub> and TXB<sub>2</sub> form the major part of the basal PG activity generated in the tissue (Jeremy *et al.*, 1984; Kasakov and Vlaskovska, 1985). By comparison, PGI<sub>2</sub> (prostacyclin) is the major prostanoid released followed by PGE<sub>2</sub>, PGF<sub>2α</sub> and TXA<sub>2</sub> in the human bladder (Poli *et al.*, 1992; Khan *et al.*, 1998).

To my knowledge, the receptor subtype(s) that mediate the actions of prostanoids on rat bladder have not been well characterized. However, the presence of EP<sub>1</sub> receptor in rat bladder is provided by a study of the selective EP<sub>1</sub> receptor antagonist, SC-19220, which increased bladder capacity in normal rats (Maggi *et al.*, 1988). The amplitude of the micturition contraction was slightly but not significantly decreased by treatment with SC-19220. On isolated strips of rat detrusor muscle, SC-19220 also shifted the log concentration-response curve for PGE<sub>2</sub> to the right affording a pA<sub>2</sub> value of 4.4 ± 0.1, the value that is less than other recognized EP<sub>1</sub> systems.

In a rat model of bladder outlet obstruction (BOO), PGE<sub>2</sub> have been shown to increase bladder capacity, micturition volume and micturition interval without increasing residual urine (Lee *et al.*, 2007). The study also demonstrated that a selective EP<sub>1</sub> antagonist; PF-2907617-02 inhibited this effect of PGE<sub>2</sub> in a dose-dependent manner. In agreement, there



were no changes in bladder parameters after intravesical PGE<sub>2</sub> instillation following EP<sub>1</sub> gene deletion in BOO mice model (Schroder *et al.*, 2004). In contrast, there were significant changes in the micturition pattern in wild type (WT) mice in response to PGE<sub>2</sub>. These results suggested that under normal condition, endogenous PGE<sub>2</sub> is not involved in modulating bladder function. In the case of the stimulated bladder, PGE<sub>2</sub> level increased and produced an overactive bladder, mediated by EP<sub>1</sub> receptor.

The existence of the EP<sub>3</sub> receptor in rat urinary bladder was documented by qualitative PCR (Su *et al.*, 2008). The study also showed that the EP<sub>3</sub> receptor antagonist DG-041 inhibited rhythmic bladder contraction, which in-turn reduced the frequency of micturition. By doing so, the visceromotor reflex (VMR) response was prevented. DG-041 and CM9 (L-798106) also increased bladder capacity in the conscious rat (Jugus *et al.*, 2009). The same study also demonstrated that DG-041 and CM9 inhibit the Ca<sup>2+</sup> influx induced by PGE<sub>2</sub> through the EP<sub>3</sub> receptor with pK<sub>i</sub> of 7.57 and 7.12; respectively. In agreement, the study showed that the EP<sub>3</sub> receptor agonist GR-63799X reduced the efficacy of the voiding, with reduction of urine per void. The selectivity effect of GR63799X were demonstrated by work on EP<sub>3</sub> receptor knockout (EP<sub>3</sub> KO) mice (McCafferty *et al.*, 2008). Intravesical administration of GR-63799X produced overactive bladder in wild-type (WT) mice. This effect was absent in EP<sub>3</sub> KO mice. EP<sub>3</sub> KO mice also presented with higher bladder capacity compared with WT mice. This implied the EP<sub>3</sub> receptor is responsible for the bladder overactivity.

PGE<sub>2</sub> and its analogue sulprostone caused a strong urgency sensation and bladder instability in healthy women (Schussler, 1990). However, sulprostone activates both EP<sub>1</sub> and EP<sub>3</sub> prostanoid receptor subtypes and has direct and neuronally-mediated actions on smooth muscle (Coleman *et al.*, 1994b; Narumiya *et al.*, 1999). The instability caused by sulprostone in human bladder could be due to activation of EP<sub>1</sub> receptors alone rather than EP<sub>3</sub> receptors.

Cyclophosphamide treatment in rats induced bladder inflammation and hyperactivity which in turn increased the expression of COX-2 (Chuang *et al.*, 2008). The study also demonstrated the up-regulation of EP<sub>4</sub> expression in the bladder is paralleled with the elevation of COX-2 expression. The increased expression of COX-2 in inflammatory bladder could lead to increased production of PGE<sub>2</sub>, which might sensitise peripheral EP<sub>4</sub> receptors and produce bladder hyperactivity. The intravesicular administration of Botulinum toxin A, prevented the release of neurotransmitter and consequently decreased the inflammatory response. Concurrently, the expression of COX-2 and EP<sub>4</sub> receptor also decreased, accompanied by loss of the bladder hyperactivity. However, the exact mechanism by which the EP<sub>4</sub> receptor causes the hypersensitivity in cystitis was not elucidated in the study. It must be borne in mind that the EP<sub>4</sub> system is always inhibitory to smooth muscles, i.e. causes relaxation.

The main aim of the current experiments is to characterize functionally the prostanoid receptor(s) in the rat urinary bladder preparation, using selective agonist and antagonists. Agonists used included PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub> (moderate EP<sub>1</sub> / EP<sub>3</sub> selectivity), sulprostone (moderate EP<sub>3</sub> / EP<sub>1</sub> selectivity), U-46619 (TP-selective), butaprost-FA (EP<sub>2</sub> selective) and latanoprost-FA (FP-selective).

Table 5.1 lists pA<sub>2</sub> values of the selective antagonists used in the rat urinary bladder experiment. Selective receptor antagonists, particularly EP<sub>1</sub> antagonists, are crucial to achieve this aim. SC-51322, a well documented and potent EP<sub>1</sub> antagonist, with pA<sub>2</sub> of 8.45 in the guinea-pig trachea (Hung *et al.*, 2006), was used. In addition, GW-848687, a recently described EP<sub>1</sub> antagonist with higher affinity than previously developed agents was also used (Giblin *et al.*, 2007). In recombinant EP<sub>1</sub> receptor study, GW-848687 has been shown to be a competitive antagonist at the EP<sub>1</sub> receptor with pA<sub>2</sub> of 9.1 (Giblin *et al.*, 2007). It also has 30-fold selectivity over the TP receptor and with very low activity on the other prostanoid receptors.

L-798106 was the only EP<sub>3</sub> antagonist available for this study. It shows high selectivity for EP<sub>3</sub> receptors based on ligand binding assays involving recombinant prostanoid receptors (Juteau *et al.*, 2001). In functional studies, it blocked the action of sulprostone with pA<sub>2</sub> values of 7.82 in guinea-pig trachea (Clarke *et al.*, 2004) and 7.43 – 8.03 in rat femoral artery (Hung *et al.*, 2006) (Table 5.1). Recently, L-798106 (CM9) have been shown to inhibit PGE<sub>2</sub>-induced Ca<sup>2+</sup> entry in cells of rat EP<sub>3</sub> receptors, with pK<sub>i</sub> value of 7.12 (Jugus *et al.*, 2009). BMS-180291 is the first choice antagonist to block TP receptors, owing to its high selectivity and affinity (pA<sub>2</sub> of 9.3 on rat aorta; Ogletree *et al.*, 1993).

However, during the period of this study, there was no selective agonist or antagonist available to assess the functional EP<sub>4</sub> receptor in the rat urinary bladder.

**Table 5.1** Antagonists and concentrations chosen for use in the rat urinary bladder experiments.

Antagonists	Reported pA <sub>2</sub>	Concentrations used in the current study	References
GW-848687	9.1	30 – 300 nM	Giblin <i>et al.</i> , 2007
SC-51322	8.45	30 nM – 1 μM	Hung <i>et al.</i> , 2006
L-798106	7.43 – 8.03  7.12 (rat)	30 nM - 1μM	Juteau <i>et al.</i> , 2001; Clarke <i>et al.</i> , 2004; Hung <i>et al.</i> , 2006  Jugus <i>et al.</i> , 2009
BMS-180291	9.3	100 nM	Ogletree <i>et al.</i> , 1993

## 5.2 Methods

The basic methodology used has been described in detail in Chapter 2. Specific methodological points not discussed previously are addressed below.

### 5.2.1 Setting up of preparations

The rat urinary bladders were obtained as described in Chapter 2. The resting tone was set at an optimal tension of 1 g, determined from preliminary experiments. Previous study also has documented 1 g as the optimal resting tension for this preparation (Maggi *et al.*, 1988; Schneider *et al.*, 2004). The four preparations were allowed to equilibrate for 1 h, and then exposed twice to 100 nM carbachol to assess the contractile function.

All subsequent sequences were done in the presence of 100 nM BMS-180291, incubated for 30 min before the sequence commenced. The preparations were allowed a further 30 - 40 min to equilibrate before exposure to 30 nM PGE<sub>2</sub> for 10 min (Chan and Jones, 2004). Preliminary experiments established that the response to cumulative addition of prostanoid agonists was improved and more reproducible following exposure to low-dose PGE<sub>2</sub>.

After washout, the preparations were allowed to stabilize for at least 30 min. The presence of the prostanoid receptors was assessed by performing the experimental protocol in the presence of various antagonists (Table 5.1). Basically, a concentration-response curve (CRC) was constructed by cumulative addition of the particular agonists. In the immediate sequences, the vehicle (serve as control) or antagonist with different concentration was added at least 30 min before the cumulative addition of the agonist (Figure 5.2A).

For assessment of antagonist potency, an inhibition-curve protocol was utilised as described in Chapter 2. Basically, the first sequence involved the construction of agonist curve in absence of antagonist of interest. After washout and stabilization, the preparation was precontracted with the fixed concentration of agonist at 80% of maximal contraction ( $EC_{80}$ ). Antagonist was then added cumulatively to the precontracted bladder strips (Figure 5.2B and 5.3). Preparation without the antagonist served as a control.

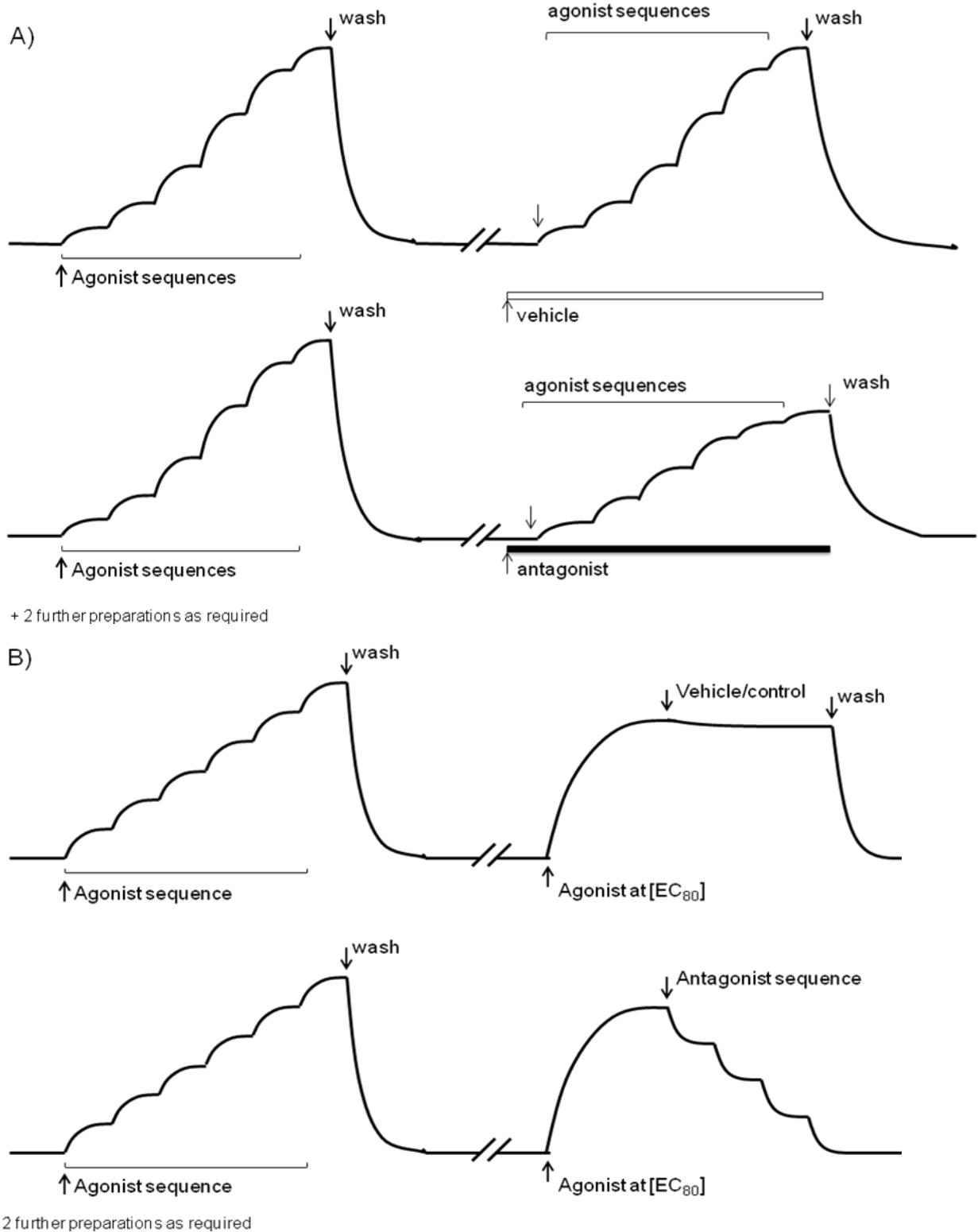
### 5.2.2 Antagonist protocol and $pA_2$ estimation

Antagonist affinity was determined by calculating the  $pA_2$ . In this study, the basic inhibition-curve protocol is used. The details have been described in Section 1.7.2. The  $pA_2$  is estimated from the equation (Lazareno and Birdsall, 1993b):

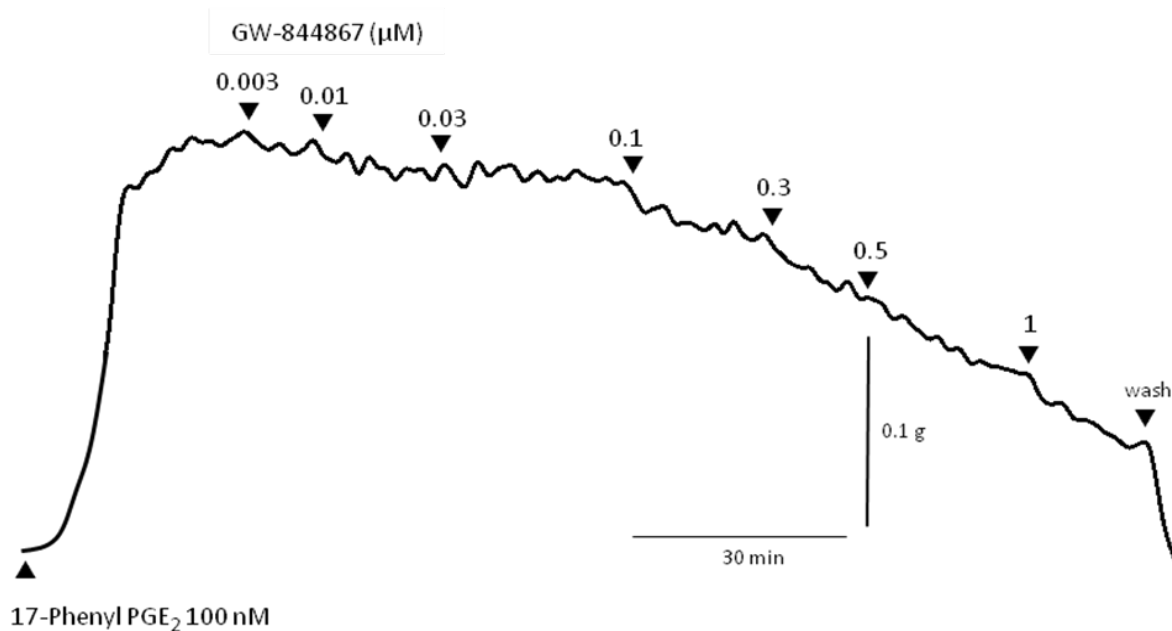
$$K_b = \frac{IC_{50}^*}{\frac{[A]}{EC_{50}^*} - 1}$$

and

$$K_b = \frac{[B_i]}{\frac{[A_f]}{[A_i]} - 1}$$



**Figure 5.2** Protocols used for rat urinary bladder to determine the profile of an antagonist and where appropriate its affinity constant. A) The basic protocol B) The inhibition-curve protocol.



**Figure 5.3** An experimental tracing of the effects of increasing concentrations of GW-844867 on the established response to 17-phenyl PGE<sub>2</sub> in rat urinary bladder, part of inhibition curve protocol (Figure 5.2B).

### 5.2.3 Statistical analysis

Contractile responses were measured as increases in tension (g) above the resting level and normalised to the second 100 nM carbachol response on each bladder preparation. Relaxant responses were expressed as a percentage of the established tone of the contractile agonist. A variable-slope sigmoidal curve was fitted to log concentration–response data using GraphPad Prism software; the bottom asymptote was constrained to zero for contraction and to 100% for relaxation. Sigmoidal curve parameters were derived from data for individual preparations. Data were further analysed by 1-factor and repeated measures 2-factor ANOVA combined with comparison of selected means by planned (orthogonal) contrasts using SuperANOVA software; all tests were two-tailed and the significance level was set at  $p < 0.05$ . Where applicable, data were analysed by the two-site competition equation using the modified F test in combination with the  $r^2$ -values comparison. All data are presented as mean  $\pm$  SEM.

## 5.3 Results

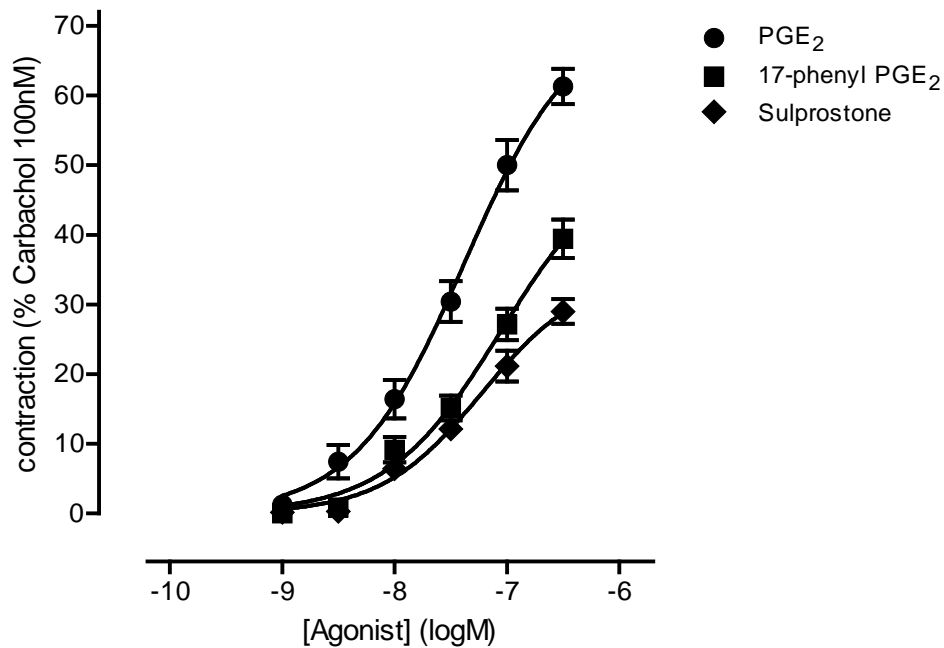
### 5.3.1 Initial investigations of rat urinary bladder strips

All experiments on rat urinary bladder strips were conducted in the presence of 1  $\mu$ M indomethacin to suppress prostanoid biosynthesis. Carbachol at 100 nM produced reproducible contractions, which did not fade. Histamine (50 – 100 nM) also contracted the bladder strips, but the response faded quickly. On the other hand, phenylephrine (10 – 500 nM) failed to produce any contractile response in the strips. In subsequent experiments, contractions were normalised to the response to the second 100 nM carbachol response obtained on each preparation.

The responses of the rat urinary bladder to cumulative addition of PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub> and sulprostone are shown in Figure 5.4, with a maximum concentration used of 300 nM ( $n = 7$ ). All three prostanoid agonists produced dose-dependent contractions. PGE<sub>2</sub> produced a higher response compared to other agonists at the maximal concentration of 300 nM ( $E_{300\text{nM}}$ :  $72 \pm 3\%$ ;  $pEC_{50}$  of  $7.41 \pm 0.08$ ,  $n = 7$ ), 17-Phenyl PGE<sub>2</sub> and sulprostone induced weaker response compared to PGE<sub>2</sub> ( $E_{300\text{nM}}$ :  $56 \pm 8\%$  and  $37 \pm 3\%$ ;  $pEC_{50}$  of  $7.29 \pm 0.08$  and  $7.13 \pm 0.13$ , respectively,  $n = 7$ ). The majority of responses to these contractile prostanoids were quick and sustained; slight fading was seen only in about one-fourth of preparations.

Cumulative addition of the selective TP agonist, U-46619 (1 nM – 1  $\mu$ M) produced no contractile response ( $n = 4$ ; data not shown). The addition of 100 nM BMS-180291 was without effect on established contraction to 100 nM PGE<sub>2</sub> ( $n = 4$ ; data not shown). In subsequent experiments, 100 nM BMS-180921 was routinely present (see Section 1.7.1).





**Figure 5.4** Concentration-response curves for PGE<sub>2</sub> and two of its analogues in rat urinary bladder ( $n = 7$ ).

Latanoprost-FA (100–300 nM) (FP selective agonist) failed to produce any response in the muscle strips ( $n = 4$ , data not shown). Cumulative addition of the selective EP<sub>2</sub> agonist butaprost-FA (100 – 300 nM) had no effect on the preparation precontracted with 80 nM carbachol (corresponding to EC<sub>60</sub> total response) ( $n = 4$ , data not shown). PGE<sub>2</sub> (100 - 300 nM) also did not relax the preparation precontracted with 80 nM carbachol (data not shown).

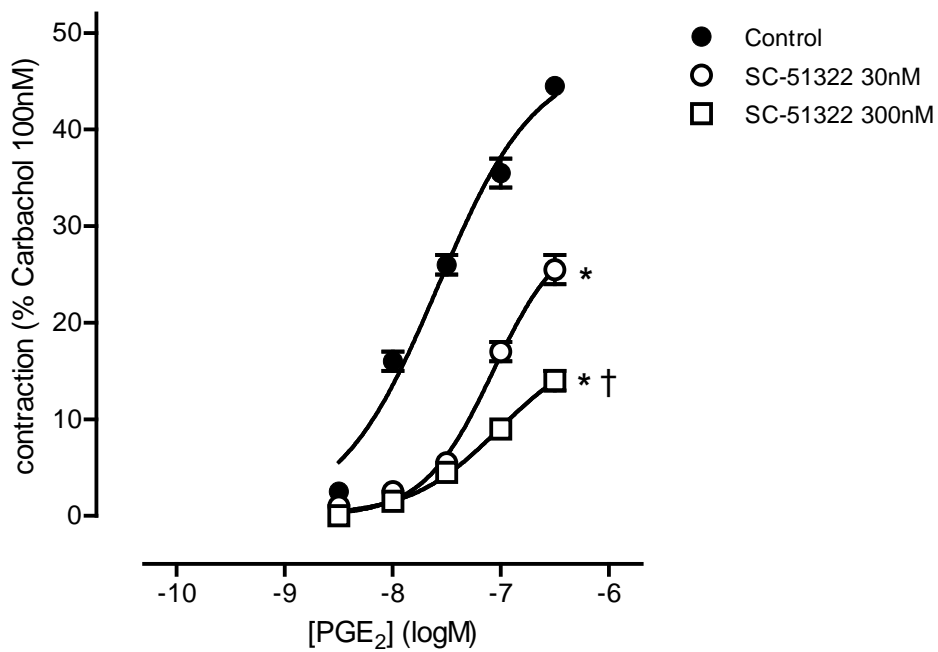
At this stage, it was concluded that relaxant EP<sub>2</sub> receptors, and probably EP<sub>4</sub> receptors, were not present in the rat urinary bladder, and that the contractile responses to prostanoids were only mediated by EP receptors. Further studies were performed to investigate the involvement of EP<sub>1</sub> and EP<sub>3</sub> receptors.

### 5.3.2 Effects of SC-51322 on contractile effects of prostanoid agonists

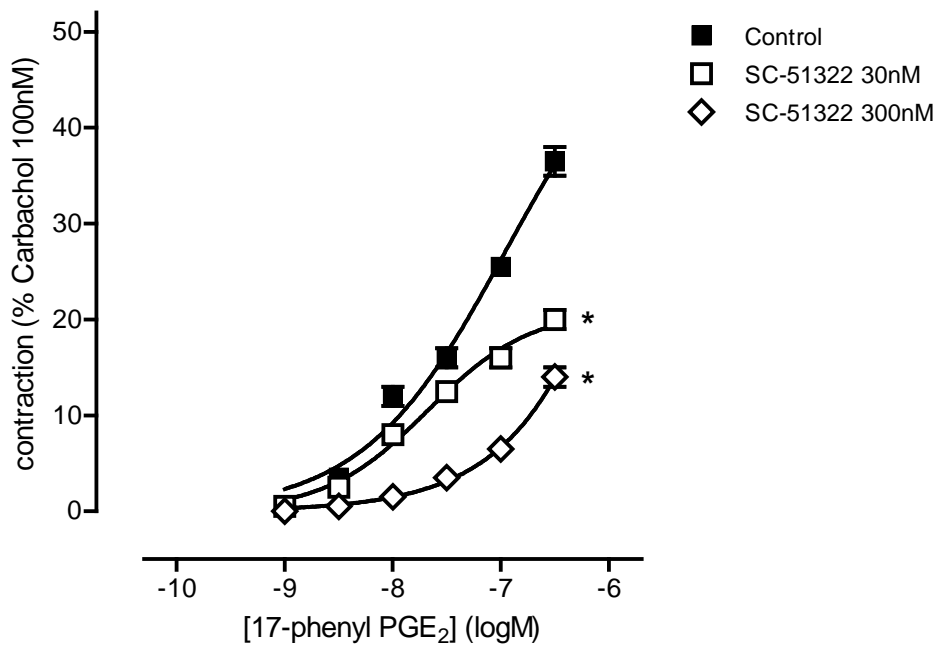
Pre-incubation of the bladder with 30 nM SC-51322 resulted in a roughly parallel right-shift of the log concentration-response curve for PGE<sub>2</sub> as shown in Figure 5.5 ( $n = 3$ ). The log interval of 1.02 (measured at EC<sub>25</sub>) afforded a pA<sub>2</sub> of 8.5. In contrast, the PGE<sub>2</sub> curve was displaced in a non-parallel manner by SC-51322 300 nM (DR comparison at EC<sub>30</sub> and EC<sub>10</sub>;  $p = 0.02$  with CI 95% of 7.41 – 40.15). There appeared to be a component of the PGE<sub>2</sub>-induced contraction that was (relatively) resistant to antagonism by SC-51322.

The CRC of 17-phenyl PGE<sub>2</sub> was constructed in the presence of SC-51322 at 30 and 300 nM (Figure 5.6,  $n = 3$ ). The curve was displaced to the right in non-parallel manner by 30 nM and 300 nM SC-51322. However, there was no significant difference between the effect of 30 nM and 300 nM SC-51322 on 17-phenyl PGE<sub>2</sub> curve ( $p = 0.65$ ).

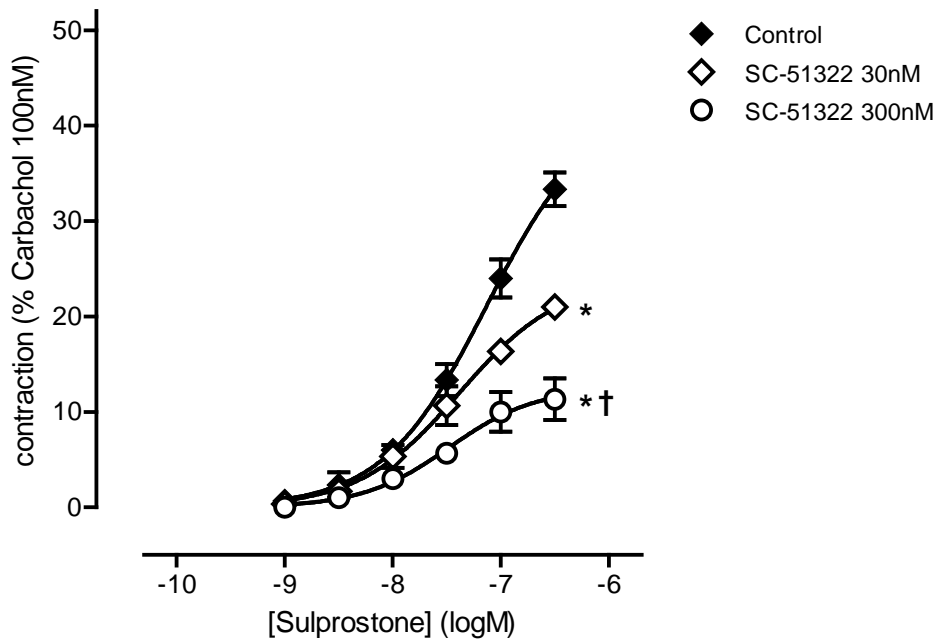
The pre-treatment of SC-51322 (30 and 300 nM) before cumulative addition of sulprostone significantly reduced the maximal contractile response ( $p = 0.001$ ) (Figure 5.7,  $n = 4$ ). Similarly with PGE<sub>2</sub> and 17-phenyl PGE<sub>2</sub>, the CRC for sulprostone is displaced non-parallel to the right. There were significant difference between sulprostone response to 30 nM and 300 nM SC-51322 as compared to control ( $p = 0.001$ , both). There was also significant difference in effect of pre-treatment with 30 nM and 300 nM SC-51322 ( $p = 0.01$ ).



**Figure 5.5** Antagonism of PGE<sub>2</sub>-induced contraction by SC-51322 in rat urinary bladder. Log concentration-response curves for PGE<sub>2</sub> following treatment with 30 nM and 300 nM SC-51322 are shown ( $n = 3$ ). \* $P < 0.01$  30 nM and 300 nM SC-51322 vs. control; † $P < 0.01$ , 300 nM SC-51322 vs. 30 nM SC-51322, using main effects contrasts.



**Figure 5.6** Antagonism of 17-phenyl PGE<sub>2</sub>-induced contraction by SC-51322 in rat urinary bladder. Log concentration-response curves for 17-phenyl PGE<sub>2</sub> following treatment with 30 nM and 300 nM SC-51322 are shown ( $n = 3$ ).  $*P < 0.01$ , 30 nM and 300 nM SC-51322 vs. control, using main effects contrasts.



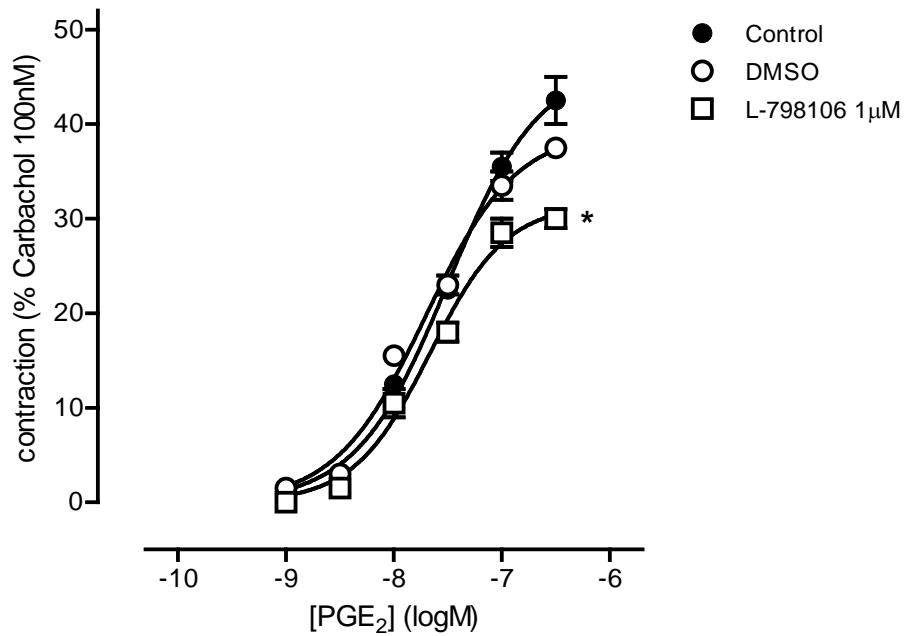
**Figure 5.7** Antagonism of sulprostone-induced contraction by SC-51322 in rat urinary bladder. Log concentration-response curves for sulprostone following treatment with 30 nM and 300 nM SC-51322 are shown ( $n = 4$ ).  $*P < 0.01$ , 30 nM and 300 nM SC-51322 vs. control;  $^{\dagger}P < 0.01$ , 300 nM SC-51322 vs. 30 nM SC-51322, using main effects contrasts.

### 5.3.3 Effects of L-798106 on contractile effects of prostanoid agonists

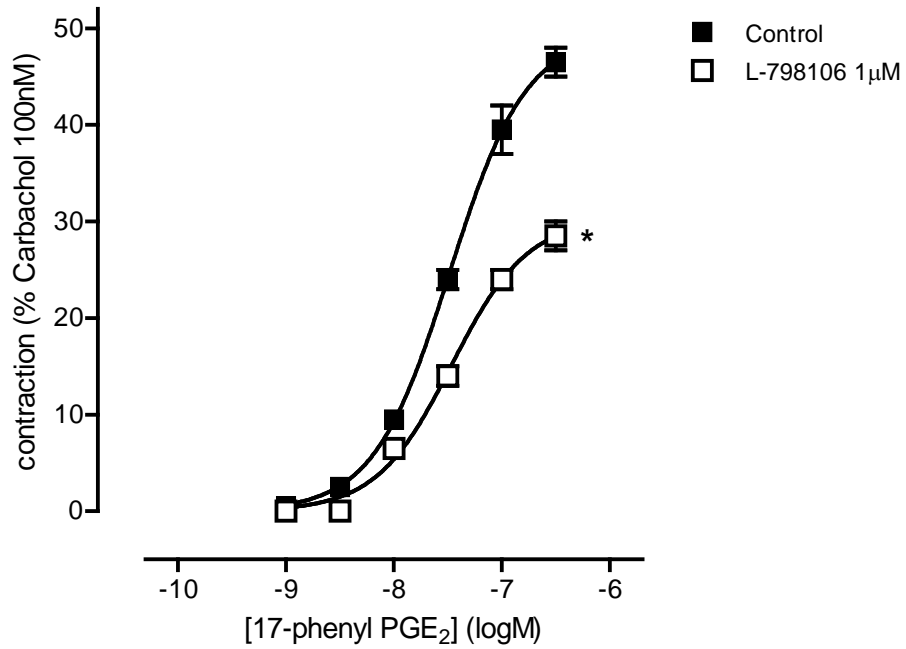
L-798106 is highly lipophilic and it was decided to make all dilutions from the stock with DMSO to be sure that the compound remained in solution. DMSO (at 14 mM) had no significant effect on the location of the PGE<sub>2</sub> curve, but may have slightly suppressed the maximum response (Figure 5.8,  $n = 3$ ). 1  $\mu$ M L-798106 significantly depressed the PGE<sub>2</sub> tone in control and in the presence of DMSO ( $p = 0.001$ ,  $p = 0.003$ ; respectively). However, there was no significant displacement of the curves.

1  $\mu$ M L-798106 significantly reduced the tone produced by 300 nM 17-phenyl PGE<sub>2</sub> ( $p = 0.001$ ) (Figure 5.9,  $n = 3$ ). The curve is displaced in non-parallel manner. The reduction of maximal response by 17-phenyl PGE<sub>2</sub> by 1  $\mu$ M L-798106 (62%) was similar as for reduction in PGE<sub>2</sub> tone (68%, Figure 5.8).

Pre-incubation of L-798106 at 1  $\mu$ M produced a significant suppression of maximal response by sulprostone ( $p = 0.001$ ) (Figure 5.10,  $n = 4$ ). The curve was displaced in non-parallel manner.

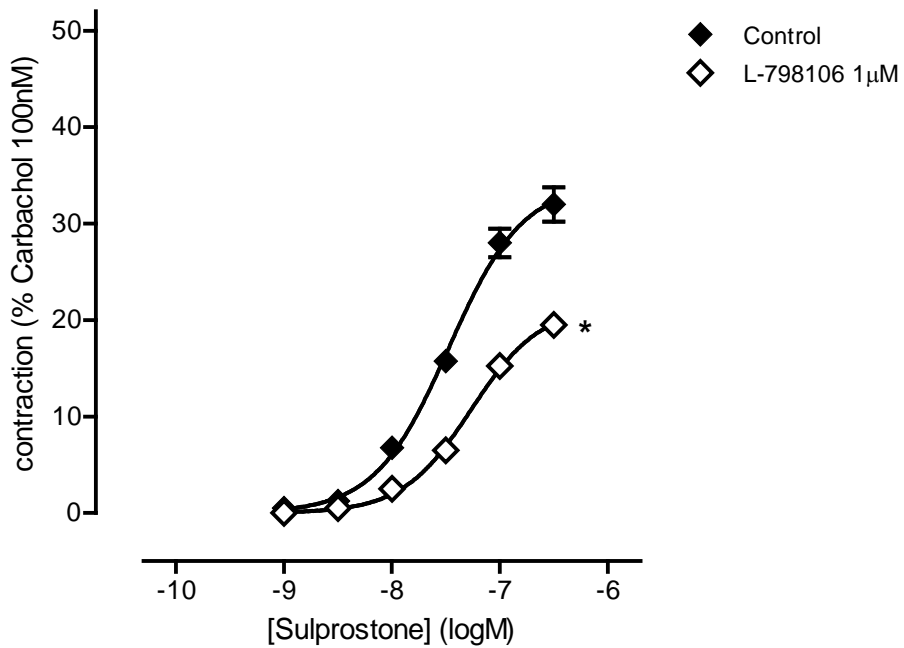


**Figure 5.8** Antagonism of PGE<sub>2</sub>-induced contraction by L-798106 in rat urinary bladder. Log concentration-response curves for PGE<sub>2</sub> following treatment with 1 µM L-798106 and DMSO are shown ( $n = 3$ ). \* $P < 0.01$ , 1 µM L-798106 vs. control and DMSO, using main effects contrasts.



**Figure 5.9** Antagonism of 17-phenyl PGE<sub>2</sub>-induced contraction by L-798106 in rat urinary bladder. Log concentration-response curve for 17-phenyl PGE<sub>2</sub> following treatment with 1 μM L-798106 is shown ( $n = 3$ ). \* $P < 0.01$ , 1 μM L-798106 vs. control, using main effects contrasts.





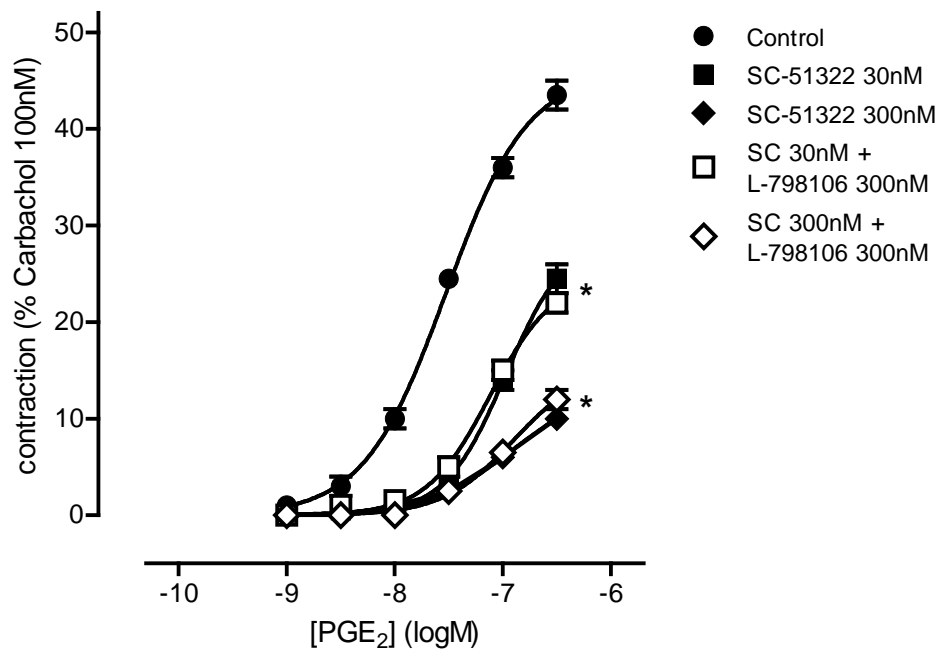
**Figure 5.10** Antagonism of sulprostone-induced contraction by L-798106 in rat urinary bladder. Log concentration-response curve for sulprostone following treatment with 1  $\mu$ M L-798106 is shown ( $n = 4$ ).  $*P < 0.01$ , 1  $\mu$ M L-798106 vs. control, using main effects contrasts.

### **5.3.4 Effects of a combination, of SC-51322 and L-798106 on contractile effects of prostanoid agonists**

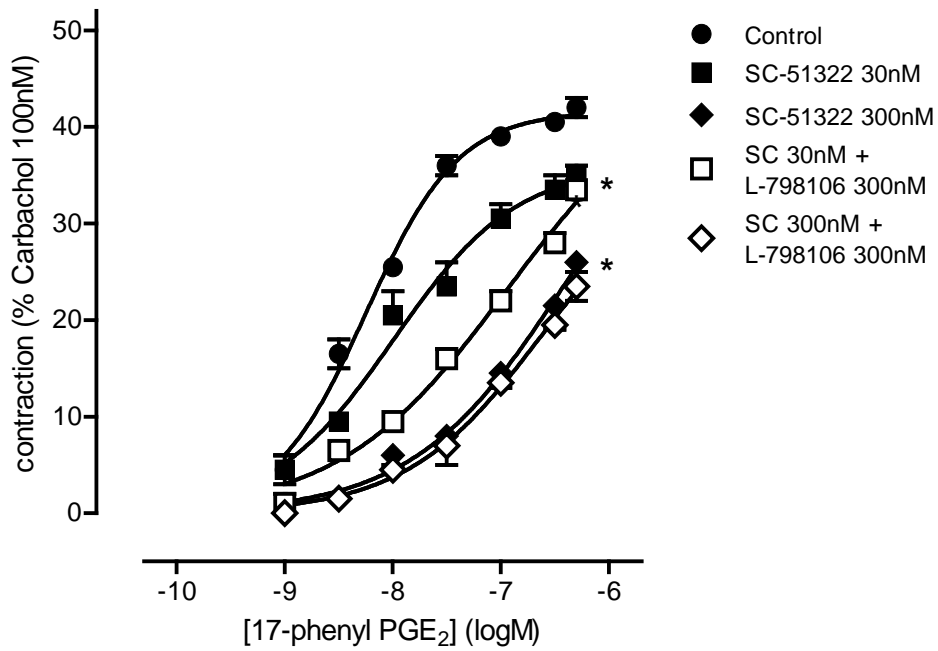
It was decided to perform further experiments in which pre-treatment with 300 nM L-798106 was added to 30 nM and 300 nM SC-51322. The combination of both drugs did not produce any significant changes to PGE<sub>2</sub> response in comparison with pre-treatment with SC-51322 alone as demonstrated in Figure 5.11 ( $n = 3$ ). Main effects analysis gave  $p$  values of 0.41 and 0.30 for comparison of 30 nM SC-51322 vs 30 nM SC-51322 / L-798106 and 300 nM SC-51322 vs 300 nM SC-51322 / L-798106 respectively.

For 17-phenyl PGE<sub>2</sub>, main effects analysis of 30 and 300 nM SC-51322 in combination with 300 nM L-798106 in comparison with control gave  $p$  values of 0.001 (Figure 5.12,  $n = 3$ ). In comparison to 300 nM SC-51322 alone, the addition of 300 nM L-798106 did not produce any significant changes. However, although the maximum response for 30 nM SC-51322 alone and in the presence of L-798106 was similar, there was a significant rightward displacement of the 17-phenyl PGE<sub>2</sub> curve ( $p = 0.001$  to  $0.004$  for cell-cell contrasts for 10, 30, 100 and 300 nM agonist).

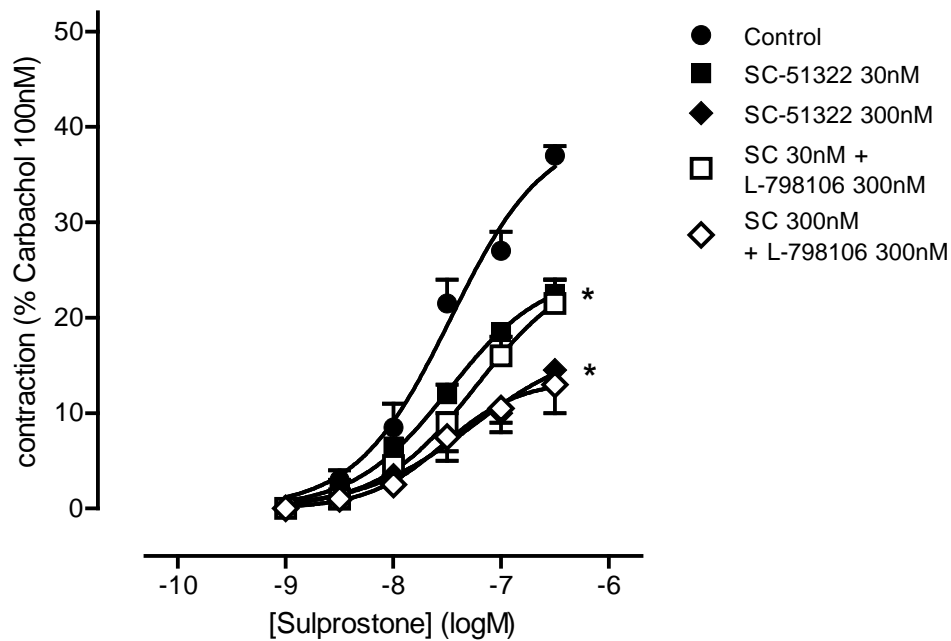
Similarly for sulprostone, the antagonist combination produced a similar finding as pre-treatment with SC-51322 alone (Figure 5.13,  $n = 3$ ). There were no significant changes in the curves between effects of 30 nM SC-51322 vs 30 nM SC-51322 / L-798106 and 300 nM SC-51322 vs 300 nM SC-51322 / L-798106 ( $p = 0.65$ ,  $p = 0.50$ ; respectively).



**Figure 5.11** Antagonism of PGE<sub>2</sub>-induced contraction by SC-51322 and L-798106 in rat urinary bladder. Log concentration-response curves for PGE<sub>2</sub> following treatment with 30 nM and 300 nM SC-51322 in the presence and absence of L-798106 300 nM are shown ( $n = 3$ ). \* $P < 0.01$ , 30 nM and 300 nM SC-51322 vs. control, using main effects contrasts.



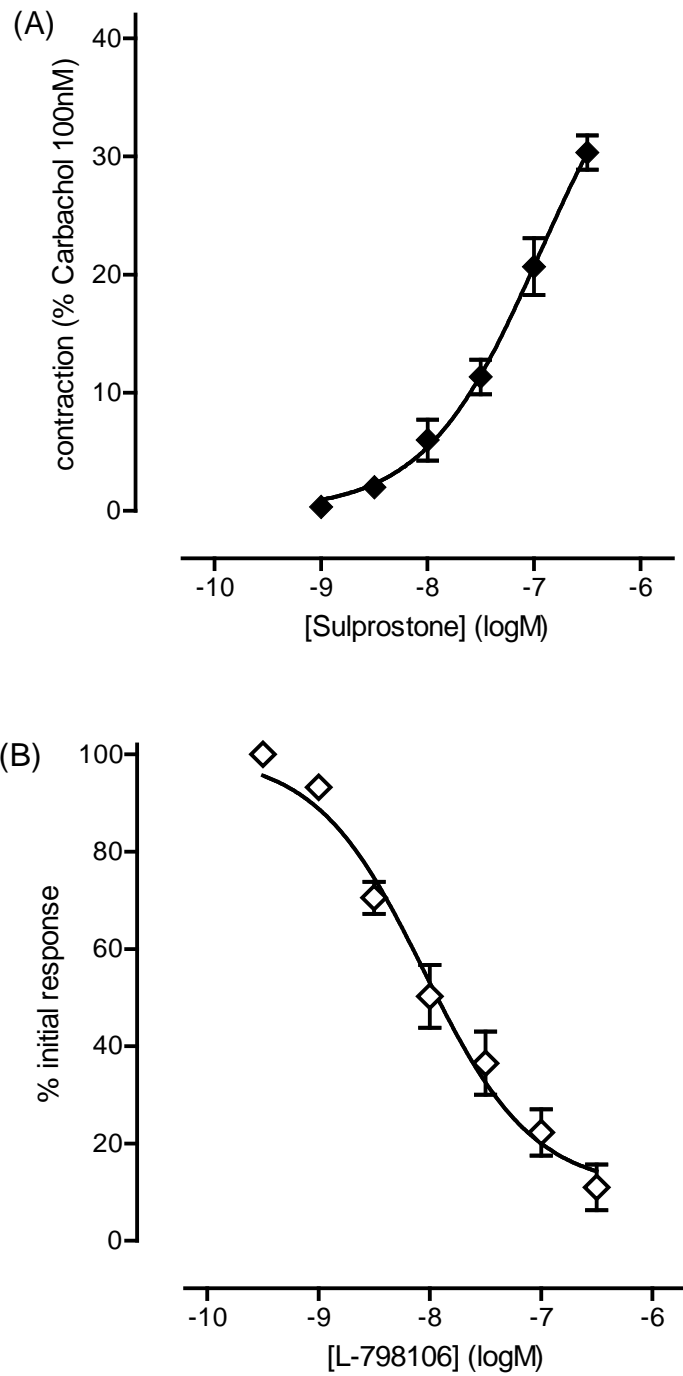
**Figure 5.12** Antagonism of 17-phenyl PGE<sub>2</sub>-induced contraction by SC-51322 and L-798106 in rat urinary bladder. Log concentration-response curves for 17-phenyl PGE<sub>2</sub> following treatment with 30 nM and 300 nM SC-51322 in the presence and absence of L-798106 300 nM are shown ( $n = 3$ ).  $*P < 0.01$ , 30 nM and 300 nM SC-51322 vs. control, using main effects contrasts.



**Figure 5.13** Antagonism of sulprostone-induced contraction by SC-51322 and L-798106 in rat urinary bladder. Log concentration-response curves for sulprostone following treatment with 30 nM and 300 nM SC-51322 in the presence and absence of L-798106 300 nM are shown ( $n = 3$ ).  $*P < 0.01$ , 30 nM and 300 nM SC-51322 vs. control, using main effects contrasts.

### **5.3.5 Antagonist potency of L-798106 under inhibition-curve protocol**

The CRC of sulprostone was constructed as part of inhibition-curve protocol to assess the potency of L-798106. The  $pEC_{50}^*$  estimated from the curve was  $7.44 \pm 0.06$  (Figure 5.14A,  $n = 4$ ). Cumulative addition of L-798106 relaxed the established tone by 100 nM sulprostone in dose-dependent manner; 87% inhibition was found at 300 nM (Figure 5.14B,  $n = 4$ ). L-798106 started to inhibit the tone at 1 nM. The inhibition was slow and took nearly 20 min to be noticeable and another 10 min before addition of the next dose. The  $pIC_{50}^*$  estimated from the inhibition curve was  $8.15 \pm 0.14$ . Calculated  $pA_2$  for L-798106 on sulprostone in the rat urinary bladder using equation described in section 5.2.4 is 8.35.

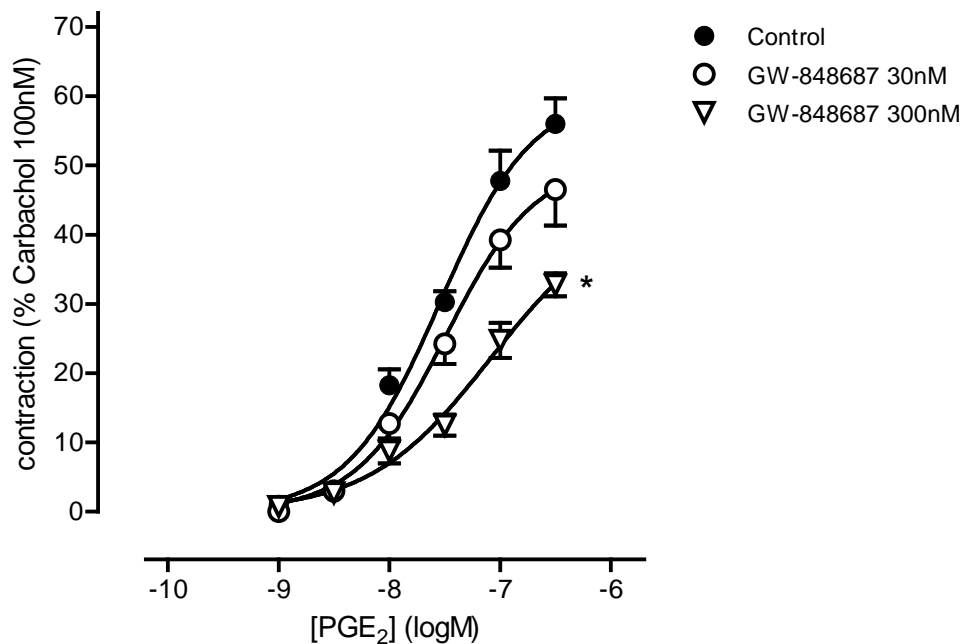


**Figure 5.14** The inhibition-curve protocol for sulprostone and L-798106 in the rat urinary bladder. (A) Log concentration curve for sulprostone ( $n = 4$ ). (B) Log concentration-inhibition curve for L-798106 using 100 nM sulprostone to establish tone ( $n = 4$ ).

### 5.3.6 Effects of GW-848687 on contractile effects of prostanoid agonists

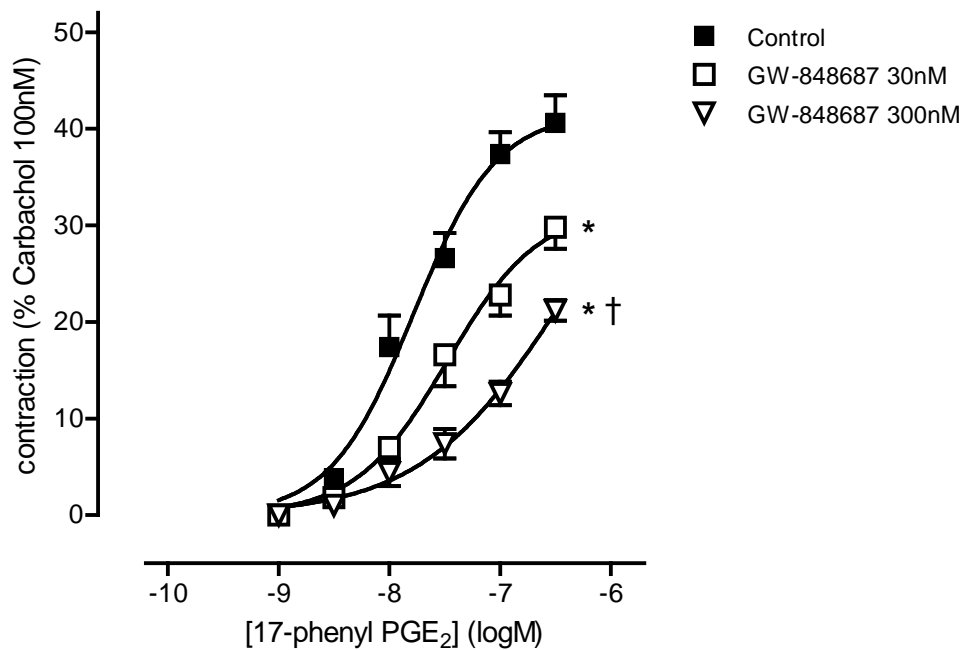
Pre-treatment with 300 nM GW-848687 significantly displaced the PGE<sub>2</sub> curve with depression of tone ( $p = 0.002$ ) (Figure 5.15,  $n = 4$ ). 30 nM GW-848687 did not significantly affect the control curve ( $p = 0.06$ ).

The effect of the GW-848687 on 17-phenyl PGE<sub>2</sub>-induced contractions is shown in Figure 5.16 ( $n = 4$ ). Pre-incubation with GW-848687 at two different concentrations (30 and 300 nM) before the cumulative addition of 17-phenyl PGE<sub>2</sub> significantly reduced the tone ( $p = 0.01$ , both). There were also significant displacement in non-parallel manner between control with 30 nM and 300 nM GW-848687 curve ( $p = 0.01$ ).



**Figure 5.15** Antagonism of PGE<sub>2</sub>-induced contraction by GW-848687 in rat urinary bladder. Log concentration-response curves for PGE<sub>2</sub> following treatment with 30 nM and 300 nM GW-848687 are shown ( $n = 4$ ). \* $P < 0.01$ , 300 nM GW-848687 vs. control and 30 nM GW-848687, using main effects contrasts.



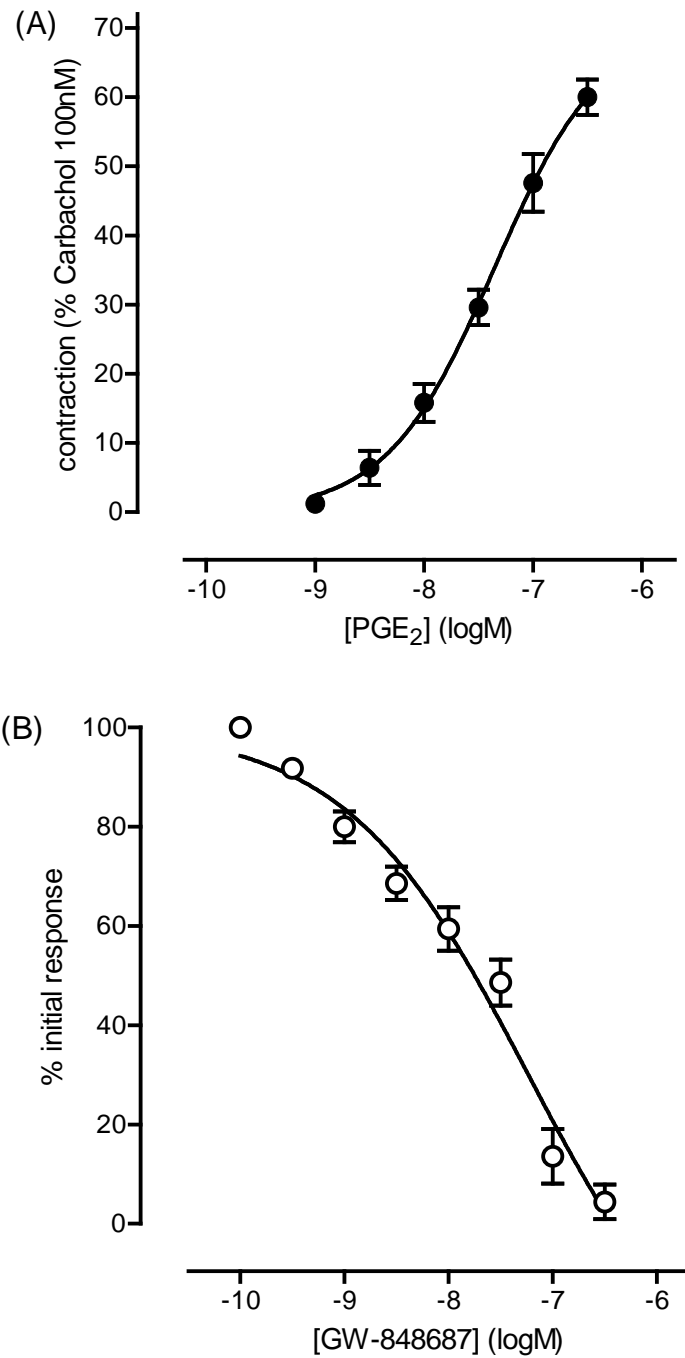


**Figure 5.16** Antagonism on 17-phenyl PGE<sub>2</sub>-induced contraction by GW-848687 in rat urinary bladder. Log concentration-response curves for 17-phenyl PGE<sub>2</sub> following treatment with 30 nM and 300 nM GW-848687 are shown ( $n = 4$ ).  $*P < 0.01$ , 30 nM and 300 nM GW-848687 vs control;  $^{\dagger}P < 0.01$ , 300 nM GW-848687 vs. 30 nM GW-848687, using main effects contrasts.

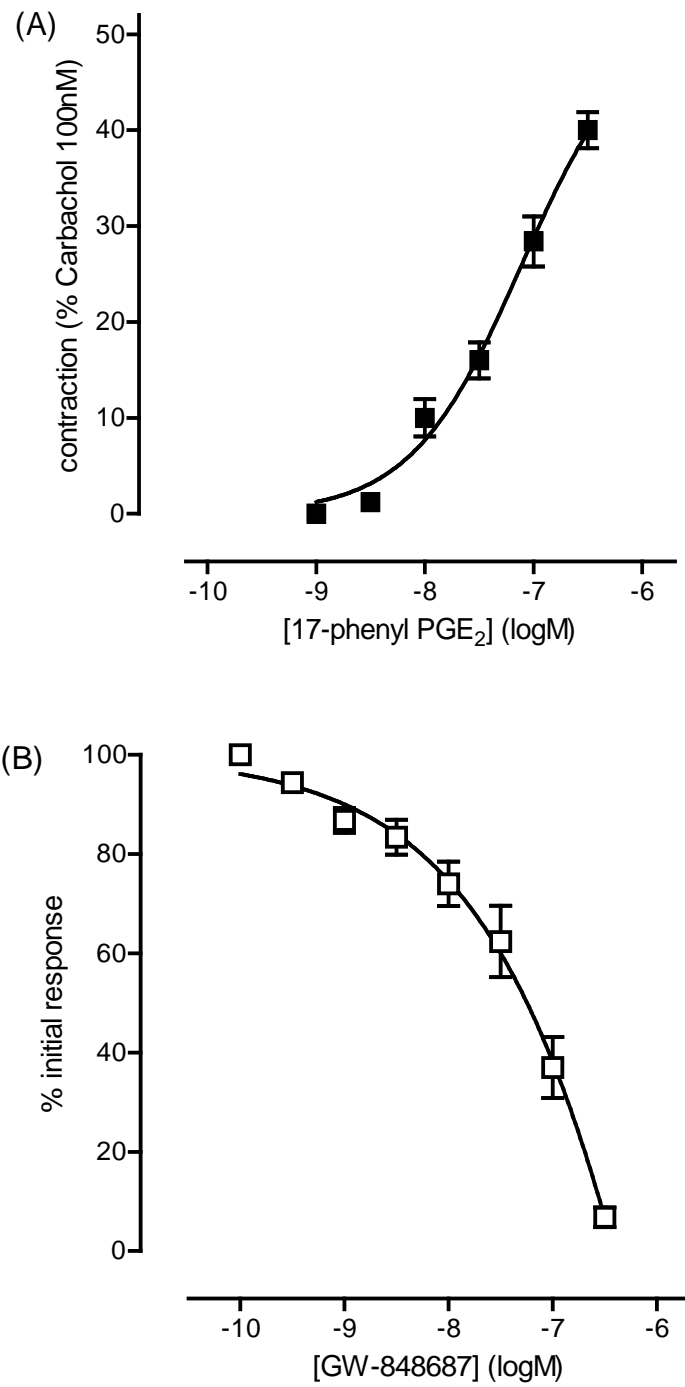
### **5.3.7 Antagonist potency of GW-848687 under inhibition-curve protocol**

The antagonist property of GW-848687 was assessed using the inhibition-curve protocol (protocol 5.2B). The response of PGE<sub>2</sub> in the absence of GW-848687 can be shown in Figure 5.17A ( $n = 5$ ). pEC<sub>50</sub>\* estimated was  $7.64 \pm 0.06$ . The effect of cumulative addition of GW-848687 (0.1 - 300 nM) on established tone to PGE<sub>2</sub> 100 nM is demonstrated in Figure 5.17B ( $n = 5$ ). The inhibition curve pIC<sub>50</sub>\* was estimated to be  $7.83 \pm 0.11$ . The pA<sub>2</sub> estimated from equation described in section 5.2.4 was 8.35.

17-Phenyl PGE<sub>2</sub> curve in the absence of antagonist was constructed as shown in Figure 5.18A ( $n = 5$ ). pEC<sub>50</sub>\* estimated from the graph was  $7.50 \pm 0.11$ . Inhibition curve of GW-848687 then constructed against the tone established with 100 nM 17-phenyl PGE<sub>2</sub> (Figure 5.18B,  $n = 5$ ). GW-848687 started to relax the tone at 0.3 nM, and continue to do so with a definitive plateau at each cumulative concentration increment. The pIC<sub>50</sub>\* estimated from the inhibition curve is  $7.39 \pm 0.15$ . Using the equation in Section 5.2.2, calculated pA<sub>2</sub> for GW-848687 against as 7.64.



**Figure 5.17** The inhibition-curve protocol for GW-848687 versus PGE<sub>2</sub> in rat urinary bladder. (A) Log concentration-response curve for PGE<sub>2</sub> ( $n = 5$ ). (B) Log concentration-inhibition curve for GW-848687, precontracted with 100 nM PGE<sub>2</sub> ( $n = 5$ ).



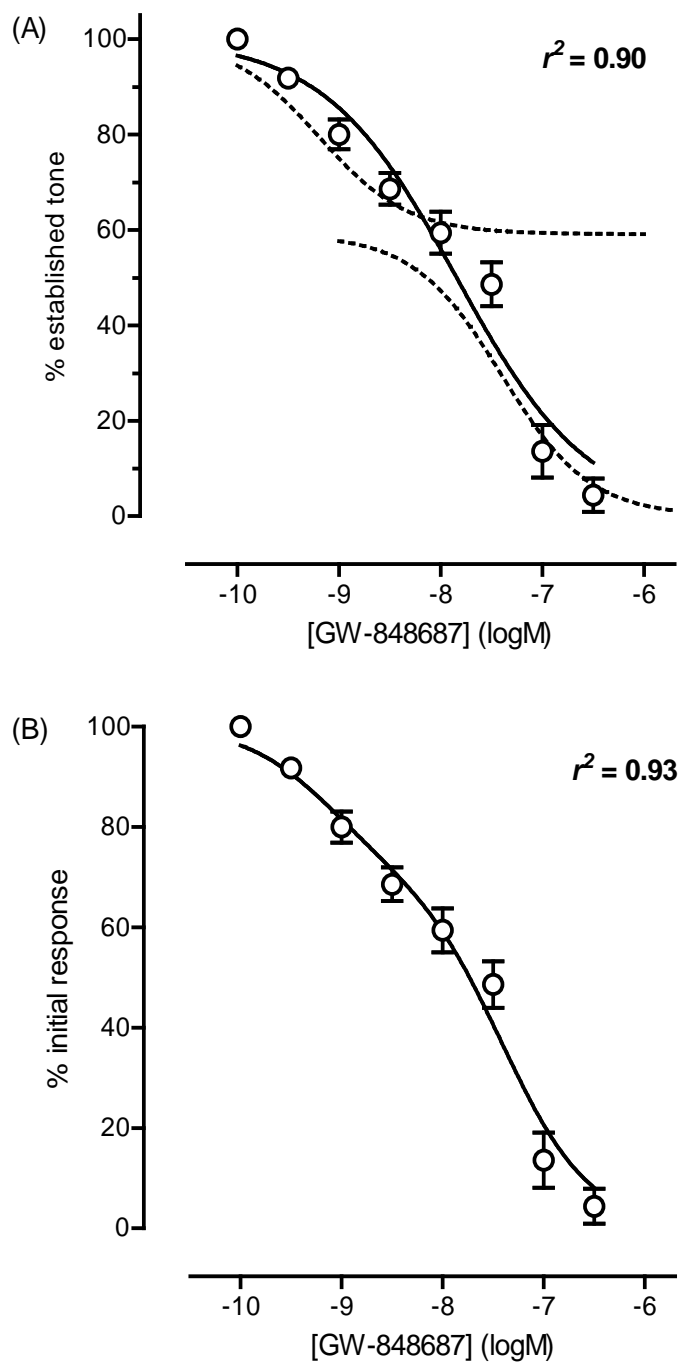
**Figure 5.18** The inhibition-curve protocol for GW-848687 versus 17-phenyl PGE<sub>2</sub> in rat urinary bladder. (A) Log concentration-response curve for 17-phenyl PGE<sub>2</sub> ( $n = 5$ ). (B) Log concentration inhibition curve for GW-848687, against 100 nM 17-phenyl PGE<sub>2</sub> ( $n = 5$ ).

### 5.3.8 Two-site competition model

It appeared that there could be two components to the inhibition-curve for GW-8488687 against both PGE<sub>2</sub> (Figure 5.17B) and 17-phenyl PGE<sub>2</sub> (Figure 5.18B). Two theoretical curves can be fitted in each graph (Figure 5.19B, Figure 5.20B). This was pursued using the two-site competition equation in GraphPad Prism. The Hill slopes of both sigmoidal components are automatically constrained to 1.0.

The best-fit two-site curve has a high affinity site with a pEC<sub>50</sub> of 9.15 comprising 28% of the sites, and a low affinity site with pEC<sub>50</sub> of 7.40 (Figure 5.19B). These results are certainly scientifically plausible. Confidence interval is also reasonably narrow (12 – 45 %). Because the results are sensible, it makes sense to compare the sum-of-squares statistically as described in Section 2.5.2.

There are forty data points in the inhibition curve in GW-848687 against 100 nM PGE<sub>2</sub>. There were four parameters fitted by the one-site equation fit and five parameters fitted for the two-site model. The calculation for the data can be summarized in Table 5.2. The F ratio is 10.08 with 1 (numerator) and 35 (denominator) degrees of freedom. Using the F table (appendix), the two-site model is significantly better than one-site model ( $p < 0.05$ ).



**Figure 5.19** The two-site competition model curve re-fitted on GW-848687 – PGE<sub>2</sub> inhibition curve (from data in Figure 5.17B) using GraphPad Prism software. (A) The two theoretical curves superimposed on one-site model fit. (B) The two-site model fitted on similar curve.

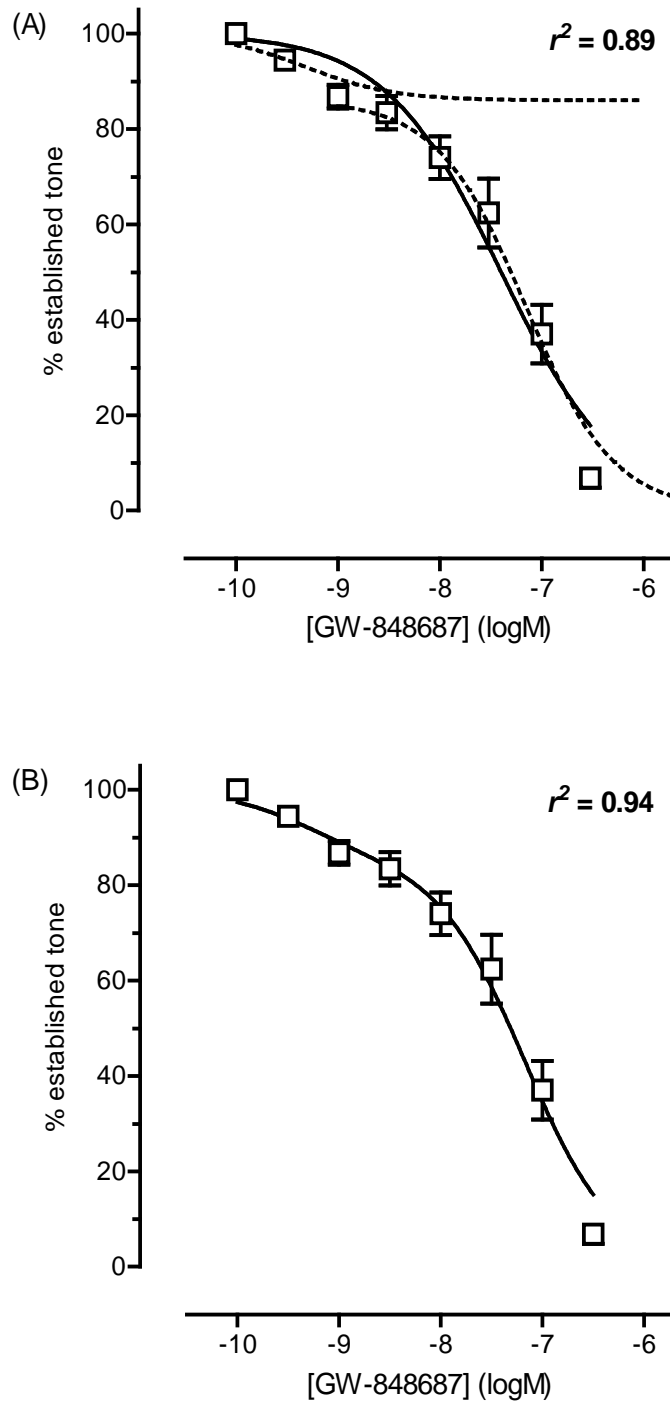
**Table 5.2** The F test for GW-848687 against 100 nM PGE<sub>2</sub>.

<b>Model</b>	<b>SS</b>	<b>df</b>
Null hypothesis (1 site)	3688	36
Null hypothesis (2 site)	2863	35
Difference	825	1
<b>Fractional difference</b>	0.2882	0.0286
<b>Ratio (F)</b>	<b>10.08</b>	

The same applied for GW-848687 inhibition curve against 100 nM 17-phenyl PGE<sub>2</sub> (Figure 5.18B). The data points were not fitted in the curve properly using the one-site model. Using the two-site model, two possible inhibition curves can be fitted in the original curve (Figure 5.20A). The best-fit two-site curve has a high affinity site with a pIC<sub>50</sub> of 9.32 comprising 14% of the sites, and a low affinity site with pIC<sub>50</sub> of 7.17 (Figure 5.20B). Even the fraction of high affinity sites is only 14% but 95% confidence interval are reasonably narrow (3 - 26%).

There are forty data points in the inhibition curve in GW-848687 against 100 nM 17-phenyl PGE<sub>2</sub>. Similarly, four parameters were fitted in one-site model and five parameters in two-site models. The F test calculation for GW-848687 against 17-phenyl PGE<sub>2</sub> is summarized in Table 5.3. The F ratio is 9.02 with 1 (numerator) and 35 (denominator) degrees of freedom. In reference to F table (appendix), the two-site model fitted is significantly better than one-site model ( $p < 0.01$ ).

The next step is to examine how well is the graph fits, the  $w$ -values. For GW-848687 against PGE<sub>2</sub>, the  $r^2$ -value is 0.93 for two-component fit and 0.90 for one-component fit. For GW-848687 against 17-phenyl PGE<sub>2</sub>,  $r^2$ -value is 0.94 for two-component fit and 0.89 for one-component fit.



**Figure 5.20** The two-site competition model curve re-fitted on GW-848687 – 17-phenyl PGE<sub>2</sub> inhibition curve (from data in Figure 5.18B) using GraphPad Prism software. (A) The two theoretical curves superimposed on one-site model fit. (B) The two-site model fitted on similar curve.



**Table 5.3** The F test for GW-848687 against 100 nM 17-phenyl PGE<sub>2</sub>.

<b>Model</b>	<b>SS</b>	<b>df</b>
Null hypothesis (1 site)	4052	36
Null hypothesis (2 site)	3221	35
Difference	831	1
<b>Fractional difference</b>	0.2580	0.0286
<b>Ratio (F)</b>	<b>9.02</b>	

## 5.4 Discussion

### 5.4.1 General overview

The rat urinary bladder is a useful tool to study functionally prostanoid antagonists due to its ready availability. The anatomical structure of the rat urinary bladder itself is similar to other hollow organs in the body with slight differences in the arrangement of the musculature layer. The muscle is not arranged in layer, instead as bundles (Gabella and Uvelius, 1990). The muscle cells are uninucleated. There is an abundance of collagen in between the muscle bundles, contributing to the high distensibility of the bladder.

The control of the rat urinary bladder involves the interplay of various factors (Figure 5.1). Prostanoids are involved in modulating and maintaining the basal tone of the bladder. Prostanoids exert their effects directly on bladder smooth muscle cells and by sensitising the afferent nerves (Schroder *et al.*, 2003). Indirectly, prostanoids are potent stimuli for neurokinins to be released, which induce the micturition reflex in the rat bladder (Ishizuka *et al.*, 1995). There has been no study documented on the direct interaction between prostanoids and autonomic nervous system on rat bladder function. However, one study has shown an interaction between PGE<sub>2</sub> and parasympathetic system at the level of spinal cord (Miura *et al.*, 2002). Parasympathetic preganglionic neuron in the rat spinal cord regulates the activity of pelvic visceral organs. The stimulation of EP<sub>1</sub> receptor in the spinal cord stimulates the parasympathetic neuron that leads to a hyperexcitability state in the bladder. Otherwise, no study has demonstrated direct interaction between prostanoids and autonomic nervous system in bladder control.

Prostanoids might be released during manipulation and removal of the bladder (Park *et al.*, 1999). In this study, it was noted that the preparation had not settled during the initial set-up of resting tension, which may be attributed to increase in endogenous prostaglandins. This

effects were prevented by the inclusion of 1  $\mu$ M indomethacin in the organ bath (Schroder *et al.*, 2003). The current study also included a selective TP antagonist (100 nM BMS-180291), which is usually included by many researchers during EP receptor studies and described in Section 1.7.1.

#### **5.4.2 Response to prostanoid agonists**

Little is known of the prostanoid receptor characterization in the rat urinary bladder. Earlier functional and qualitative studies mainly concentrated on a particular prostanoid receptor (Maggi *et al.*, 1988; Schroder *et al.*, 2004; Chuang *et al.*, 2008; Su *et al.*, 2008). Therefore, the overall aim of this study was to characterize the prostanoid receptor in the rat urinary bladder.

In the current study, initial results showed that the EP agonists had contractile effects that are rapid in onset. PGE<sub>2</sub> is a prostanoid agonist that can activate all four EP receptor subtypes with different affinity; in the rank order: EP<sub>3</sub> > EP<sub>4</sub> >> EP<sub>2</sub> > EP<sub>1</sub> with lower affinity for FP, DP, TP and negligible affinity for IP receptors (> 10,000) (Abramovitz *et al.*, 2000). PGE<sub>2</sub> has been demonstrated to increase bladder pressure in the normal, conscious rat (Ishizuka *et al.*, 1995). In agreement, in the current study, PGE<sub>2</sub> contracted the quiescence preparation. 17-Phenyl PGE<sub>2</sub> (EP<sub>1</sub> > EP<sub>3</sub>, with minimal activity on EP<sub>2</sub>) and sulprostone (EP<sub>3</sub> > EP<sub>1</sub>, with some activity on FP) also contracted the bladder strips with different potency and maximal response in the presence of indomethacin and the selective TP receptor antagonist (Figure 5.4) (Table 5.4).

**Table 5.4** The pEC<sub>50</sub> and E<sub>max</sub> for three prostanoid agonist used in the current experiment, at maximal concentration used (300 nM).

Agonists	pEC <sub>50</sub>	E <sub>max</sub> (300 nM) (%)
PGE <sub>2</sub>	7.41 ± 0.08	72 ± 3
17-Phenyl PGE <sub>2</sub>	7.29 ± 0.08	56 ± 8
Sulprostone	7.13 ± 0.13	37 ± 3

Latanoprost-FA, is an analogue of a prostaglandin F<sub>2</sub> alpha (PGF<sub>2α</sub>) and highly selective to FP receptor with K<sub>i</sub> value of 2.8 nM (Abramovitz *et al.*, 2000). In this study, latanoprost-FA (100 - 300 nM) failed to produce any contractile response. The lack of activity of latanoprost-FA suggested the absence of FP receptor in the rat urinary bladder.

The potent TP analogue, U-46619 (Coleman *et al.*, 1981; Abramovitz *et al.*, 2000) did not contract the muscle strips even with the higher dose up to 1 μM. 100 nM BMS-180291 had a weak inhibition (< 10%) effect on 100 nM carbachol, but this unlikely to be due to activation of TP receptor. The involvement of endogenous TXA<sub>2</sub> released by the mechanical stimulation can be ruled out as indomethacin was present in the bath fluid throughout the experiment. It is unlikely that the TP receptor is present in the rat urinary bladder.

Butaprost has a high affinity for the EP<sub>2</sub> receptor with negligible activity on the other EP receptors (Gardiner, 1986; Coleman *et al.*, 1994a; Narumiya *et al.*, 1999; Abramovitz *et al.*, 2000; Wilson *et al.*, 2004). It is assumed that this activity results from hydrolysis of its C1-ester group by non-specific esterases present in most tissues. In the present study, butaprost-free acid (FA) failed to relax carbachol precontracted bladder strips in spite of higher concentration (100-300 nM) used. The possible explanation is there was no EP<sub>2</sub> receptor in the rat urinary bladder. Moreover, butaprost-FA also has been shown to contract the guinea-

pig ileum at concentration as low as 30 nM (Lawrence *et al.*, 1992). The contraction has been attributed to EP<sub>1</sub> receptor in the guinea-pig ileum. It is possible that the absence of relaxation with butaprost-FA in the rat bladder could be due to the presence of highly sensitive EP<sub>1</sub> receptor system.

A study by Chuang *et al.* (2008) has demonstrated the over-expression of EP<sub>4</sub> receptor in parallel to an increase in COX-2 level in the bladder of the rat with cyclophosphamide-induced cystitis. However, the basal level of the EP<sub>4</sub> receptor in the normal rat urinary bladder was not documented. The exact mechanism of the EP<sub>4</sub> receptor causing the hypersensitivity in cystitis was not explained in the study. In general, the EP<sub>4</sub> is inhibitory to smooth muscles, i.e. to cause relaxation. A selective EP<sub>4</sub> antagonist was not available at the time of the current experiment to assess the EP<sub>4</sub> activity in the rat urinary bladder. PGE<sub>2</sub> however, is known to have high affinity for EP<sub>4</sub> receptor with  $K_i$  value of 0.79 nM (Abramovitz *et al.*, 2000). In the current experiment, PGE<sub>2</sub> (100 - 300 nM) did not relax the preparation precontracted with 80 nM carbachol. Thus, it is unlikely that the EP<sub>4</sub> receptor involved directly in modulating bladder function in the current experiment. In view of the above results, the next step was to characterize the contractile EP receptor in the rat urinary bladder using the antagonist studies.

### **5.4.3 The nature of the EP receptor(s) mediating contraction of the rat urinary bladder preparation**

The high agonist potency of 17-phenyl PGE<sub>2</sub> and sulprostone on the rat urinary bladder preparation indicated the potential presence of both EP<sub>1</sub> and EP<sub>3</sub> receptors. Both Schild plot and inhibition curve protocols with EP<sub>1</sub> and EP<sub>3</sub> antagonists were used to clarify this situation. Under the Schild protocol, the antagonism profile of SC-51322 and GW-848687 did not conform to classical competitive antagonism at a single receptor site. Thus, the agonist log concentration-response curves (CRCs) under antagonist treatment were not parallel to the vehicle curve and there was some evidence of non-surmountability. In guinea-pig trachea preparations, SC-51322 (Hung *et al.*, 2006) and GW-848687 (Jones RL, personal

communication) produced a parallel right-shifts of the 17-phenyl PGE<sub>2</sub> curve; Schild plot slopes were not significantly different from unity, thereby allowing pA<sub>2</sub> values to be calculated (8.45 and 9.75; respectively). Two explanations appear possible in the first instance: either both EP<sub>1</sub> antagonists behave non-competitively (see later) or two receptors contribute to the contractile activity of the prostaglandin E analogues examined.

In examining the second proposal, the right-shifts of the agonist logCRC caused by SC-51322 and GW-848687 are consistent with their known affinities for the EP<sub>1</sub> receptor. From published data (Hung *et al.*, 2006; Giblin *et al.*, 2007), one would expect selective block of EP<sub>1</sub> receptors at 30 nM of either SC-51322 or GW-848687. The same could be true of 300 nM, but it is a common occurrence for the high selectivity of a novel antagonists reported initially to be subsequently downgraded. Similarly, the right-shifts of the logCRCs for PGE<sub>2</sub>, 17-phenyl PGE<sub>2</sub> and sulprostone by 1 μM L-798106 indicate the existence of a functional EP<sub>3</sub> system (Figure 5.8, 5.9 and 5.10).

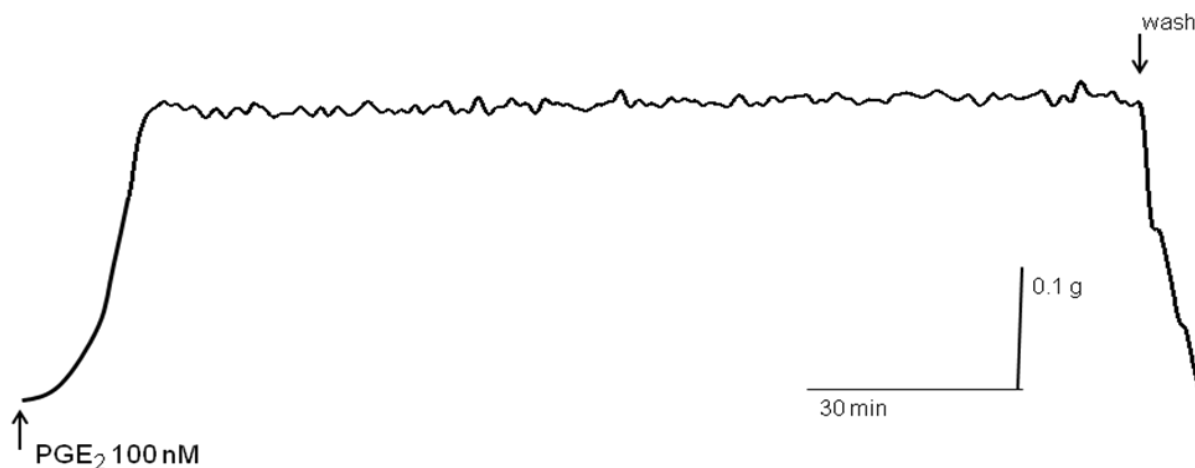
However, the combination of EP<sub>1</sub> and EP<sub>3</sub> antagonists under the Schild protocol does not support the above findings. Figure 5.13 clearly shows that the addition of L-798106 to SC-51322 pre-treatment did not produce further right-shifts of the sulprostone curve, thereby providing little support for the presence of an EP<sub>3</sub> contractile system.

Turning to inhibition curve protocols, there is statistically sound evidence using SC-51322 and GW-848687 for a two-component contractile system in the rat bladder preparation. Again, the higher sensitivity system is compatible with selective EP<sub>1</sub> antagonism, with GW-848687 at 1 nM showing clear inhibition of PGE<sub>2</sub>-induced contraction (Figure 5.19). The lower sensitivity component may represent the EP<sub>3</sub> receptor, but it was expected that GW-848687 would show higher selectivity than the current experiments inform.

There is some discrepancy between the inhibitory potencies of the EP<sub>1</sub> blockers under the Schild and inhibition-curve protocols, with greater potency accorded to the latter. It is unlikely that this is due to fade of the agonist response under the inhibition curve protocol. As shown in Figure 5.21, tension induced by the EP agonist was well maintained for at least 90 min.

The inhibition curve protocol using the L-798106 provided strong evidence for an EP<sub>3</sub> system, with inhibition of sulprostone-induced contraction starting at 1 nM and proceeding monotonically to 87% inhibition at 300 nM (Figure 5.14). However, this large degree of inhibition was unexpected and also clearly disagrees with data in Figure 5.11.

Several issues must be considered in relation to the above discrepancies. In relation to the kinetics properties of the antagonists, had the antagonists reached steady-state at the time of agonist addition? Thirty minutes pre-treatment was allowed for L-798106 in the Schild protocol, while 10 - 15 min was allowed for each concentration increment in the inhibition-curve protocol. It seems unlikely that this is relevant issue given the lower L-798106 concentrations effective in the inhibition-curve protocols. In other words, we would expect to see slower inhibition at the lower concentrations thereby potentially underestimating the potency of the antagonist – this was not the case. The constancy of the contribution of the two contractile components between bladder preparations from different animals needs to be considered also. It is difficult to assess this factor, given that sulprostone and 17-phenyl PGE<sub>2</sub> have both EP<sub>1</sub> and EP<sub>3</sub> agonist activity. Finally, how do the two contractile systems interact when simultaneously activated? In the rat femoral artery, there is synergism between EP<sub>3</sub> agonist and phenylephrine ( $\alpha_1$  agonist), K<sup>+</sup> (depolarizing agent) or U-46619 (TP agonist) (Hung *et al.*, 2006). In guinea-pig aorta, where there is synergism between EP<sub>3</sub> receptor and  $\alpha_1$ -adrenoreceptor systems, comprehensive EP<sub>3</sub> antagonism does not completely return established contraction to the level of  $\alpha_1$  agonist alone (Jones RL, personal communication). Could there be synergism between the EP<sub>1</sub> and EP<sub>3</sub> systems in the rat urinary bladder preparation?



**Figure 5.21** Typical tracing of 100 nM PGE<sub>2</sub>-induced contraction response in the rat bladder in the absence of antagonists, time-control preparation. The established response was well sustained over the period of more than 90 min.

#### 5.4.4 Two-site competition model versus one-site competition model

The possibility of two contractile receptor systems in the rat urinary bladder can be further examine by refitting the antagonist-inhibition curve as described in details in Section 2.5.2. The non-parallel right displacement by GW-848687 on PGE<sub>2</sub> curves suggested the presence of two receptor systems in the rat bladder, possibly EP<sub>1</sub> and EP<sub>3</sub> receptors. The graph of the one site fit model for GW-848687 against 100 nM PGE<sub>2</sub> seems to deviate systematically from the data point (Figure 5.17B). The possibility of two contractile receptor systems is demonstrated by re-fitting the curve on Figure 5.17B using the two-site model in GraphPad Prism program (Figure 5.19B). The data seems better fitted in two-site model.

Both, the F test and  $r^2$ -values comparison results suggested the incomplete inhibition by GW-848687 on tone established by PGE<sub>2</sub> and 17-phenyl PGE<sub>2</sub> could be due to presence of two functional EP-receptors system in the rat urinary bladder.

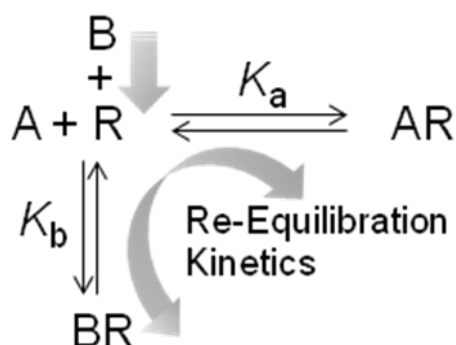


### 5.4.5 Insurmountable antagonism and the hemi-equilibrium concept

The depression of maximal agonist response with a relatively small right-shift of the agonist curve is usually termed 'insurmountable antagonism' (Kenakin *et al.*, 2006). Insurmountable antagonism can be due to non-competitive antagonism, whereby the antagonist either binds irreversibly at or close to the agonist binding site or affects receptor function at a site distinct from the agonist binding site. SC-51322 and GW-848687 have been documented to behave as competitive antagonists in functional and / or recombinant receptor studies (Hung *et al.*, 2006; Giblin *et al.*, 2007), but this does not necessarily mean that they behave similarly on the EP<sub>1</sub> receptor system in the rat urinary bladder.

Another possibility involves true competition for the agonist binding site, but a hemi-equilibrium state exists for the antagonist-receptor interaction. In an ideal situation, on addition of agonist to determine the extent of the antagonism, the receptor should fully re-equilibrate with the antagonist and agonist in the receptor compartment. For a low- to moderate-affinity antagonist, the antagonist dissociation rate constant will be relatively large and antagonist will quickly dissociate, thereby allowing agonist to occupy the empty receptors. For a high-affinity antagonist, dissociation will be much slower and the agonist response may be measured at a time when the dissociation process is incomplete. An insurmountable antagonist profile may thus occur (Figure 5.22, redrawn from Kenakin *et al.*, 2006). To overcome this hemi-equilibrium situation, the antagonist contact time must be increased, i.e. increase the duration before addition of the consecutive agonist dose.

### Equilibration Kinetics



**Figure 5.22** The re-equilibration kinetics that contributes to the hemi-equilibration state; A = agonist, B = antagonist, R = receptor (redrawn from Kenakin *et al.*, 2006).

#### 5.4.6 EP<sub>4</sub> receptor

The functional EP<sub>4</sub> receptor in the rat bladder cannot be determined in the current study due to non-availability of selective EP<sub>4</sub> agonist or antagonist. However, the failure of PGE<sub>2</sub> to inhibit the precontracted preparation suggested the absence of EP<sub>4</sub> in the rat urinary bladder. Despite over-expression of EP<sub>4</sub> receptor in rat bladder following cystitis (Chuang *et al.*, 2008) this study failed to show the contribution from EP<sub>4</sub> receptor in bladder function. Nevertheless, the functionality of EP<sub>4</sub> in bladder can be explored once the appropriate receptor agonist or antagonist made available.

## 5.5 Conclusions

In the present work, it has been shown that the prostanoid agonists produced a contractile effect on the rat urinary bladder. It seems likely that a EP<sub>1</sub> receptor is present. However, there was some doubt as to the identity of the second receptor as an EP<sub>3</sub> receptor. This arose from discrepancies between the degree of antagonism seen a) with combined use of EP<sub>1</sub> and EP<sub>3</sub> antagonists and EP<sub>1</sub> or EP<sub>3</sub> antagonist alone under the Schild protocol, and b) under the Schild and inhibition curve protocols. There was no functional relaxant prostanoid receptor documented in this study.

GW-848687 did not show surmountable antagonism on the bladder preparation. This may be explained on the basis of two contractile EP receptors as alluded to above. Another possibility to consider is that GW-848687 is selective towards the EP<sub>1</sub> receptor, but has a non-competitive mechanism or achieves a hemi-equilibrium state. This situation needs to be resolved before one could fully recommend GW-848687 as a robust tool for characterizing EP<sub>1</sub> receptor systems.

**CHAPTER SIX**

**RAT FEMORAL ARTERY**

**AND**

**DIABETES**

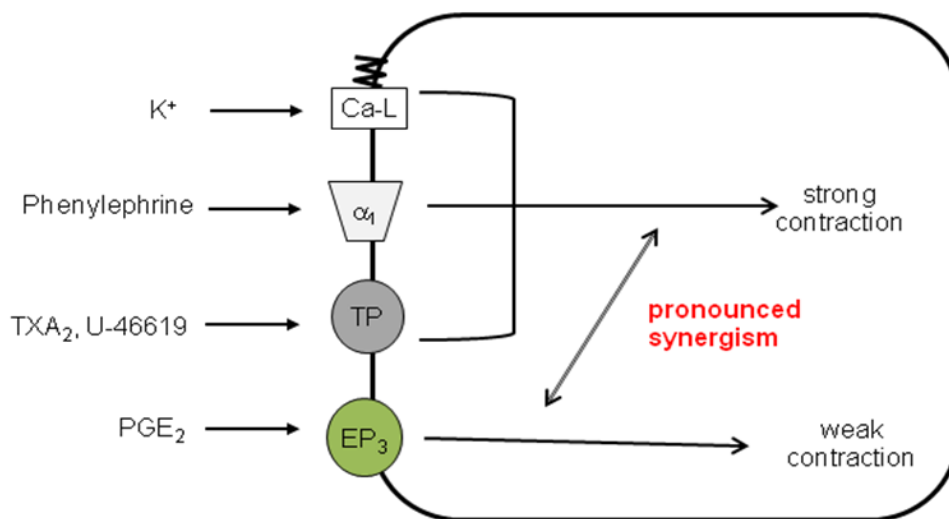
## 6.1 Introduction

Endothelium dysfunction has been reported in diabetic people and experimental animals with diabetes (De Vriese *et al.*, 2000, Schofield *et al.*, 2002). The dysfunction is caused by the high glucose content in tissues, which triggers a cascade of functional and structural alterations in vascular cells, leading to macrovascular and microvascular diseases (Schalkwijk and Stehouwer, 2005). A hallmark of endothelial dysfunction in diabetes is alteration in the biosynthesis of prostaglandins and oxidative stress products and decreased release and / or bioavailability of nitric oxide (NO) (Tesfamariam *et al.*, 1989; Vanhoutte *et al.*, 2005). The hyperglycaemic state in diabetes alone is the stimulation for productions of prostaglandins (Tesfamariam *et al.*, 1990). Endothelial COX, which is upregulated in various pathological conditions including diabetes, is thought to be responsible (Tesfamariam *et al.*, 1989; Ge *et al.*, 1995; Shi *et al.*, 2006; Shi *et al.*, 2007b). In contrast, Peredo *et al.* (2001) showed impairment of prostanoid production in the mesenteric vascular bed of the diabetic rat.

Endogenous prostanoids are likely to exert their constrictor or dilator actions on blood vessels by activating prostanoid receptors located on smooth muscle cells (Narumiya *et al.*, 1999; Breyer *et al.*, 2001). With reference to the diabetic state, the increased levels of vasoconstrictor prostaglandins act mainly through TP receptors in femoral artery from rats with early stage STZ-induced diabetes (Peredo *et al.*, 1999; Shi *et al.*, 2007a) and in aorta from alloxan-diabetic rabbits (Tesfamariam *et al.*, 1989). In contrast, the synthesis of the vasodilator prostacyclin is decreased in STZ-induced diabetic rats (Harrison *et al.*, 1978; Peredo *et al.*, 1999). Moreover, Shi *et al.* (2007a) showed that the TP antagonist, S-18886 partially inhibited the contractile response to A-23187 (calcium ionophore) in 12 weeks STZ-rat femoral artery. S-18886 in combination with SC-19220 (EP<sub>1</sub> antagonist) and AH-6809 (DP<sub>1</sub>, EP<sub>1</sub>, EP<sub>2</sub> antagonist) abolished the contraction. In support of this picture, there was a mixed production of vasoconstrictor prostaglandins and TXA<sub>2</sub>, in rat mesenteric arterial bed (Peredo *et al.*, 1999).

While the vascular action of TXA<sub>2</sub> is straightforward, that of PGE<sub>2</sub> is potentially complicated. Firstly, PGE<sub>2</sub> may cause vasodilatation through activation of EP<sub>2</sub> and / or EP<sub>4</sub> receptors. Secondly, it may activate TP receptors at high concentrations, as shown by Jones *et al.* (1982) in rabbit aorta and by Dorn *et al.* (1992) in rat aorta. Thirdly, PGE<sub>2</sub> may activate an EP receptor, most likely an EP<sub>3</sub> receptor, to cause vessel contraction. Thus, in rat femoral artery, PGE<sub>2</sub> acting alone induced very weak contraction, but showed pronounced synergism with strong contractile agents including K<sup>+</sup>, phenylephrine and the TP agonist U-46619 (Figure 6.1) (Hung *et al.*, 2006). One presumes that it would show a similar synergistic interaction with the natural TP agonist TXA<sub>2</sub>. However, this EP<sub>3</sub> receptor-driven synergism has not been taken into account when PGE<sub>2</sub> has been implicated in the diabetic state. For example the TP receptor antagonist S-18886 only partially blocked the contractile response to the calcium ionophore, A-23187 in femoral artery from the 12-week STZ diabetic rat (Shi *et al.*, 2007a). The possibility of activation of both TP and EP receptors in the chronic diabetic state was proposed by the authors, but no reference was made to the potential interaction of these two prostanoid systems.

The effect of diabetes in rat can be examined in various models. The Zucker fatty (ZF) rat (Okamoto *et al.*, 2008) and Goto-Kakizaki (GK) rat (Goto *et al.*, 1976) are both developed by cross-breeding and manifest type 2 diabetes. A more convenient model is the streptozotocin (STZ) induced-diabetic rat. STZ is an *N*-nitroso derivative of D-glucosamine that is chemically unstable in saline or distilled water at room temperature and neutral pH. It has been utilised to induce diabetes in a variety of experimental animals and to determine the short and long-term complication of diabetes (Table 6.1). The dose of streptozotocin varies between 25 – 200 mgkg<sup>-1</sup> in different species. It was further suggested that the intensity of the damage to the β-cells could be graded according to the dosage used (Junod *et al.*, 1969).



**Figure 6.1** Prostanoid pharmacology in rat femoral artery. The synergism between phenylephrine and PGE<sub>2</sub> produced a strong contraction response. *Ca-L*: L-type calcium channel (Hung *et al.*, 2006).

It was shown that the diabetogenic action of streptozotocin results primarily from its highly specific cytotoxic action on insulin producing cells ( $\beta$ -cells of the islets of Langerhans) leading to rapid and irreversible necrosis (Srivastava *et al.*, 1982). The induction of diabetes by streptozotocin causes a triphasic blood-sugar response. Initially, there will be increases in blood sugar level secondary to glycogenolysis or an increase in free fatty acid (FFA). The glucose cannot be utilised properly due to reduction in circulating insulin, thus the body used the glycogen and fat store for the energy, leading to weight loss eventually. In the second phase, there is a profound hypoglycaemic state resulting from augmented endogenous release of insulin from the degenerating  $\beta$ -cells. In the final stage within 24 hours post-injection, an irreversible diabetic state developed (Szkudelski, 2001). The animals will be totally devoid of insulin, resulting in a hyperglycaemic state.

The initial aim of the experiments was to determine which prostanoid receptors are activated in the isolated femoral artery of the STZ-diabetic rat when enhanced endogenous prostaglandin production occurs, and how these prostanoid receptors interact with each other

and with other vaso-active agents. Shi *et al.* (2008) used exogenous hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) to generate an oxidative stress environment in rat femoral artery, based on the finding that  $\text{H}_2\text{O}_2$  produces contraction response in this artery and rat mesenteric artery (Matoba *et al.*, 2000). However, my initial experiments showed minimal or non-sustained contraction with  $\text{H}_2\text{O}_2$  despite the higher concentration used (more than 10 mM). The focus of the study was therefore switched to examine the role of prostanoids in the associated enhancement of contractile response to  $\alpha_1$ -adrenoceptor agonism or  $\text{K}^+$ -induced depolarization in the diabetic state. The evidence for the involvement of prostanoids in this enhancement derives from the partial inhibition of the contractile responses by the COX inhibitor indomethacin (Shi and Vanhoutte, 2008). Two critical decisions were necessary – which diabetic state to induce and how to differentiate the roles of TP and  $\text{EP}_3$  receptors.

In the literature, blood vessels have been harvested at varying intervals (1 – 52 weeks) after STZ injection; these studies are summarized in Table 6.1. Peredo *et al.* (1999) have shown the response to noradrenaline in endothelium intact STZ-rat mesenteric bed is similar in all duration of diabetes (2 to 8 weeks), and comparable to control. However, the maximal response was reduced in STZ-rats beyond 8 weeks. The maximal response in denuded STZ-rat femoral artery to U-46619 and 60 mM KCl were also shown to be similar to control at 4 weeks after STZ-induction (Shi *et al.*, 2007a). In contrast, there was significant reduction in response to U-46619 in 12 weeks STZ-rats. Based on these results, it was decided to use the denuded rat femoral artery after 4 weeks of STZ-induction as this is the first experience with STZ-rats model.

Phenylephrine, a selective  $\alpha_1$ -adrenoreceptor agonist, has been shown to contract femoral artery in both normal and diabetes rat (Jarajapu *et al.*, 2001; Shi *et al.*, 2006). Its action is mediated via  $\alpha_1$ -adrenoceptors, while the post-junctional action of  $\alpha_2$  agonists on rat femoral artery is negligible (Jarajapu *et al.*, 2001). Initial experiments confirmed good reproducibility for contraction of rat femoral artery by phenylephrine, which is essential if the synergism with  $\text{EP}_3$  agonist is to be accurately assessed.



**Table 6.1** Summary of published studies on effects of vasoactive agents on isolated blood vessels from control and streptozotocin-treated rats

Author, year	STZ dose and route	Induction duration (weeks)	Tissues	Endothelium status	Agents	Responses	Effects of antagonists / inhibitors
<b>Chang and Stevens, 1992</b>	55mg/kg IV	12 52	Rat aorta	Intact and removed	Phenylephrine	- increased sensitivity in 52 weeks STZ (increased EC <sub>50</sub> ; E <sub>max</sub> same) in intact endothelium compared to control - no difference in without endothelium	- not inhibited by indomethacin 5 µM - implicated as diabetes progressed, reduced EDRF due to endothelial dysfunction
<b>Dai et al., 1993</b>	55mg/kg IV	6 16 24	Renal artery	Intact and removed	Phenylephrine  Acetylcholine  Histamine	- no change in contractile response in both: intact vs. without endothelium control vs. STZ  - as diabetes progressed, reduction in ACh relaxation (intact endothelium)  - same as Ach	- as diabetes progressed, increase in endothelial and smooth muscle dysfunction due to increase in free radicals
<b>Mulhern and Docherty, 1989</b>	65mg/kg IP	2-3	Rat aorta	Intact and removed	Noradrenaline  KCl 40mM	- no difference between DM & controls - no difference between with & without endothelium  - as for noradrenaline	
<b>Peredo et al., 1999</b>	60mg/kg IP	1 2 3 4	Rat mesenteric bed	Intact	Noradrenaline	- contractile response, no difference of maximal response between control / STZ and duration of induction	- all contraction not affected by 10 µM indomethacin, except in 8 weeks animals (STZ / control)

Author, year	STZ dose and route	Induction duration (weeks)	Tissues	Endothelium status	Agents	Responses	Effects of antagonists / inhibitors
		8			KCl 80mM	- no difference between control / STZ and duration	- not affected by indomethacin
<b>Peredo, 2001</b>	100mg/kg IP	18-20	Rat mesenteric bed	Intact and <b>removed</b>	Noradrenaline	contractile response - no difference in control & STZ (same in with / without endothelium)	- indomethacin reduced the responses in control but not in STZ (with / without endothelium)
					KCl	- contractile response (as for noradrenaline)	- not affected by indomethacin (with / without endothelium)
<b>Pfaffman et al., 1982</b>	65mg/kg IP	2 – 12	Rat aorta	Intact	Noradrenaline	- significantly decreased E <sub>max</sub> in STZ but comparable EC <sub>50</sub>	- the response restored as in control with insulin administration
					KCl	- significantly decreased in STZ (EC <sub>50</sub> & E <sub>max</sub> )	
<b>Shi and Vanhoutte, 2008</b>	55mg/kg IV	12	Rat femoral artery	Intact	U-46619	- significantly reduced contractile response in STZ - curve shifted to the LEFT (hyper-responsiveness)	- 5 μM indomethacin / 100 nM naproxen– restored the response in STZ compared to control rats
					Phenylephrine	- contractile response reduced in STZ	
					KCl	- comparable between STZ & control	
<b>Shi et al., 2007a</b>	55mg/kg IV	4 12	Rat femoral artery	Intact & <b>removed</b>	Phenylephrine	- contraction response, no difference E <sub>max</sub> between STZ / control - no difference in duration - no differences in with / without endothelium	

Author, year	STZ dose and route	Induction duration (weeks)	Tissues	Endothelium status	Agents	Responses	Effects of antagonists / inhibitors
					A23187	<p><i>precontracted by phenylephrine</i>  <i>4 weeks</i>            - relaxation on lower dose, then contraction on higher dose            - <b>without endothelium</b>, no relaxation phase on STZ</p> <p><i>12 weeks</i>            - in control, response same as 4 weeks            - in STZ: no relaxation phase in with / without endothelium</p>	<p><i>4 weeks</i>            - the contraction by Phe inhibited by 5 <math>\mu</math>M indomethacin and 0.1 <math>\mu</math>M S-18886</p> <p><i>12 weeks</i>            - contraction in STZ inhibited by 5 <math>\mu</math>M indomethacin and 0.1 <math>\mu</math>M S-18886</p>
					A23187	<p><i>quiescent preparation</i>  <i>4 weeks</i>            - <b>without endothelium</b>: no response in STZ / control            - intact endothelium:            -contraction, STZ &gt;&gt; control</p> <p><i>12 weeks</i>            - without endothelium: as 4 weeks            - intact endothelium:            - significant contraction response                STZ &gt;&gt;&gt; control                STZ &gt;&gt;&gt; 4 weeks</p>	<p><i>4 weeks</i>            - contraction response in intact endothelium in both inhibited by 5 <math>\mu</math>M indomethacin and 0.1 <math>\mu</math>M S-18886</p> <p><i>12 weeks</i>            - contraction in intact endothelium partially inhibited by 5 <math>\mu</math>M indomethacin and 0.1 <math>\mu</math>M S-18886            - AH-6809 / SC-19220 / S-18886 completely inhibited the contraction response</p>
					U-46619	<p>- hyper-responsive – shift curve to <b>LEFT</b>, and reduced <math>E_{max}</math> in <b>without endothelium</b> of 12 weeks STZ (not in 4 weeks, comparable to control)</p>	

Author, year	STZ dose and route	Induction duration (weeks)	Tissues	Endothelium status	Agents	Responses	Effects of antagonists / inhibitors
					KCl 60 mM	- <b>without endothelium</b> : significant reduction of contraction in 12 weeks STZ	
<b>Shi et al., 2007b</b>	55mg/kg IV	12	Rat femoral artery	Intact and <b>removed</b>	A23187	+ L-NAME - intact endothelium: contraction with higher response in STZ - without endothelium: none	- contraction in intact endothelium inhibited by 5 µM indomethacin and 0.1 µM S-18886
					H <sub>2</sub> O <sub>2</sub>	- <b>without endothelium</b> : contractions; STZ >> control with LEFT shift	-contraction inhibited by 5 µM indomethacin and 0.1 µM S-18886
<b>Taylor et al., 1992</b>	56mg/kg IP	5-6	Rat mesenteric bed	Intact	Noradrenaline	- contraction response, increased in STZ compared to control	- L-NAME increased the response of control nearly as in STZ rats

\* *IP* = intraperitoneal; *IV* = intravenous; *KCl* = potassium chloride; *ACh* = acetylcholine; *Phe* = phenylephrine; *S-18886* (*TP* antagonist)

BMS-180291 was chosen as the TP antagonist. Its profile is well-documented and it blocked the U-46619 activity in rat aortic rings at pA<sub>2</sub> of 9.3 (Ogletree *et al.*, 1993) (Table 6.2). L-798106 was chosen as the EP<sub>3</sub> antagonist. It shows high selectivity for EP<sub>3</sub> receptors based on ligand binding at recombinant prostanoid receptors (Juteau *et al.*, 2001). In functional studies, it blocked the action of sulprostone with pA<sub>2</sub> values of 7.82 in guinea-pig trachea (Clarke *et al.*, 2004) and 7.43 – 8.03 in rat femoral artery (Hung *et al.*, 2006). In the most recent functional study in rat urinary bladder, L-798106 inhibit the PGE<sub>2</sub>-induced Ca<sup>2+</sup> influx through activation of EP<sub>3</sub> receptor with pK<sub>i</sub> of 7.12 (Jugus *et al.*, 2009).

The final aim of the experiment was to examine the effect of endothelium on the response of phenylephrine on STZ-rat femoral artery. Prostaglandins are synthesised by both endothelial and smooth muscle COX (Weksler *et al.*, 1978; Narumiya *et al.*, 1999). This study will establish whether the augmented prostanoids production in diabetes is endothelium dependent or not.

**Table 6.2** Antagonists and concentrations chosen for use in the experiment.

Antagonists	Reported pA <sub>2</sub>	Concentration used in the current experiment	References
L-798106	7.43 – 8.03	1 µM	Hung <i>et al.</i> , 2006; Clarke <i>et al.</i> , 2004
BMS-180291	9.3	100 nM	Ogletree <i>et al.</i> , 1993
CAY-10441	8.82	1 µM	Clark <i>et al.</i> , 2004

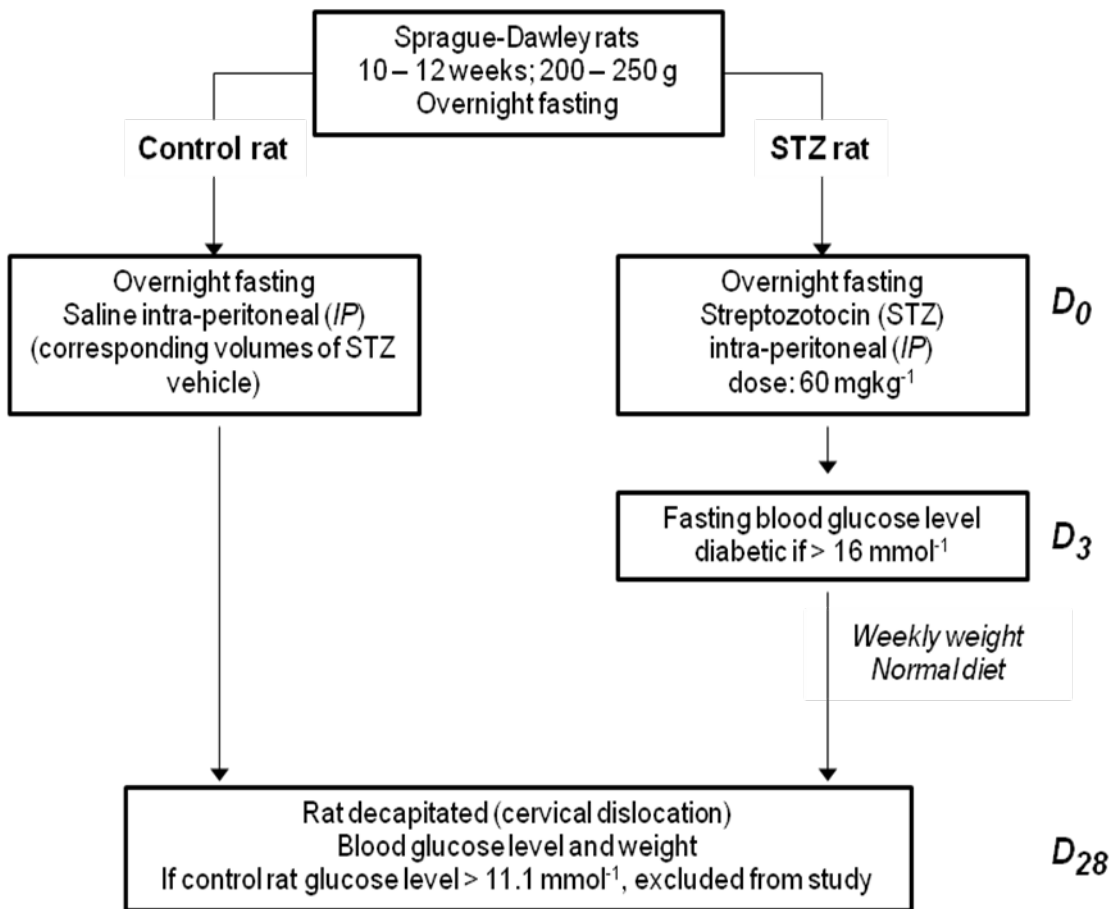
## 6.2 Methods

The basic methodology used has been described in detail in Chapter 2. Specific methodological points not discussed previously are addressed below.

### 6.2.1 Streptozotocin-induced diabetes

The experiment was carried out on 10 - 12 week old male Sprague-Dawley rats (200 - 250 g). Diabetes was induced with streptozotocin (STZ; 60 mgkg<sup>-1</sup>, i.p) after overnight fasting (Figure 6.2) (Pfaffman *et al.*, 1982; Peredo *et al.*, 1999; Burke *et al.*, 2006). Streptozotocin (20 mgml<sup>-1</sup>) was freshly prepared on the day of induction in 40 mM Na-Citrate buffer solution at pH 4.5 and used within 10 - 15 min. Control rats were injected with corresponding volumes of the STZ vehicle (saline). At 72 hours after STZ injection, a blood sample was taken from the tail artery and the fasting blood glucose level was measured using an Ascensia Breeze 2 glucometer (Bayer HealthCare, UK). Rats were considered diabetic if the blood glucose level was higher than 16 mmol<sup>-1</sup>. Body weight was measured weekly and rats were kept under optimal conditions and given a normal diet.

At 4 weeks after induction, the rats were decapitated. Blood glucose and final weight were measured on the day of death. Control rats with non-fasting blood glucose higher than 11.1 mmol<sup>-1</sup> were excluded from the study. All procedures and protocols were performed under Home Office Project Licence guidelines (project licence holder: Professor Brian Furman, University of Strathclyde).



**Figure 6.2** The protocol for streptozotocin (STZ)-induced diabetes in rat.

## 6.2.2 Setting up of preparations

The rat femoral arteries were obtained and mounted in a wire myograph as described in Chapter 2. When indicated, the endothelium was removed mechanically from the artery by rolling the intimal surface against a piece of roughened wire slightly thinner than the lumen of the artery. The resting tone was set at an optimal tension of 1.0 g, determined from preliminary experiments and previous study on the same settings (Shi *et al.*, 2007a; Shi *et al.*, 2007b). Four preparations were used contemporaneously. The preparations were allowed to equilibrate for 1 h, and then exposed twice to KCl at 60 mM for viability assessment. The criteria of exclusion as described in Section 2.2.1.2 were applied.

The artery was contracted with 100 nM phenylephrine (60% maximal response,  $EC_{60}$ ), followed by cumulative addition of acetylcholine (1, 10, 100 nM) to assess the completeness of endothelium removal (Furchgott and Zawadzki, 1980; Furchgott, 1981). Where applicable, the antagonists used were preincubated for 30 min before cumulative addition of phenylephrine. Repeated concentration response curve to phenylephrine were reproducible and no corrections for time-dependent changes were required.



### 6.2.3 Antagonist protocol and pA<sub>2</sub> estimation

Antagonist affinity was determined by calculating the pA<sub>2</sub>. In this study, the basic inhibition-curve protocol is used. The details have been described in Section 1.7.2. The pA<sub>2</sub> (= -logK<sub>b</sub>) is estimated from the equation (Lazareno and Birdsall, 1993b):

$$K_b = \frac{IC_{50}^*}{\frac{[A]}{EC_{50}^*} - 1}$$

and

$$K_b = \frac{[B_i]}{\frac{[A_f]}{[A_i]} - 1}$$

### 6.2.4 Statistical analysis

Contractile responses were measured as increases in tension (g) above the resting level and normalised to the second 60 mM KCl response on each artery preparation. A variable-slope sigmoidal curve was fitted to log concentration–response data using GraphPad Prism software; the bottom asymptote was constrained to zero for contraction. Sigmoidal curve parameters were derived from data for individual preparations. Data were further analysed by 1-factor and repeated measures 2-factor ANOVA combined with comparison of selected means by planned (orthogonal) contrasts using SuperANOVA software. Two means of value were compared with unpaired Student *t-test*. All tests were two-tailed and the significance level was set at  $p < 0.05$ . All data are presented as mean  $\pm$  SEM.

## 6.3 Results

### 6.3.1 General condition of animals

At the beginning of the study, the two groups of rats had a comparable body weight (200 – 250 g). Four weeks later, the control rats had gained weight ( $p = 0.003$ ), whereas STZ-treated rats exhibited considerable weight loss ( $p = 0.009$ ) (Table 6.3). The STZ-treated rats also had polyuria; the cage bed material had to be changed more frequently compared to control rats. There was a single mortality in the control group, as the animal developed an abdominal cyst at three weeks after saline injection and was terminated under Schedule 1. The femoral arteries from control animals had generous deposits of fats, whereas those from STZ-treated animals looked almost fat-free.

**Table 6.3** Comparison of body weight in control and STZ-rats at the start of the study and at four weeks

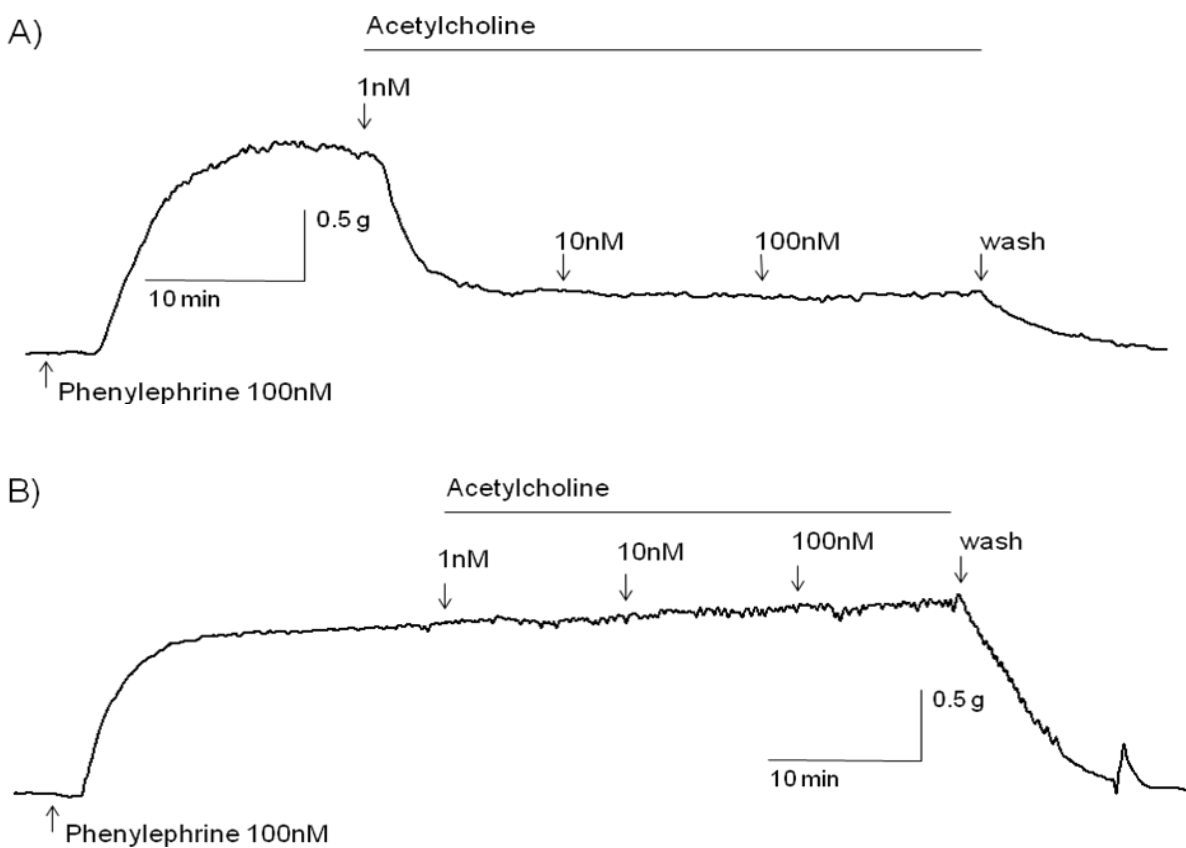
Time / duration	Control rats (g)	STZ-rats (g)
0 week (start)	218 ± 13	249 ± 26
4 weeks	367 ± 28†	186 ± 15*

\* $P < 0.05$ , 4 weeks vs 0 weeks STZ-rats

† $P < 0.05$ , 4 weeks vs 0 weeks control rats

### 6.3.2 Effects of endothelium removal on acetylcholine

In preparations with intact endothelium, acetylcholine (1 – 100  $\mu\text{M}$ ) produced a graded relaxation of the tone induced by 100 nM phenylephrine (Figure 6.3A). In comparisons, in preparation without endothelium, acetylcholine had no effect (Figure 6.3B).

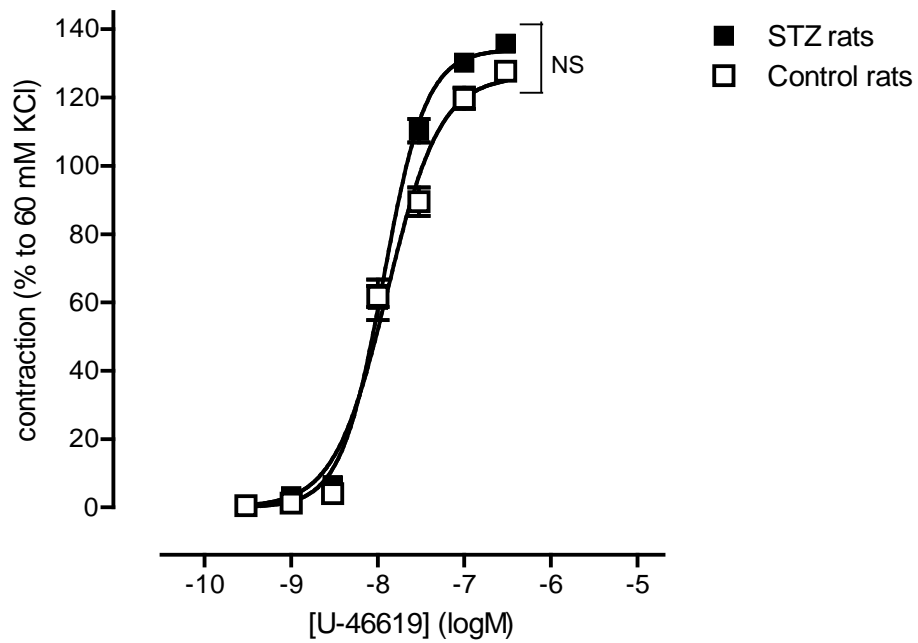


**Figure 6.3** The traces from the current study illustrating the relaxant effects of acetylcholine on 100 nM phenylephrine-induced tone on rat femoral artery. (A) The relaxant effect in artery with intact endothelium, (B) the loss of the relaxant response in artery with endothelium removed.

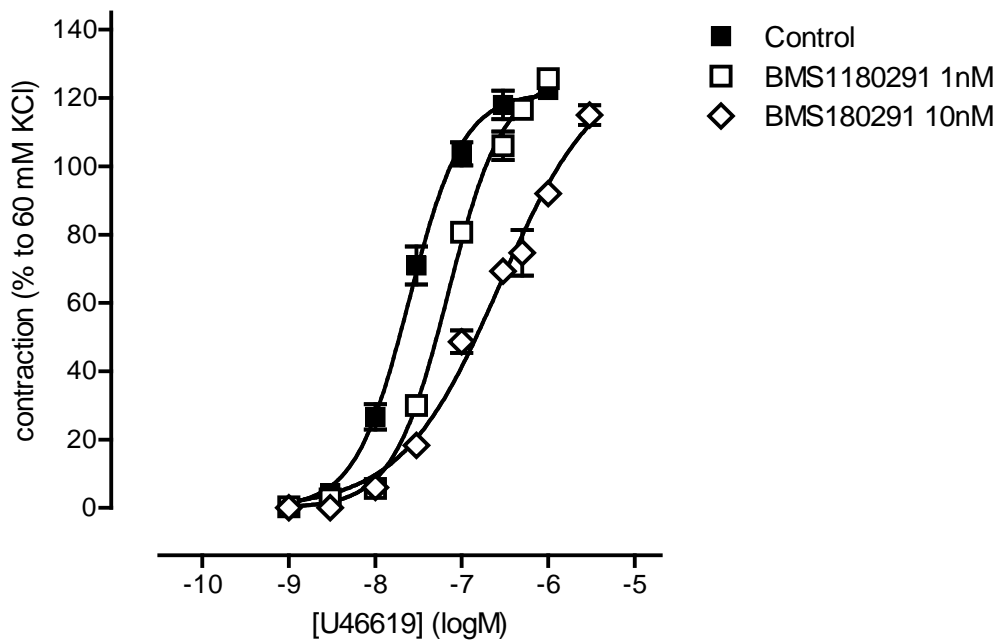
### 6.3.3 Effects of U-46619 and its block by BMS-180291

U-46619 (1 – 300 nM) potently contracted rat femoral artery preparations. The log concentration-response curves were similar in preparations from control and STZ-rats ( $E_{\max}$ :  $130.6 \pm 2\%$ ,  $125.6 \pm 5.4\%$ ;  $pEC_{50} = 7.95 \pm 0.03$ ;  $7.95 \pm 0.06$ ) (Figure 6.4,  $n = 4$ ). The main effects contrast for control and STZ treatment were not significantly different ( $p = 0.38$ ). The slope of the two curves was almost the same (control: 1.8; STZ: 1.6).

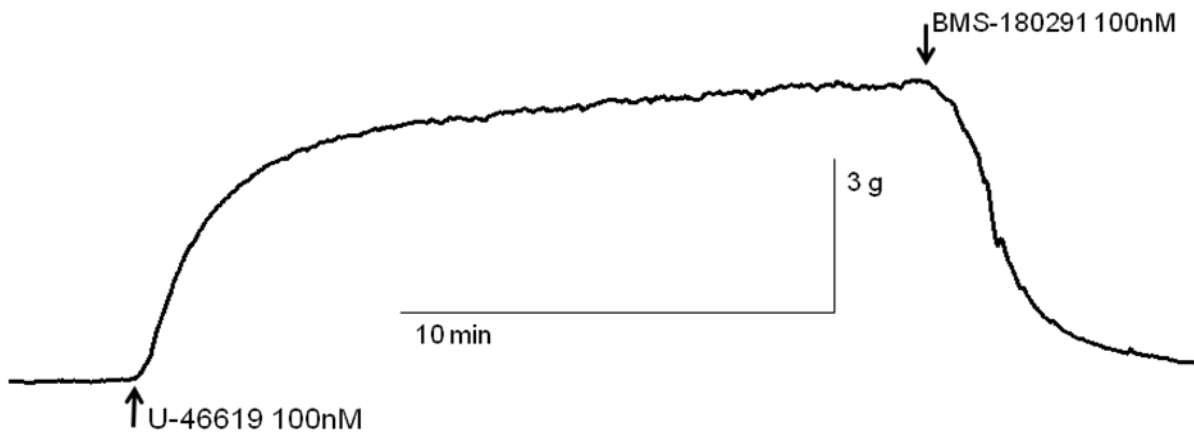
BMS-180291 at 1 and 10 nM right-shifted the log concentration-response curves for U-46619 in femoral arteries from STZ-rats (Figure 6.5,  $n = 4$ ). Dose-ratios were 3.16 and 11.2 respectively, affording  $pA_2$  values of 9.33 and 9.01. BMS-180291 at 100 nM abolished the contraction to 100 nM U-46619 (Figure 6.6). After three washouts at 15 min intervals, the preparation had recovered full sensitivity to U-46619.



**Figure 6.4** Effects of U-46619 on denuded femoral arteries from control and STZ rats ( $n = 4$ ). *NS*, no significant difference between control and STZ-rats, using main effects contrasts.



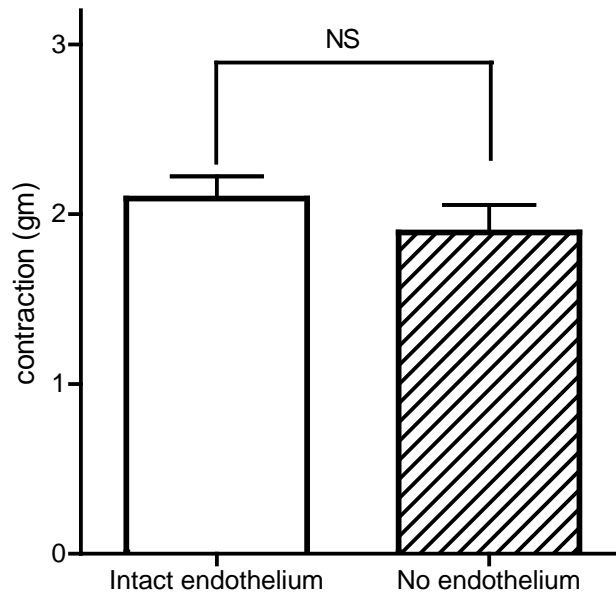
**Figure 6.5** Antagonism of U-46619-induced contraction in denuded STZ-rat femoral artery by BMS-180291. Log concentration-response curves for U-46619 following treatment with 1 nM and 10 nM SC-51322 are shown ( $n = 4$ ).



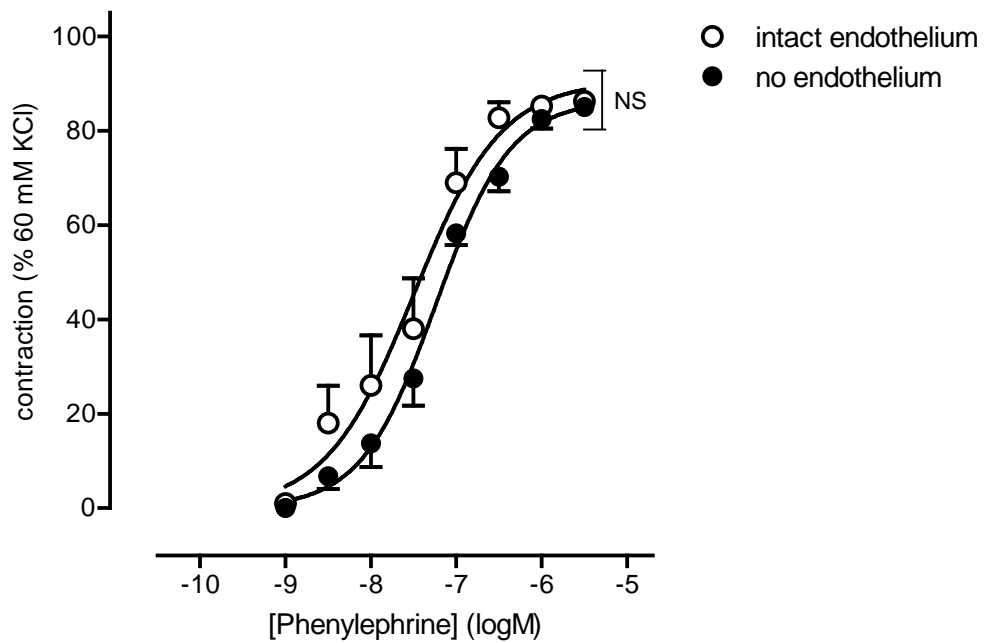
**Figure 6.6** Experimental trace illustrating the fast onset and complete inhibition by 100 nM BMS-180291 (TP antagonist) on 100 nM U-46619-induced contraction of endothelium-denuded femoral artery from STZ- rat.

### 6.3.4 Effects of endothelium on potassium chloride and phenylephrine contraction

The responses to 60 mM KCl for intact and without endothelium preparations were not significantly different in STZ-rats ( $p = 0.36$ ; Figure 6.7,  $n = 5$ ). Similar response was observed in denuded and intact endothelium artery of control rat. Thus, the contractile response in control and STZ-rats were normalised to 60 mM KCl ( $K^+$ -standard). Log concentration-response curves for phenylephrine (1 nM – 3  $\mu$ M) in STZ femoral artery preparations with and without endothelium are shown in Figure 6.8 ( $n = 4$ ). Main effects contrasts for with and without endothelium were not significantly different ( $p = 0.67$ ). ( $E_{max}$  values were  $92.9 \pm 12.0$  and  $88.3 \pm 6.2\%$ ;  $pEC_{50}$  values were  $7.46 \pm 0.21$  and  $7.21 \pm 0.10$ ; respectively).



**Figure 6.7** Contraction induced by 60 mM KCl in intact endothelium and denuded femoral arteries of STZ-rats ( $n = 5$ ). *NS*, no significant difference using Student *t*-test.

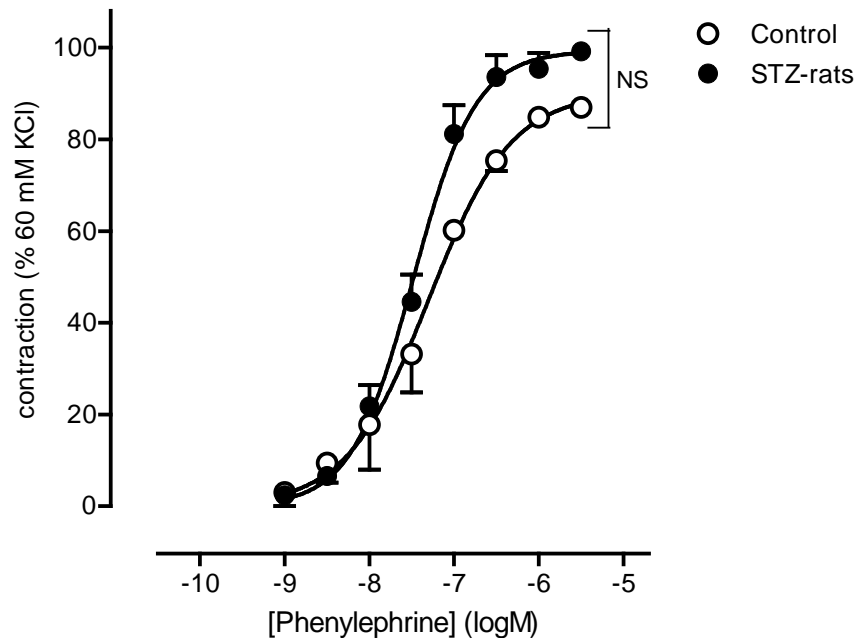


**Figure 6.8** Log concentration-response curves for phenylephrine in STZ-rat femoral artery with endothelium and without endothelium ( $n = 4$ ). *NS*, no significant difference between intact and without endothelium preparation using main effects contrasts.



### 6.3.5 Effects of phenylephrine on streptozotocin and control arteries

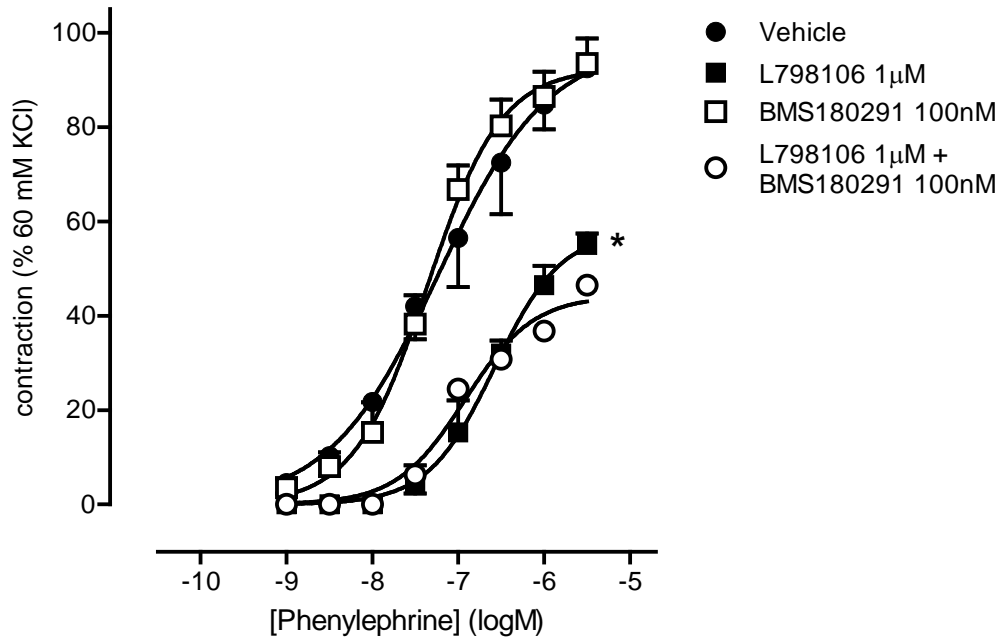
Log concentration-response curves for phenylephrine (1 – 300 nM) in endothelium-denuded femoral artery preparations from STZ and control rats are shown in Figure 6.9 ( $n = 5$ ). The denuded femoral artery from STZ-rats showed similar contractile responses to phenylephrine (1 – 300 nM) compared to control rats. In arteries from STZ-treated rats, the maximal response to phenylephrine was greater than those from control rats but statistically not significant ( $E_{\max}$ :  $88.8 \pm 2.6\%$  and  $75.4 \pm 2.6\%$  respectively,  $p = 0.47$ ). The  $pEC_{50}$  values for STZ-rats and control preparations were:  $7.39 \pm 0.07$  and control:  $7.55 \pm 0.08$  respectively ( $p = 0.98$ ).



**Figure 6.9** Log concentration-response curves for phenylephrine on denuded femoral arteries from control and STZ rats ( $n = 5$ ). *NS*, no significant difference between control and STZ-rats using main effects contrasts.

### 6.3.6 Effects of prostanoid antagonists on phenylephrine-induced contraction

The TP receptor antagonist, BMS-180291 at 100 nM did not affect the contractile response to phenylephrine in STZ preparations:  $E_{max}$ : vehicle (DMSO)  $90.3 \pm 11.5\%$ ; STZ  $92.3 \pm 7.3\%$ ;  $p = 0.63$ ) (Figure 6.10;  $n = 4$ ). The  $pEC_{50}$  values were similar between two groups (Table 6.4). In contrast, the  $EP_3$  receptor antagonist, L-798106 at 1  $\mu$ M, significantly reduced the maximal response to phenylephrine compared to vehicle ( $80.3 \pm 11.5$  vs.  $52.3 \pm 9.8\%$ ,  $p < 0.05$ ), with the curve shifted significantly to the right (Table 6.4). The addition of 100 nM BMS-180291 to 1  $\mu$ M L-798106 produced no significant changes in the two parameters ( $p = 0.87$ )



**Figure 6.10** Effects of phenylephrine in denuded STZ-rat femoral artery, in the presence of vehicle (DMSO), 1  $\mu$ M L-798106 and 100 nM BMS-180291 ( $n = 4$ ).  $*P < 0.05$ , 1  $\mu$ M L-798106 and vs. vehicle, using main effects contrasts.

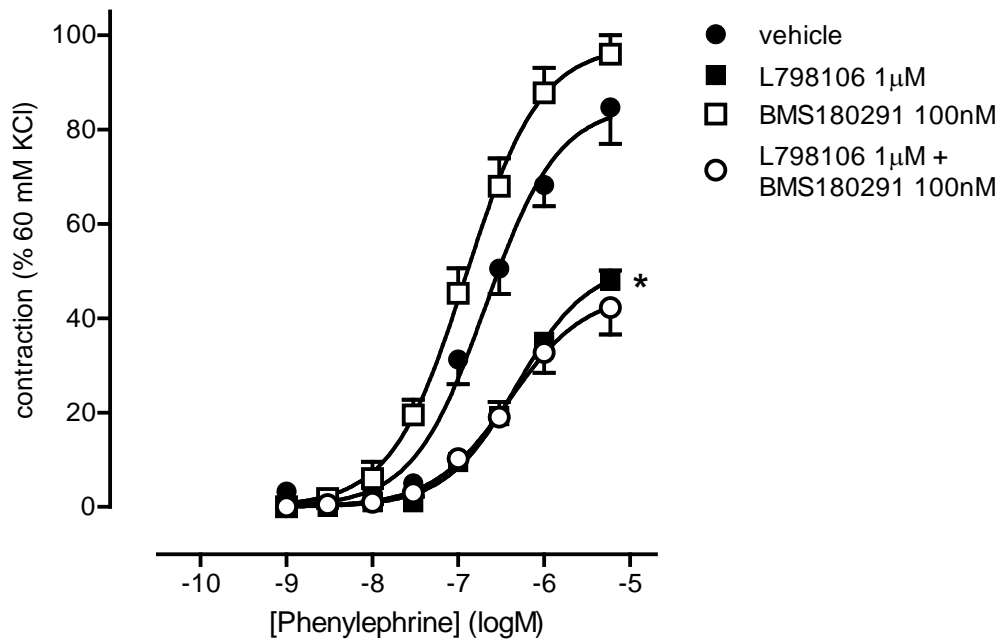
**Table 6.4** pEC<sub>50</sub> of phenylephrine response in denuded control and STZ-rats femoral artery, following treatment with L-798106 and BMS-180291.

	pEC <sub>50</sub>	
	Control	STZ
Vehicle	6.44 ± 0.04	7.17 ± 0.16
L-798106 1 μM	6.21 ± 0.09 *	6.87 ± 0.06 *
BMS-180291 100 nM	6.91 ± 0.13	7.10 ± 0.31
BMS-180291 100 nM + L-798106 1 μM	6.32 ± 0.08	7.22 ± 0.01

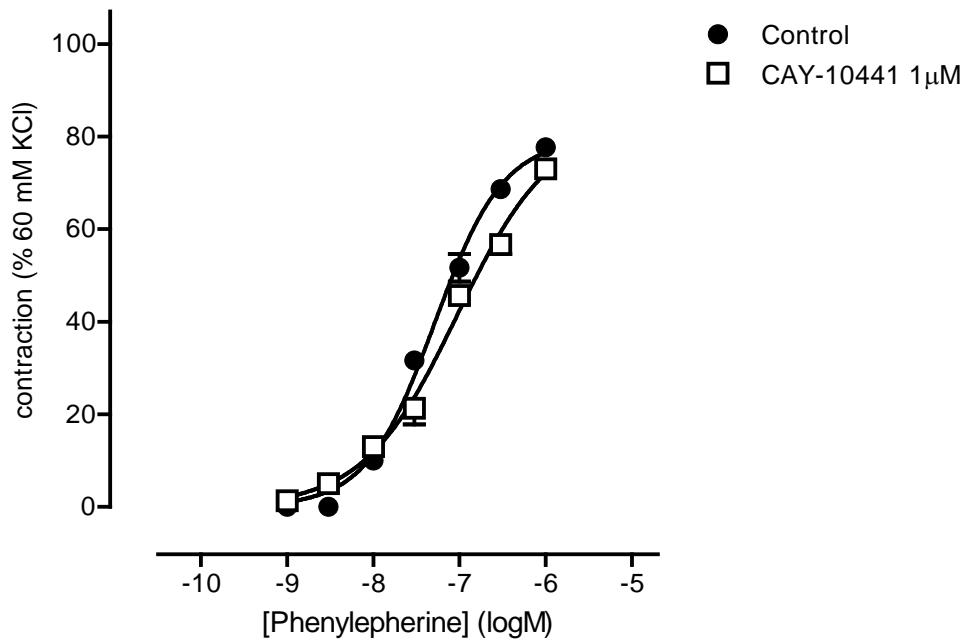
\*  $P < 0.05$ , L-798106 1 μM vs vehicle (control and STZ-rats)

The antagonist profiles were similar in control rats. Maximal response to phenylephrine in vehicle-treated preparations was no different in the presence of 100 nM BMS-180291 ( $88.3 \pm 8.2\%$ ,  $90.2 \pm 6.3\%$  respectively) (Figure 6.11,  $n = 4$ ). The pEC<sub>50</sub> values were also similar (Table 6.4). Similar to STZ-rats, 1 μM L-798106 reduced the phenylephrine maximum and shifted the concentration-response curve to the right in comparison to the control preparation. 100 nM BMS-180291 in combination with 1 μM L-798106 did not produce any significant changes compared with L-798106 alone.

CAY-10441 (IP-antagonist) at 1 μM did not affect the maximal response in diabetic rat femoral artery (Figure 6.12;  $n = 5$ ).



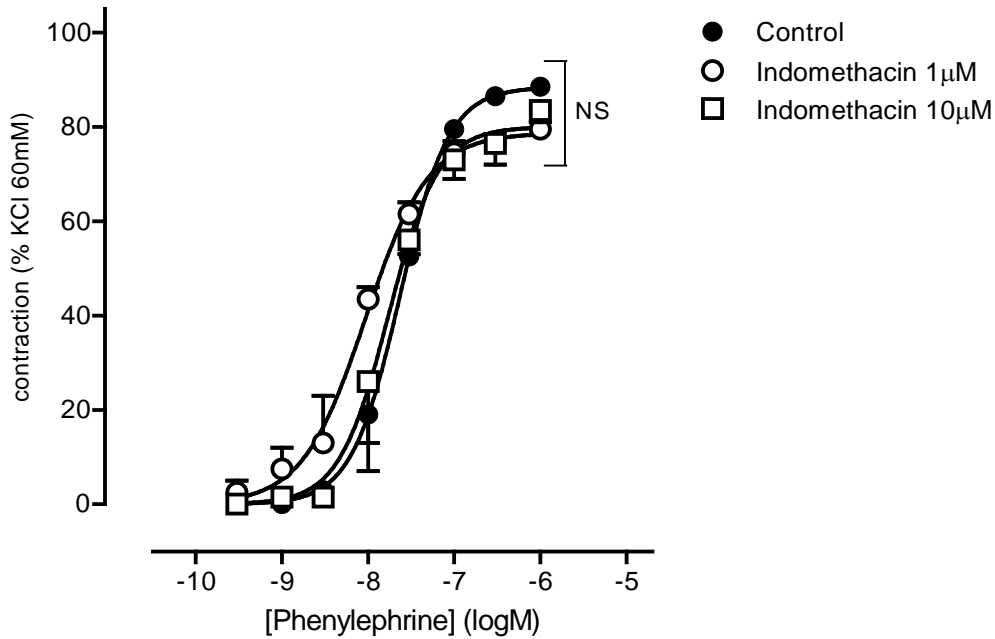
**Figure 6.11** Effects of phenylephrine in denuded control rat femoral artery following treatment with vehicle (DMSO), 1 µM L-798106 and 100 nM BMS-180291 ( $n = 4$ ).  $*P < 0.05$ , 1 µM L-798106 vs. vehicle, using main effects contrasts.



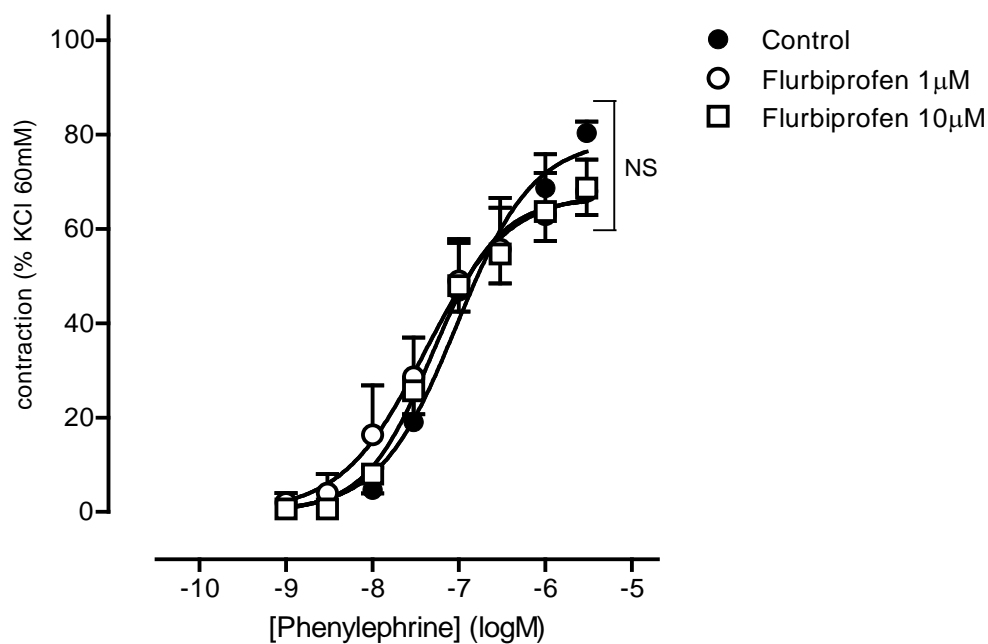
**Figure 6.12** Effects of phenylephrine in denuded STZ-rat femoral artery following treatment with 1  $\mu$ M CAY-10441 ( $n = 5$ ).

### 6.3.7 Effects of COX-inhibitors on phenylephrine responses

Indomethacin (non-specific COX inhibitors) at 1 and 10  $\mu\text{M}$  did not affect the maximal response produced by phenylephrine in non-endothelium rat femoral artery of STZ-rats (Figure 6.13,  $n = 4$ ). Similar results were found with flurbiprofen at 1 and 10  $\mu\text{M}$  (Figure 6.14,  $n = 4$ ) (Table 6.5). A small increased in spontaneous tone was observed after incubation with flurbiprofen (Figure 6.15).



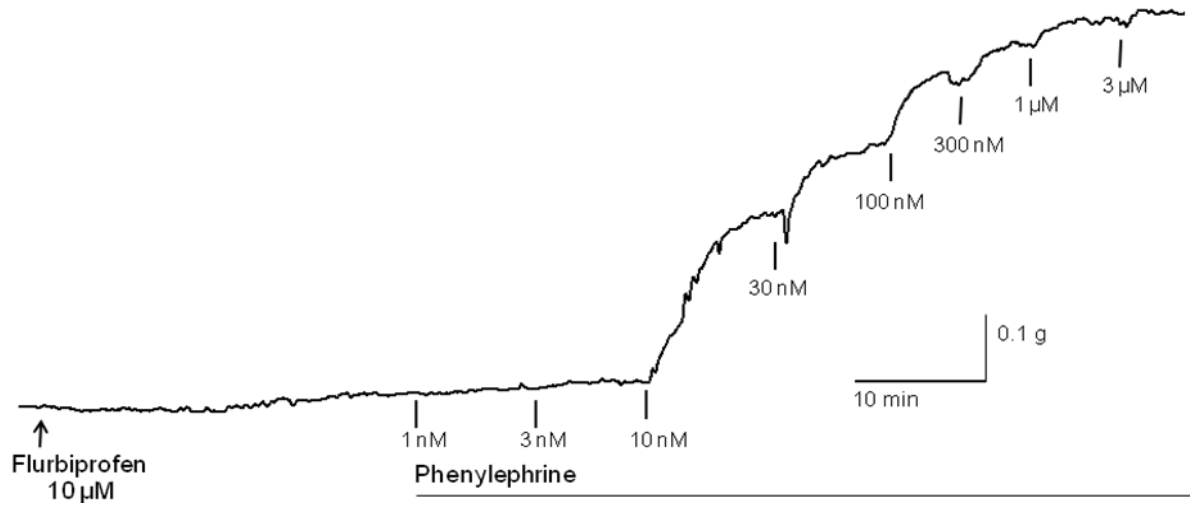
**Figure 6.13** Effects of indomethacin on the log-concentration-response curve for phenylephrine in denuded STZ-rat femoral artery ( $n = 4$ ). *NS*, not significant using main effects contrasts.



**Figure 6.14** Effects of flurbiprofen on the log-concentration-response curve for phenylephrine in denuded STZ-rat femoral artery ( $n = 4$ ). *NS*, not significant using main effects contrasts.

**Table 6.5**  $pEC_{50}$  of phenylephrine response in denuded STZ-rat femoral artery, following indomethacin and flurbiprofen.

	<b><math>pEC_{50}</math></b>
Vehicle	$7.61 \pm 0.06$
Indomethacin 1 $\mu$ M	$8.03 \pm 0.07$
Indomethacin 10 $\mu$ M	$7.75 \pm 0.08$
Vehicle	$7.35 \pm 0.16$
Flurbiprofen 1 $\mu$ M	$7.45 \pm 0.21$
Flurbiprofen 10 $\mu$ M	$7.31 \pm 0.09$



**Figure 6.15** Experimental trace illustrating the increased in spontaneous tone induced after addition of 10 μM flurbiprofen on the denuded STZ-rat femoral artery.



## **6.4 Discussion**

### **6.4.1 General response to streptozotocin**

Diabetes mellitus is an endocrine disease characterized by the lack of insulin production (Type 1) or resistance of the target-organ to insulin or insulin resistant (Type 2) (Gale, 2001). In the early stage of diabetes, the subject usually presents with increasing passing out of urine (polyuria) and increase water uptake (polydipsia). Polyuria in STZ-treated rats was evident in the current study as the cage material needed to be change more frequently. Weight loss in the STZ-rats was due to inability of the rat to utilise the carbohydrate due to lack of insulin. Instead, the body fat was utilised for energy. In the current study, the STZ-rats were noticed to be thin and the dissected femoral artery was totally devoid of any connective tissue. The symptoms and signs resemble Type 1 diabetes mellitus in human and are in agreement with previous studies on STZ-induced diabetes in the rat (Harris and MacLeod, 1988; Furman and Sneddon, 1993; Shi *et al.*, 2007a; Shi *et al.*, 2007b; Shi and Vanhoutte, 2008).

### **6.4.2 Endothelium removal and effects on contractile response**

Saponin was used initially in an attempt to remove the endothelium. Saponin is a natural product, obtained from the bark of the South American soap tree, *Quillaja saponaria*. It has been used as a chemical method to remove endothelium in dog femoral artery (Samata *et al.*, 1986) and dog coronary artery (Nakane *et al.*, 1986). Nakane *et al.* (1986) used intraluminal bolus injection of saponin (1 mg) in isolated and perfused dog coronary arteries. In the current study, the arterial rings were incubated with Krebs-Henseleit solution containing 0.3 mgml<sup>-1</sup> of saponin for 35 min at 37 °C as described earlier by Samata *et al.* (1986). However, responses to KCl and phenylephrine were abolished by this treatment. Instead, the endothelium was mechanically

removed by rubbing roughened myograph wire against the intimal surface of the artery. The method was described previously in a study of acetylcholine mechanism of action in cat cerebral artery (Lee, 1982).

In the current study, contractions to phenylephrine were similar in the presence and absence of endothelium. This agrees with similar studies of the action of noradrenaline on rat mesenteric bed (Fortes *et al.*, 1983; Peredo, 2001) and rat aorta (Fortes *et al.*, 1983; Mulhern and Docherty, 1989). The response is dependent on release of intracellular  $\text{Ca}^{2+}$  and to lesser extent on entry of extracellular  $\text{Ca}^{2+}$  (Owen and Carrier, 1980; Hung *et al.*, 2006). In contrast, others have shown that the removal of endothelium increased the maximal response and sensitivity to phenylephrine in comparison to an intact endothelium preparation in STZ-rat aorta (Martin *et al.*, 1986; Alosachie and Godfraind, 1988).

The removal of endothelium had no effect on the response to 60 mM KCl. This agreed with the previous finding in canine femoral artery where removal of endothelium cause slight depression of KCl-induced contraction (De Mey and Vanhoutte, 1981). The response was also similar between denuded artery of control and STZ-rats, in agreement with the earlier work done in diabetic rat aorta where the contractile response to KCl is endothelium independent (Mulhern and Docherty, 1989; Peredo, 2001; Tang *et al.*, 2008).

The present results demonstrated that there were no differences in response to phenylephrine in both denuded femoral artery of control and diabetes rat. This is in consistent with previous work done on rat femoral artery (Dai *et al.*, 1993; Shi *et al.*, 2007a). These results are also similar to work done on denuded diabetic rat aorta (Chang and Stevens, 1992; Tang *et al.*, 2008) and rat mesenteric arteries (Harris and MacLeod, 1988; White and Carrier, 1988; Furman and Sneddon, 1993; Peredo, 2001).

### 6.4.3 Induction of endogenous prostanoids by diabetes

The induction of endogenous prostanoids varies according to the duration of diabetes. In rat mesenteric bed, the vasoconstrictor prostanoid production (TXA<sub>2</sub> / PGF<sub>2α</sub>) were increased up to 100% from basal level in the first week of STZ-induction diabetes, (Peredo *et al.*, 1999). The levels of PGI<sub>2</sub> and PGE<sub>2</sub> were unchanged within the same period. The ratio between the production of PGI<sub>2</sub> and TXA<sub>2</sub> was recovered to reach values similar to those of the controls at four weeks (Peredo *et al.*, 1999; Shi and Vanhoutte, 2008).

There appears to be enhanced activation of the COX enzyme in arteries following induction of diabetes, leading to the production of endogenous contractile factors. In the early stage of the diabetes, the main COX product generated in rat femoral artery is TXA<sub>2</sub> (Shi *et al.*, 2007b). TXA<sub>2</sub> also shown to be increased in diabetic dog mesenteric arteries (Sterin-Borda *et al.*, 1984). TXA<sub>2</sub> mainly activates TP receptors on the smooth muscle cells as shown by the ability of S-18886 (TP receptor antagonist) to inhibit responses to phenylephrine in diabetic rat femoral artery (Shi *et al.*, 2007a).

In the current experiment, the TXA<sub>2</sub> mimetic, U-46619 produced a similar contractile profile in femoral arteries from STZ and control rats. These results suggest that in diabetes neither the sensitivity nor the density of the TP receptor is changed. Previous work using radioligand binding studies showed there were no significant differences in the density (B<sub>max</sub>) or affinity (K<sub>d</sub>) of TP receptors on isolated platelets or glomerular membranes between control and STZ-rats (Morinelli *et al.*, 1993). The current results also agreed with the view that the underlying smooth muscle is not affected by STZ diabetes (Chang and Stevens, 1992). However, BMS-180291 at 100 nM did not affect phenylephrine-induced contraction, thus arguing against a facilitatory role for endogenous TXA<sub>2</sub>. BMS-180291 (TP antagonist) competitively inhibited U-46619 with pA<sub>2</sub> of 9.33 and 9.01, corresponding to its expected affinity for the TP receptor. Furthermore, 100 nM

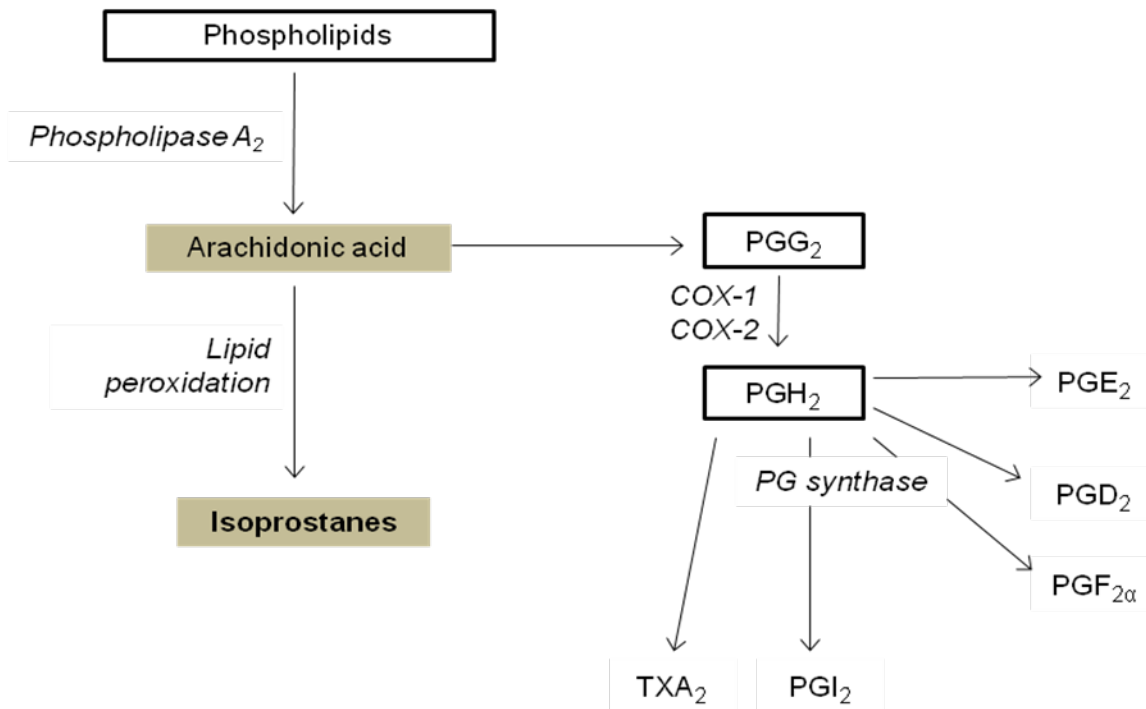
BMS-180291 completely antagonised the contractile response to 100 nM U-46619 (~ 60 - 70% maximal response) in the STZ-rats (Figure 6.6). In contrast, Shi *et al* (2007b) showed an increase in sensitivity to U-46619 but decrease in maximal response in STZ-rat femoral artery compared to the control preparation.

In the experiments of Hung *et al.* (2006), PGE<sub>2</sub> alone produced a weak contractile response of the rat femoral artery. However, when the preparation was primed with phenylephrine (10 nM, 10% of maximal response), a strong contractile effect was seen with PGE<sub>2</sub> (1 – 300 nM). 10 nM PGE<sub>2</sub> also enhanced the response to phenylephrine, demonstrating that the synergism is mutual. This synergism was blocked by 1 µM L-798106, with pA<sub>2</sub> of 7.35 – 8.10. These experiments were conducted in the presence of indomethacin, so that endogenous prostanoids did not interfere with the assessment of the exogenous prostanoid.

The current study would tend to support a facilitator role for PGE<sub>2</sub> in the contraction of rat femoral artery. In the absence of COX-inhibition, L-798106 at 1 µM inhibited the response to phenylephrine, with shift of EC<sub>50</sub> to a higher value and suppression of the maximum response. This profile is compatible with the generation of PGE<sub>2</sub> within the tissue, which then acts on EP<sub>3</sub> receptors on the smooth muscle cells to enhance α<sub>1</sub>-adrenoceptor mediated events. This profile was very similar in control and STZ-diabetic rats. However, in the current experiments, indomethacin and flurbiprofen did not affect phenylephrine action in the femoral artery from STZ-rat. This result agreed with work done by Chang *et al.* (1992) where 5 µM indomethacin did not affect the contractile response to phenylephrine in STZ-rat aorta (endothelium removed). In addition, Peredo (2001) showed no effect of 10 µM indomethacin on the contractile response to noradrenaline in STZ-rat mesenteric beds.

A possible explanation for the nil effect of indomethacin on contractile response to phenylephrine is that a non-COX pathway generates the prostaglandin or prostaglandin-like compounds. One group of compounds of interest are the isoprostanes (isoPs), prostaglandin (PG)-like compounds produced by non-enzymatic degradation (lipid peroxidation) of arachidonic acid (Roberts and Morrow, 1995; Dogne *et al.*, 2005). Examples include 8-iso PGE<sub>2</sub> and 8-iso PGF<sub>2α</sub>. This lipid peroxidation is independent of the COX enzyme in human and animals (Figure 6.16). Indeed, isoP levels increased as a consequence of enhanced lipid peroxidation in diabetes (Davi *et al.*, 1999). The plasma level of isoPs was about two-fold higher in 8 weeks STZ-rat than in control rats (Wigg *et al.*, 2004). In response to bolus injection of noradrenaline, the level of isoPs and TXA<sub>2</sub> significantly increased in human umbilical vein (Sametz *et al.*, 2000b). Thus, the same stimulation that enhanced prostaglandin production will also increase the level of isoPs in the tissue.

Previously, radio-ligand binding studies on rat smooth muscle cells have shown receptors for isoprostanes distinct from prostanoid receptors (Fukunaga *et al.*, 1993; Fukunaga *et al.*, 1997). However, in high concentration, there was a cross-activation of TP receptors. Moreover, recent studies on isolated vascular preparation have shown the contractile effect is due to activation on EP and TP receptors with no evidence for distinct isoPs receptors. IsoPs have shown to contract various tissues, including guinea-pig trachea (Clarke *et al.*, 2004), rat gastric fundus, guinea-pig ileum (Sametz *et al.*, 2000a), human umbilical vein (Daray *et al.*, 2003; Daray *et al.*, 2004) and pig pulmonary artery (Janssen and Tazzeo, 2002). The isoPs constrictive effect is also not dependent on intact endothelium as demonstrated earlier on a canine intestine preparation (Elmhurst *et al.*, 1997). In the current experiment, a TP antagonist did not affect the response to phenylephrine. In view of this result, it is most likely in STZ-rat femoral artery isoPs potentially synergise with phenylephrine through EP<sub>3</sub> system in the artery as evidenced by the inhibitory effect of L-798106.



**Figure 6.16** Phospholipids are metabolised by phospholipase A<sub>2</sub> to form arachidonic acid in response to various stimuli, like the diabetes process. Oxidation of arachidonic acid via a non-enzymatic pathway (lipid peroxidation) produces a family of isoprostanes (redrawn from Dogne *et al.*, 2005).

Another possibility for the inhibitory effect of L-798106 is a non-specific mechanism. However, it would be difficult to distinguish between, say, block of synergism induced by an endogenous non-prostanoid and a non-specific inhibitory effect since the responses to other contractile agents may be inhibited by L-798106 to the same extent. The alternative method is to use a chemically different EP<sub>3</sub> antagonist that is more selective on the same settings. For example, ONO-AE3-240 is reported to be a highly selective EP<sub>3</sub> antagonist (mouse EP<sub>3</sub> / EP<sub>1</sub> selectivity ratio = 2500; Amano *et al.*, 2003), which significantly suppressed tumor-associated angiogenesis and tumor growth in WT mice; perhaps it would show high selectivity in general.

#### **6.4.4 Roles of prostacyclin**

Rat femoral artery is relaxed by prostacyclin analogues (Hung *et al.*, 2006). Prostacyclin (PGI<sub>2</sub>) could therefore be part of the prostaglandins released, exerting its relaxant action through IP receptor (Narumiya *et al.*, 1999). The nil effect of the indomethacin and flurbiprofen could then be due to inhibition of the production of both TXA<sub>2</sub> (contractile) and PGI<sub>2</sub> (relaxant). If this is the reason, by blocking the IP receptor, the contractile response to phenylephrine in the preparation should be higher than in the absence of any antagonist. However, the IP antagonist CAY-10441 (pA<sub>2</sub> 8.82; Clark *et al.*, 2004) did not affect the maximal response to phenylephrine in the STZ-rats. Interestingly, there is a substantial evidence that in the STZ-induced diabetes rat, the production of PGI<sub>2</sub> is decreased (Harrison *et al.*, 1978; Peredo *et al.*, 1999).

## 6.5 Conclusions

The evidence for a role of endogenous prostanoid(s) in enhancing the contractility of rat femoral artery is conflicting. In both control and STZ treatment situations, the results with L-798106 suggest that an endogenous EP<sub>3</sub> agonist may enhance  $\alpha_1$ -adrenoreceptor-induced contraction. However, the COX-inhibitor data would argue against this proposal. It is possible that the endogenous facilitator could be a product of a non-COX biosynthetic pathway, such as an isoprostane. A co-role for prostacyclin acting as a functional antagonist seems unlikely in view of the lack of effect of the IP antagonist CAY-10441. Additionally, a thorough investigation of the specificity of L-798106 is required. It may be useful to repeat the current experiments using an EP<sub>3</sub> antagonist of a different chemical class.

Nevertheless, the current study has shown that there was no significant difference in response to phenylephrine in control and STZ-induced diabetes rat. The discrepancy between other published works cannot be readily explained. Several possibilities are due to variations between the duration of STZ-induced diabetes, the different tissue preparations, and sources of animal or experimental conditions.



# **CHAPTER SEVEN**

## **GENERAL DISCUSSION**

**AND**

## **CONCLUSION**

The present study has used classical organ bath pharmacology and utilised fundamental principles of pharmacology to characterize prostanoid receptors in several isolated vessels or tissues. In the process, the utility of certain novel prostanoid agonists and antagonists has been assessed.

## **7.1 Functional studies**

Organ bath studies were carried out to determine the functional responses of the guinea-pig trachea, rat mesenteric artery, rat urinary bladder and rat femoral artery to prostaglandin analogues. This was used to provide information on the population of key prostanoid receptors within these tissues.

Although Schild antagonism protocols remain the gold standard to determine the affinity constant of an antagonist, the current study has utilised inhibition-curve protocols based on the Cheng-Prusoff relationship. The decision to choose this method was based on the limited availability of prostanoid analogues as the inhibition-curve protocol is operating over a low concentration range. Furthermore, the inhibition protocol provides direct observation of the rate of onset of antagonism in the case of highly potent and / or lipophilic antagonists, which were expected to be slower in onset. The affinity constants estimated for particular antagonists were in agreement with other functional studies using Schild protocols. The inhibition-curve protocol has not been used frequently in previous studies for prostanoid receptor antagonists

In the guinea-pig trachea, the current study findings were consistent with the presence of contractile EP<sub>1</sub> and TP receptors and a relaxant EP<sub>2</sub> receptor. All the EP<sub>2</sub> agonists tested in the GPT study behaved as full agonists. CAY-10399 was shown to be the most potent EP<sub>2</sub> receptor agonist of those studied. The EP<sub>2</sub> receptor agonist AH-13205 appeared to have two relaxant

components. This profile may arise from the existence of another relaxant receptor in the trachea. Alternatively, it is possible that the two isomers present in AH-13205 may be responsible. It should be possible to separate these isomers by high-performance liquid chromatography and study each separately.

In the rat mesenteric artery, the absence of relaxant prostanoid receptors was of considerable assistance in establishing the presence of contractile TP, EP<sub>1</sub> and EP<sub>3</sub> prostanoid receptors. In addition, the use of the potent and selective TP antagonist BMS-180291 was crucial to studying only EP receptor-mediated effects. The presence of these functional receptors was confirmed by the reduction of established contractile response in the presence of the respective selective antagonists. EP<sub>1</sub> and EP<sub>3</sub> receptors were activated simultaneously by PGE<sub>2</sub> analogues such as 17-phenyl PGE<sub>2</sub> and sulprostone. The two-site competition equation fitted the inhibition-curve data significantly better than one-site competition model, as shown by a modified F test. This is the first time that this type of analysis has been used for a prostanoid system. The novel EP<sub>1</sub> agonist, ONO-D1-004 was shown to be a selective on EP<sub>1</sub> receptor. However, the activity of novel EP<sub>3</sub> agonist, ONO-AE-248 on rat mesenteric artery needs to be investigated further due to contradictory results.

In similar situation in the rat urinary bladder, only contractile EP-type receptors were shown to be present functionally. Antagonist studies demonstrated both EP<sub>1</sub> and EP<sub>3</sub> receptors contributed to the contractile response induced by the prostanoid agonists. As in the rat mesenteric artery, inhibition-curve protocols underpinned the finding of two contractile systems. Again, the two-site competition model fitted better than the one-site competition model. Non-parallel right-shifts of EP agonist curves were found with the novel EP<sub>1</sub> antagonist, GW-848687. The profile GW-844687 may be due to presence of two contractile receptors or non-competitive antagonism. In relation to the latter, a state of hemi-equilibrium for the antagonist-agonist interaction needs to be considered.

The current studies were able to characterize the prostanoid receptors by utilising the inhibition curve protocol instead of full Gaddum-Schild protocol. Further studies, in availability of the more selective prostanoid would help to clarify the uncertainty results in this study. Similarly, binding studies on this isolated preparation could be performed to allow identification of the specific receptors that the analogues bind.

The studies reported here were carried out over the same time period as related work on several guinea-pig smooth muscle preparations by one of my supervisors. The guinea-pig trachea (EP<sub>1</sub> and TP preparation) and aorta rings (EP<sub>3</sub> and TP preparation) were used in the experiments in a conventional organ bath. The experiments were often plagued by the slow approach of prostanoid agonists and antagonists to steady state, particularly when the ligands showed high potency or affinity. Thus, inhibition-curve protocols could not be performed satisfactorily. In addition, it is never clear whether the slow onset has affected the absolute potency of the ligand under study. Onset rates of both agonists and antagonists in my experiments were much faster, thereby allowing construction of cumulative relationships within a reasonable time period. This difference may be due to the thinner nature of my tissues, which allows rapid diffusion to centrally situated receptors.

The use of conventional organ bath experiments has declined of late in favour of recombinant receptor studies, usually involving radioligand binding. The transfer of these data to complex *in vivo* functional systems soon follows. The results are often equivocal however. Part of the problem relates to a lack of the sorts of information that isolated tissue studies readily provide. For example: the true potency and selectivity of the agent when a complex functional response is measured and opposing transduction systems are present; the rate of attainment of steady-state when access to the receptor pool is not instantaneous as it is the case for receptors located in the cell membrane of a carrier cell. It is hoped that some of the data contained in this thesis will help to bridge this gap as far as prostanoid systems are concerned.

## 7.2 Streptozotocin-induced diabetes study

Optimising the diabetic state of the experimental rat is a crucial aspect in determining the effect of the diabetic disease process on the arterial response to prostaglandin analogues. The period of four weeks after the administration of streptozotocin was chosen as the cut-off point in the current experiment, based on previous studies by various authors.

In STZ study, attention has been focused on the TP and EP<sub>3</sub> receptors in rat femoral artery. The rationale for this was that previous studies in the same setting in the diabetic rat only concentrated on the TP receptor /  $\alpha_1$ -adrenoceptor interaction and failed to take into account that an EP<sub>3</sub> agonist (e.g. PGE<sub>2</sub> in the *in vivo* situation) can markedly enhance the contractile actions of other agonists (including noradrenaline *in vivo*). The presence of TP receptor in the study was demonstrated by complete inhibition of U-46619-induced contraction by the selective TP antagonist BMS-180291. In addition, the study also focused on the effect of the endothelium on the phenylephrine-induced response. The endothelium was removed mechanically, by rolling the intima side of the femoral artery in order to remove the effects of endothelial dysfunction secondary to diabetes. Another method involving chemical removal using saponin proved difficult to achieve.

The current study demonstrated that the contractile response of KCl and phenylephrine in rat femoral artery were endothelium independent, in agreement with other studies. The response to phenylephrine was partially inhibited by the selective EP<sub>3</sub> antagonists L-798106 and L-826266, implying that the tissue continuously generates PGE<sub>2</sub> or a similar endogenous EP<sub>3</sub> agonist. Indeed, it is known that diabetes itself induces the production of the endogenous prostaglandins. However, two COX-inhibitors, indomethacin and flurbiprofen, failed to produce a similar inhibition of the phenylephrine response. *Per se*, this finding does not rule out a role for EP<sub>3</sub> receptors in controlling the sensitivity of the  $\alpha_1$ -adrenoceptor (or other) contractile systems.

There is still the possibility that compounds with EP<sub>3</sub> agonist properties are generated through a non-COX pathway. Another possibility, namely that endogenous PGI<sub>2</sub> acting via IP receptors opposes the contractile action of the endogenous EP<sub>3</sub> agonist, was ruled out through the use of a selective IP antagonist.

The time factor was the main limitation of why the duration of four weeks STZ-induction was chosen for the study. In reviewing of the results obtained, there were no significant differences between the contractility of the femoral arteries from control and STZ-induced diabetes rat. Future work should include a longer duration of the diabetes state with increased availability of the STZ-rats. This is necessary to provide greater understanding of the effects of diabetes on prostanoid production and the vascular response to the prolonged diabetes.

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# APPENDIX

**Table of F-statistics  $P = 0.05$**

df2\df1	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	22	24	26	28	30	35
3	10.13	9.55	9.28	9.12	9.01	8.94	8.89	8.85	8.81	8.79	8.76	8.74	8.73	8.71	8.70	8.69	8.68	8.67	8.67	8.66	8.65	8.64	8.63	8.62	8.62	8.60
4	7.71	6.94	6.59	6.39	6.26	6.16	6.09	6.04	6.00	5.96	5.94	5.91	5.89	5.87	5.86	5.84	5.83	5.82	5.81	5.80	5.79	5.77	5.76	5.75	5.75	5.73
5	6.61	5.79	5.41	5.19	5.05	4.95	4.88	4.82	4.77	4.74	4.70	4.68	4.66	4.64	4.62	4.60	4.59	4.58	4.57	4.56	4.54	4.53	4.52	4.50	4.50	4.48
6	5.99	5.14	4.76	4.53	4.39	4.28	4.21	4.15	4.10	4.06	4.03	4.00	3.98	3.96	3.94	3.92	3.91	3.90	3.88	3.87	3.86	3.84	3.83	3.82	3.81	3.79
7	5.59	4.74	4.35	4.12	3.97	3.87	3.79	3.73	3.68	3.64	3.60	3.57	3.55	3.53	3.51	3.49	3.48	3.47	3.46	3.44	3.43	3.41	3.40	3.39	3.38	3.36
8	5.32	4.46	4.07	3.84	3.69	3.58	3.50	3.44	3.39	3.35	3.31	3.28	3.26	3.24	3.22	3.20	3.19	3.17	3.16	3.15	3.13	3.12	3.10	3.09	3.08	3.06
9	5.12	4.26	3.86	3.63	3.48	3.37	3.29	3.23	3.18	3.14	3.10	3.07	3.05	3.03	3.01	2.99	2.97	2.96	2.95	2.94	2.92	2.90	2.89	2.87	2.86	2.84
10	4.96	4.10	3.71	3.48	3.33	3.22	3.14	3.07	3.02	2.98	2.94	2.91	2.89	2.86	2.85	2.83	2.81	2.80	2.79	2.77	2.75	2.74	2.72	2.71	2.70	2.68
11	4.84	3.98	3.59	3.36	3.20	3.09	3.01	2.95	2.90	2.85	2.82	2.79	2.76	2.74	2.72	2.70	2.69	2.67	2.66	2.65	2.63	2.61	2.59	2.58	2.57	2.55
12	4.75	3.89	3.49	3.26	3.11	3.00	2.91	2.85	2.80	2.75	2.72	2.69	2.66	2.64	2.62	2.60	2.58	2.57	2.56	2.54	2.52	2.51	2.49	2.48	2.47	2.44
13	4.67	3.81	3.41	3.18	3.03	2.92	2.83	2.77	2.71	2.67	2.63	2.60	2.58	2.55	2.53	2.51	2.50	2.48	2.47	2.46	2.44	2.42	2.41	2.39	2.38	2.36
14	4.60	3.74	3.34	3.11	2.96	2.85	2.76	2.70	2.65	2.60	2.57	2.53	2.51	2.48	2.46	2.44	2.43	2.41	2.40	2.39	2.37	2.35	2.33	2.32	2.31	2.28
15	4.54	3.68	3.29	3.06	2.90	2.79	2.71	2.64	2.59	2.54	2.51	2.48	2.45	2.42	2.40	2.38	2.37	2.35	2.34	2.33	2.31	2.29	2.27	2.26	2.25	2.22
16	4.49	3.63	3.24	3.01	2.85	2.74	2.66	2.59	2.54	2.49	2.46	2.42	2.40	2.37	2.35	2.33	2.32	2.30	2.29	2.28	2.25	2.24	2.22	2.21	2.19	2.17
17	4.45	3.59	3.20	2.96	2.81	2.70	2.61	2.55	2.49	2.45	2.41	2.38	2.35	2.33	2.31	2.29	2.27	2.26	2.24	2.23	2.21	2.19	2.17	2.16	2.15	2.12
18	4.41	3.55	3.16	2.93	2.77	2.66	2.58	2.51	2.46	2.41	2.37	2.34	2.31	2.29	2.27	2.25	2.23	2.22	2.20	2.19	2.17	2.15	2.13	2.12	2.11	2.08
19	4.38	3.52	3.13	2.90	2.74	2.63	2.54	2.48	2.42	2.38	2.34	2.31	2.28	2.26	2.23	2.21	2.20	2.18	2.17	2.16	2.13	2.11	2.10	2.08	2.07	2.05
20	4.35	3.49	3.10	2.87	2.71	2.60	2.51	2.45	2.39	2.35	2.31	2.28	2.25	2.23	2.20	2.18	2.17	2.15	2.14	2.12	2.10	2.08	2.07	2.05	2.04	2.01
22	4.30	3.44	3.05	2.82	2.66	2.55	2.46	2.40	2.34	2.30	2.26	2.23	2.20	2.17	2.15	2.13	2.11	2.10	2.08	2.07	2.05	2.03	2.01	2.00	1.98	1.96
24	4.26	3.40	3.01	2.78	2.62	2.51	2.42	2.36	2.30	2.25	2.22	2.18	2.15	2.13	2.11	2.09	2.07	2.05	2.04	2.03	2.00	1.98	1.97	1.95	1.94	1.91
26	4.23	3.37	2.98	2.74	2.59	2.47	2.39	2.32	2.27	2.22	2.18	2.15	2.12	2.09	2.07	2.05	2.03	2.02	2.00	1.99	1.97	1.95	1.93	1.91	1.90	1.87
28	4.20	3.34	2.95	2.71	2.56	2.45	2.36	2.29	2.24	2.19	2.15	2.12	2.09	2.06	2.04	2.02	2.00	1.99	1.97	1.96	1.93	1.91	1.90	1.88	1.87	1.84
30	4.17	3.32	2.92	2.69	2.53	2.42	2.33	2.27	2.21	2.16	2.13	2.09	2.06	2.04	2.01	1.99	1.98	1.96	1.95	1.93	1.91	1.89	1.87	1.85	1.84	1.81
35	4.12	3.27	2.87	2.64	2.49	2.37	2.29	2.22	2.16	2.11	2.08	2.04	2.01	1.99	1.96	1.94	1.92	1.91	1.89	1.88	1.85	1.83	1.82	1.80	1.79	1.76
40	4.08	3.23	2.84	2.61	2.45	2.34	2.25	2.18	2.12	2.08	2.04	2.00	1.97	1.95	1.92	1.90	1.89	1.87	1.85	1.84	1.81	1.79	1.77	1.76	1.74	1.72
45	4.06	3.20	2.81	2.58	2.42	2.31	2.22	2.15	2.10	2.05	2.01	1.97	1.94	1.92	1.89	1.87	1.86	1.84	1.82	1.81	1.78	1.76	1.74	1.73	1.71	1.68
50	4.03	3.18	2.79	2.56	2.40	2.29	2.20	2.13	2.07	2.03	1.99	1.95	1.92	1.89	1.87	1.85	1.83	1.81	1.80	1.78	1.76	1.74	1.72	1.70	1.69	1.66
60	4.00	3.15	2.76	2.53	2.37	2.25	2.17	2.10	2.04	1.99	1.95	1.92	1.89	1.86	1.84	1.82	1.80	1.78	1.76	1.75	1.72	1.70	1.68	1.66	1.65	1.62
70	3.98	3.13	2.74	2.50	2.35	2.23	2.14	2.07	2.02	1.97	1.93	1.89	1.86	1.84	1.81	1.79	1.77	1.75	1.74	1.72	1.70	1.67	1.65	1.64	1.62	1.59
80	3.96	3.11	2.72	2.49	2.33	2.21	2.13	2.06	2.00	1.95	1.91	1.88	1.84	1.82	1.79	1.77	1.75	1.73	1.72	1.70	1.68	1.65	1.63	1.62	1.60	1.57
100	3.94	3.09	2.70	2.46	2.31	2.19	2.10	2.03	1.97	1.93	1.89	1.85	1.82	1.79	1.77	1.75	1.73	1.71	1.69	1.68	1.65	1.63	1.61	1.59	1.57	1.54
200	3.89	3.04	2.65	2.42	2.26	2.14	2.06	1.98	1.93	1.88	1.84	1.80	1.77	1.74	1.72	1.69	1.67	1.66	1.64	1.62	1.60	1.57	1.55	1.53	1.52	1.48
500	3.86	3.01	2.62	2.39	2.23	2.12	2.03	1.96	1.90	1.85	1.81	1.77	1.74	1.71	1.69	1.66	1.64	1.62	1.61	1.59	1.56	1.54	1.52	1.50	1.48	1.45
1000	3.85	3.00	2.61	2.38	2.22	2.11	2.02	1.95	1.89	1.84	1.80	1.76	1.73	1.70	1.68	1.65	1.63	1.61	1.60	1.58	1.55	1.53	1.51	1.49	1.47	1.43
>1000	1.04	3.00	2.61	2.37	2.21	2.10	2.01	1.94	1.88	1.83	1.79	1.75	1.72	1.69	1.67	1.64	1.62	1.61	1.59	1.57	1.54	1.52	1.50	1.48	1.46	1.42
df2/df1	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	22	24	26	28	30	35



**Table of F-statistics  $P = 0.01$**

df2\df1	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	22	24	26	28	30	35
3	34.12	30.82	29.46	28.71	28.24	27.91	27.67	27.49	27.35	27.23	27.13	27.05	26.98	26.92	26.87	26.83	26.79	26.75	26.72	26.69	26.64	26.60	26.56	26.53	26.50	26.45
4	21.20	18.00	16.69	15.98	15.52	15.21	14.98	14.80	14.66	14.55	14.45	14.37	14.31	14.25	14.20	14.15	14.11	14.08	14.05	14.02	13.97	13.93	13.89	13.86	13.84	13.79
5	16.26	13.27	12.06	11.39	10.97	10.67	10.46	10.29	10.16	10.05	9.96	9.89	9.82	9.77	9.72	9.68	9.64	9.61	9.58	9.55	9.51	9.47	9.43	9.40	9.38	9.33
6	13.75	10.92	9.78	9.15	8.75	8.47	8.26	8.10	7.98	7.87	7.79	7.72	7.66	7.61	7.56	7.52	7.48	7.45	7.42	7.40	7.35	7.31	7.28	7.25	7.23	7.18
7	12.25	9.55	8.45	7.85	7.46	7.19	6.99	6.84	6.72	6.62	6.54	6.47	6.41	6.36	6.31	6.28	6.24	6.21	6.18	6.16	6.11	6.07	6.04	6.02	5.99	5.94
8	11.26	8.65	7.59	7.01	6.63	6.37	6.18	6.03	5.91	5.81	5.73	5.67	5.61	5.56	5.52	5.48	5.44	5.41	5.38	5.36	5.32	5.28	5.25	5.22	5.20	5.15
9	10.56	8.02	6.99	6.42	6.06	5.80	5.61	5.47	5.35	5.26	5.18	5.11	5.05	5.01	4.96	4.92	4.89	4.86	4.83	4.81	4.77	4.73	4.70	4.67	4.65	4.60
10	10.04	7.56	6.55	5.99	5.64	5.39	5.20	5.06	4.94	4.85	4.77	4.71	4.65	4.60	4.56	4.52	4.49	4.46	4.43	4.41	4.36	4.33	4.30	4.27	4.25	4.20
11	9.65	7.21	6.22	5.67	5.32	5.07	4.89	4.74	4.63	4.54	4.46	4.40	4.34	4.29	4.25	4.21	4.18	4.15	4.12	4.10	4.06	4.02	3.99	3.96	3.94	3.89
12	9.33	6.93	5.95	5.41	5.06	4.82	4.64	4.50	4.39	4.30	4.22	4.16	4.10	4.05	4.01	3.97	3.94	3.91	3.88	3.86	3.82	3.78	3.75	3.72	3.70	3.65
13	9.07	6.70	5.74	5.21	4.86	4.62	4.44	4.30	4.19	4.10	4.02	3.96	3.91	3.86	3.82	3.78	3.75	3.72	3.69	3.66	3.62	3.59	3.56	3.53	3.51	3.46
14	8.86	6.51	5.56	5.04	4.70	4.46	4.28	4.14	4.03	3.94	3.86	3.80	3.75	3.70	3.66	3.62	3.59	3.56	3.53	3.51	3.46	3.43	3.40	3.37	3.35	3.30
15	8.68	6.36	5.42	4.89	4.56	4.32	4.14	4.00	3.89	3.80	3.73	3.67	3.61	3.56	3.52	3.49	3.45	3.42	3.40	3.37	3.33	3.29	3.26	3.24	3.21	3.17
16	8.53	6.23	5.29	4.77	4.44	4.20	4.03	3.89	3.78	3.69	3.62	3.55	3.50	3.45	3.41	3.37	3.34	3.31	3.28	3.26	3.22	3.18	3.15	3.12	3.10	3.05
17	8.40	6.11	5.19	4.67	4.34	4.10	3.93	3.79	3.68	3.59	3.52	3.46	3.40	3.35	3.31	3.27	3.24	3.21	3.19	3.16	3.12	3.08	3.05	3.03	3.00	2.96
18	8.29	6.01	5.09	4.58	4.25	4.01	3.84	3.71	3.60	3.51	3.43	3.37	3.32	3.27	3.23	3.19	3.16	3.13	3.10	3.08	3.03	3.00	2.97	2.94	2.92	2.87
19	8.19	5.93	5.01	4.50	4.17	3.94	3.77	3.63	3.52	3.43	3.36	3.30	3.24	3.19	3.15	3.12	3.08	3.05	3.03	3.00	2.96	2.92	2.89	2.87	2.84	2.80
20	8.10	5.85	4.94	4.43	4.10	3.87	3.70	3.56	3.46	3.37	3.29	3.23	3.18	3.13	3.09	3.05	3.02	2.99	2.96	2.94	2.90	2.86	2.83	2.80	2.78	2.73
22	7.95	5.72	4.82	4.31	3.99	3.76	3.59	3.45	3.35	3.26	3.18	3.12	3.07	3.02	2.98	2.94	2.91	2.88	2.85	2.83	2.78	2.75	2.72	2.69	2.67	2.62
24	7.82	5.61	4.72	4.22	3.90	3.67	3.50	3.36	3.26	3.17	3.09	3.03	2.98	2.93	2.89	2.85	2.82	2.79	2.76	2.74	2.70	2.66	2.63	2.60	2.58	2.53
26	7.72	5.53	4.64	4.14	3.82	3.59	3.42	3.29	3.18	3.09	3.02	2.96	2.90	2.86	2.82	2.78	2.75	2.72	2.69	2.66	2.62	2.58	2.55	2.53	2.50	2.45
28	7.64	5.45	4.57	4.07	3.75	3.53	3.36	3.23	3.12	3.03	2.96	2.90	2.84	2.79	2.75	2.72	2.68	2.65	2.63	2.60	2.56	2.52	2.49	2.46	2.44	2.39
30	7.56	5.39	4.51	4.02	3.70	3.47	3.30	3.17	3.07	2.98	2.91	2.84	2.79	2.74	2.70	2.66	2.63	2.60	2.57	2.55	2.51	2.47	2.44	2.41	2.39	2.34
35	7.42	5.27	4.40	3.91	3.59	3.37	3.20	3.07	2.96	2.88	2.80	2.74	2.69	2.64	2.60	2.56	2.53	2.50	2.47	2.44	2.40	2.36	2.33	2.31	2.28	2.23
40	7.31	5.18	4.31	3.83	3.51	3.29	3.12	2.99	2.89	2.80	2.73	2.66	2.61	2.56	2.52	2.48	2.45	2.42	2.39	2.37	2.33	2.29	2.26	2.23	2.20	2.15
45	7.23	5.11	4.25	3.77	3.45	3.23	3.07	2.94	2.83	2.74	2.67	2.61	2.55	2.51	2.46	2.43	2.39	2.36	2.34	2.31	2.27	2.23	2.20	2.17	2.14	2.09
50	7.17	5.06	4.20	3.72	3.41	3.19	3.02	2.89	2.79	2.70	2.63	2.56	2.51	2.46	2.42	2.38	2.35	2.32	2.29	2.27	2.22	2.18	2.15	2.12	2.10	2.05
60	7.08	4.98	4.13	3.65	3.34	3.12	2.95	2.82	2.72	2.63	2.56	2.50	2.44	2.39	2.35	2.31	2.28	2.25	2.22	2.20	2.15	2.12	2.08	2.05	2.03	1.98
70	7.01	4.92	4.07	3.60	3.29	3.07	2.91	2.78	2.67	2.59	2.51	2.45	2.40	2.35	2.31	2.27	2.23	2.20	2.18	2.15	2.11	2.07	2.03	2.01	1.98	1.93
80	6.96	4.88	4.04	3.56	3.26	3.04	2.87	2.74	2.64	2.55	2.48	2.42	2.36	2.31	2.27	2.23	2.20	2.17	2.14	2.12	2.07	2.03	2.00	1.97	1.94	1.89
100	6.90	4.82	3.98	3.51	3.21	2.99	2.82	2.69	2.59	2.50	2.43	2.37	2.31	2.27	2.22	2.19	2.15	2.12	2.09	2.07	2.02	1.98	1.95	1.92	1.89	1.84
200	6.76	4.71	3.88	3.41	3.11	2.89	2.73	2.60	2.50	2.41	2.34	2.27	2.22	2.17	2.13	2.09	2.06	2.03	2.00	1.97	1.93	1.89	1.85	1.82	1.79	1.74
500	6.69	4.65	3.82	3.36	3.05	2.84	2.68	2.55	2.44	2.36	2.28	2.22	2.17	2.12	2.07	2.04	2.00	1.97	1.94	1.92	1.87	1.83	1.79	1.76	1.74	1.68
1000	6.66	4.63	3.80	3.34	3.04	2.82	2.66	2.53	2.43	2.34	2.27	2.20	2.15	2.10	2.06	2.02	1.98	1.95	1.92	1.90	1.85	1.81	1.77	1.74	1.72	1.66
>1000	1.04	4.61	3.78	3.32	3.02	2.80	2.64	2.51	2.41	2.32	2.25	2.19	2.13	2.08	2.04	2.00	1.97	1.94	1.91	1.88	1.83	1.79	1.76	1.73	1.70	1.64
df2\df1	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	22	24	26	28	30	35

**Table of F-statistics  $P = 0.001$**

df2\df1	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	22	24	26	28	30	35
3	167.03	148.50	141.11	137.10	134.58	132.85	131.59	130.62	129.86	129.25	128.74	128.32	127.96	127.65	127.38	127.14	126.93	126.74	126.57	126.42	126.16	125.94	125.75	125.59	125.45	125.17
4	74.14	61.25	56.18	53.44	51.71	50.53	49.66	49.00	48.48	48.05	47.71	47.41	47.16	46.95	46.76	46.60	46.45	46.32	46.21	46.10	45.92	45.77	45.64	45.53	45.43	45.24
5	47.18	37.12	33.20	31.09	29.75	28.84	28.16	27.65	27.25	26.92	26.65	26.42	26.22	26.06	25.91	25.78	25.67	25.57	25.48	25.40	25.25	25.13	25.03	24.95	24.87	24.72
6	35.51	27.00	23.70	21.92	20.80	20.03	19.46	19.03	18.69	18.41	18.18	17.99	17.83	17.68	17.56	17.45	17.35	17.27	17.19	17.12	17.00	16.90	16.81	16.74	16.67	16.54
7	29.25	21.69	18.77	17.20	16.21	15.52	15.02	14.63	14.33	14.08	13.88	13.71	13.56	13.43	13.32	13.23	13.14	13.06	12.99	12.93	12.82	12.73	12.66	12.59	12.53	12.41
8	25.42	18.49	15.83	14.39	13.49	12.86	12.40	12.05	11.77	11.54	11.35	11.20	11.06	10.94	10.84	10.75	10.67	10.60	10.54	10.48	10.38	10.30	10.22	10.16	10.11	10.00
9	22.86	16.39	13.90	12.56	11.71	11.13	10.70	10.37	10.11	9.89	9.72	9.57	9.44	9.33	9.24	9.15	9.08	9.01	8.95	8.90	8.80	8.72	8.66	8.60	8.55	8.45
10	21.04	14.91	12.55	11.28	10.48	9.93	9.52	9.20	8.96	8.75	8.59	8.45	8.33	8.22	8.13	8.05	7.98	7.91	7.86	7.80	7.71	7.64	7.57	7.52	7.47	7.37
11	19.69	13.81	11.56	10.35	9.58	9.05	8.66	8.36	8.12	7.92	7.76	7.63	7.51	7.41	7.32	7.24	7.18	7.11	7.06	7.01	6.92	6.85	6.79	6.73	6.68	6.59
12	18.64	12.97	10.80	9.63	8.89	8.38	8.00	7.71	7.48	7.29	7.14	7.01	6.89	6.79	6.71	6.63	6.57	6.51	6.45	6.41	6.32	6.25	6.19	6.14	6.09	6.00
13	17.82	12.31	10.21	9.07	8.35	7.86	7.49	7.21	6.98	6.80	6.65	6.52	6.41	6.31	6.23	6.16	6.09	6.03	5.98	5.93	5.85	5.78	5.72	5.67	5.63	5.54
14	17.14	11.78	9.73	8.62	7.92	7.44	7.08	6.80	6.58	6.40	6.26	6.13	6.02	5.93	5.85	5.78	5.71	5.66	5.60	5.56	5.48	5.41	5.35	5.30	5.25	5.17
15	16.59	11.34	9.34	8.25	7.57	7.09	6.74	6.47	6.26	6.08	5.94	5.81	5.71	5.62	5.54	5.46	5.40	5.35	5.29	5.25	5.17	5.10	5.04	4.99	4.95	4.86
16	16.12	10.97	9.01	7.94	7.27	6.81	6.46	6.20	5.98	5.81	5.67	5.55	5.44	5.35	5.27	5.21	5.14	5.09	5.04	4.99	4.91	4.85	4.79	4.74	4.70	4.61
17	15.72	10.66	8.73	7.68	7.02	6.56	6.22	5.96	5.75	5.58	5.44	5.32	5.22	5.13	5.05	4.99	4.92	4.87	4.82	4.78	4.70	4.63	4.58	4.53	4.48	4.40
18	15.38	10.39	8.49	7.46	6.81	6.36	6.02	5.76	5.56	5.39	5.25	5.13	5.03	4.94	4.87	4.80	4.74	4.68	4.63	4.59	4.51	4.45	4.39	4.34	4.30	4.22
19	15.08	10.16	8.28	7.27	6.62	6.18	5.85	5.59	5.39	5.22	5.08	4.97	4.87	4.78	4.70	4.64	4.58	4.52	4.47	4.43	4.35	4.29	4.23	4.19	4.14	4.06
20	14.82	9.95	8.10	7.10	6.46	6.02	5.69	5.44	5.24	5.08	4.94	4.82	4.72	4.64	4.56	4.50	4.44	4.38	4.33	4.29	4.21	4.15	4.09	4.05	4.01	3.92
22	14.38	9.61	7.80	6.81	6.19	5.76	5.44	5.19	4.99	4.83	4.70	4.58	4.49	4.40	4.33	4.26	4.20	4.15	4.10	4.06	3.98	3.92	3.86	3.82	3.78	3.69
24	14.03	9.34	7.55	6.59	5.98	5.55	5.24	4.99	4.80	4.64	4.51	4.39	4.30	4.21	4.14	4.07	4.02	3.96	3.92	3.87	3.80	3.74	3.68	3.63	3.59	3.51
26	13.74	9.12	7.36	6.41	5.80	5.38	5.07	4.83	4.64	4.48	4.35	4.24	4.14	4.06	3.99	3.92	3.86	3.81	3.77	3.72	3.65	3.59	3.53	3.49	3.45	3.36
28	13.50	8.93	7.19	6.25	5.66	5.24	4.93	4.70	4.51	4.35	4.22	4.11	4.01	3.93	3.86	3.80	3.74	3.69	3.64	3.60	3.52	3.46	3.41	3.36	3.32	3.24
30	13.29	8.77	7.05	6.13	5.53	5.12	4.82	4.58	4.39	4.24	4.11	4.00	3.91	3.83	3.75	3.69	3.63	3.58	3.54	3.49	3.42	3.36	3.30	3.26	3.22	3.14
35	12.90	8.47	6.79	5.88	5.30	4.89	4.60	4.36	4.18	4.03	3.90	3.79	3.70	3.62	3.55	3.48	3.43	3.38	3.33	3.29	3.22	3.16	3.10	3.06	3.02	2.93
40	12.61	8.25	6.60	5.70	5.13	4.73	4.44	4.21	4.02	3.87	3.75	3.64	3.55	3.47	3.40	3.34	3.28	3.23	3.19	3.15	3.07	3.01	2.96	2.91	2.87	2.79
45	12.39	8.09	6.45	5.56	5.00	4.61	4.32	4.09	3.91	3.76	3.64	3.53	3.44	3.36	3.29	3.23	3.17	3.12	3.08	3.04	2.96	2.90	2.85	2.80	2.76	2.68
50	12.22	7.96	6.34	5.46	4.90	4.51	4.22	4.00	3.82	3.67	3.55	3.44	3.35	3.27	3.20	3.14	3.09	3.04	2.99	2.95	2.88	2.82	2.77	2.72	2.68	2.60
60	11.97	7.77	6.17	5.31	4.76	4.37	4.09	3.87	3.69	3.54	3.42	3.32	3.23	3.15	3.08	3.02	2.96	2.91	2.87	2.83	2.76	2.69	2.64	2.60	2.56	2.47
70	11.80	7.64	6.06	5.20	4.66	4.28	3.99	3.77	3.60	3.45	3.33	3.23	3.14	3.06	2.99	2.93	2.88	2.83	2.78	2.74	2.67	2.61	2.56	2.51	2.47	2.39
80	11.67	7.54	5.97	5.12	4.58	4.20	3.92	3.71	3.53	3.39	3.27	3.16	3.07	3.00	2.93	2.87	2.81	2.76	2.72	2.68	2.61	2.55	2.49	2.45	2.41	2.32
100	11.50	7.41	5.86	5.02	4.48	4.11	3.83	3.61	3.44	3.30	3.18	3.07	2.99	2.91	2.84	2.78	2.73	2.68	2.63	2.59	2.52	2.46	2.41	2.36	2.32	2.24
200	11.16	7.15	5.63	4.81	4.29	3.92	3.65	3.43	3.26	3.12	3.01	2.90	2.82	2.74	2.67	2.61	2.56	2.51	2.47	2.42	2.35	2.29	2.24	2.19	2.15	2.07
500	10.96	7.00	5.51	4.69	4.18	3.81	3.54	3.33	3.16	3.02	2.91	2.81	2.72	2.64	2.58	2.52	2.46	2.41	2.37	2.33	2.26	2.20	2.14	2.10	2.05	1.97
1000	10.89	6.96	5.46	4.66	4.14	3.78	3.51	3.30	3.13	2.99	2.87	2.77	2.69	2.61	2.54	2.48	2.43	2.38	2.34	2.30	2.23	2.16	2.11	2.06	2.02	1.94
>1000	1.04	6.92	5.43	4.62	4.11	3.75	3.48	3.27	3.10	2.96	2.85	2.75	2.66	2.58	2.52	2.46	2.40	2.35	2.31	2.27	2.20	2.14	2.08	2.04	1.99	1.91
df2\df1	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	22	24	26	28	30	35